

UNIVERSIDADE FEDERAL DE SANTA CATARINA CENTRO DE CIÊNCIAS BIOLÓGICAS PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA

Fabiano Faga-Pacheco

Projeções futuras da distribuição de *Sargassum* (Fucales, Phaeophyceae) no Oceano Atlântico Ocidental em diferentes cenários de mudanças climáticas

Florianópolis 2023 Fabiano Faga-Pacheco

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Orientador(a): Prof. Dr. Carlos Frederico Deluqui Gurgel

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Projeções futuras da distribuição de *Sargassum* (Fucales, Phaeophyceae) no Oceano Atlântico Ocidental em diferentes cenários de mudanças climáticas

O presente trabalho em nível de Mestrado foi avaliado e aprovado, em 24 de outubro de 2022, pela banca examinadora composta pelos seguintes membros:

Prof. Carlos Frederico Deluqui Gurgel, Dr. Universidade Federal do Rio de Janeiro

Profa. Fernanda Thiesen Brum, Dra. Universidade Federal do Paraná

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Certificamos que esta é a versão original e final do trabalho de conclusão que foi julgado adequado para obtenção do título de Mestre em Ecologia.

> Profa. Natalia Hanazaki, Dra. Coordenação do Programa de Pós-Graduação

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Florianópolis, 2023



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UNIVERSIDADE FEDERAL DE SANTA CATARINA PRÓ-REITORIA DE PÓS-GRADUAÇÃO PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA

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Ata de sessão pública de apresentação e defesa de Dissertação de Mestrado de Fabiano Faga Pacheco.

Aos vinte e quatro dias do mês de outubro de dois mil e vinte e dois, às 14h00min, por videoconferência, foi realizada a sessão pública de apresentação e defesa da Dissertação de Mestrado, na Área de Concentração Ecologia, do mestrando Fabiano Faga-Pacheco como requisito final para obtenção do título de MESTRE em ECOLOGIA, de acordo com a Portaria nº 024/2022/PPGECO. A banca foi composta pelos seguintes professores: Dr. Carlos Frederico Deluqui Gurgel - Instituto de Biodiversidade e Sustentabilidade/UFRJ (Orientador), Dr^a. Fernanda Thiesen Brum – UFPR (Membro Titular – Externo à UFSC) e Dr. Eduardo Luís Hettwer Giehl - ECZ/CFM/UFSC (Membro Titular - Interno à UFSC), sob a presidência do primeiro. A dissertação tem como título Projeções futuras da distribuição de Sargassum (Fucales, Phaeophyceae) no Oceano Atlântico Ocidental em diferentes cenários de mudanças climáticas.

Houve participação por meio de sistemas de interação áudio e vídeo em tempo real pelos seguintes professores: Dr. Carlos Frederico Deluqui Gurgel, Dra. Fernanda Thiesen Brum e Dr. Eduardo Luís Hettwer Giehl.

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Às 17 horas e 40 minutos foi lavrada a presente ata e encerrada a sessão, que vai assinada pelo candidato e pela banca examinadora.

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RESUMO

Mudanças climáticas estão alterando a biota da Terra. Alterações na fenologia, morfologia, diversidade genética e distribuição das espécies marinhas têm sido reportadas em escala regional e global. Uma das mais profundas mudanças no ambiente marinho refere-se à tropicalização de comunidades bentônicas temperadas, com estimativas de alterações futuras na distribuição de espécies devido às mudanças no clima. Entretanto, pouco ainda é conhecido sobre os impactos do aquecimento global em comunidades de macroalgas marinhas tropicais. Espécies bentônicas formadoras de dossel, como as algas pardas conhecidas como sargaços (gênero Sargassum), têm um papel importante em substratos consolidados, providenciando complexidade ambiental e abrigo para uma variedade de organismos. Apesar da grande importância pelos serviços ecossistêmicos proporcionados por estas algas, evidências de perdas significativas de hábitat e biomassa têm sido reportadas nos últimos anos. Para compreender como mudanças futuras nos preditores ambientais podem afetar a distribuição de espécies de sargaço ao longo do oceano Atlântico Ocidental, foram aplicados Modelos de Nicho Ecológico (MNE) para 12 espécies amplamente conhecidas. Um outro MNE foi calculado para uma espécie aqui chamada de Sargassum WA, que representa seis espécies morfologicamente definidas que se encontram em uma politomia em árvores filogenéticas recentes. Projeções de adequabilidade ambiental foram feitas para o presente e para os cenários, um mais realista ("Representative Concentration Pathway" - RCP 4.5) e um mais pessimista (RCP 8.5) do IPCC. Os resultados indicam mudanças na adequabilidade ambiental latitudinal para as espécies, variando de 0.5º a 8.1º em direção ao norte e entre 0º e 5.5º em direção ao sul. Seis espécies de Sargassum devem ter reduzidas a sua área adequada entre 10% e 80%, enquanto outras seis espécies devem ter sua área adequada aumentada entre 4% e 168%. De maneira geral, as alterações previstas apresentam um padrão predominante de aumento da adequabilidade ambiental em maiores latitudes ao longo do oceano Atlântico Ocidental para todas as espécies, bem como uma diminuição da área de adequabilidade em latitudes menores para metade das espécies. Esse padrão é consistente com uma esperada tropicalização de ambientes em latitudes temperadas. Essas mudanças podem produzir perdas consideráveis em serviços ecossistêmicos mantidos por bancos de Sargassum, em

particular em menores latitudes. Os resultados realçam a necessidade de políticas de conservação e estratégicas de manejo de espécies bentônicas de *Sargassum*, a fim de evitar efeitos negativos de mudanças na adequabilidade dos hábitats.

Palavras-chave: ambiente marinho bentônico; aquecimento global; ecologia de algas; espécies bentônicas de *Sargassum;* modelagem de nicho ecológico; modelo de distribuição de espécies; mudanças climáticas; Oceano Atlântico Ocidental.

ABSTRACT

Climate changes have impacted phenology, morphology, genetic diversity and distribution of marine species at regional and global scales. One of the more pervasive changes in the marine environment refers to the tropicalization of temperate benthic communities. Recent climate warming has changed the global redistribution of marine species, whether by poleward pattern of expansions or regional contractions at lower latitude. However, the impacts of future global warming on tropical marine macroalgal communities have been underrepresented. Sargassum species are foundation species that play an important role for marine ecosystems by providing complexity and shelter for a wide range of other organisms. To understand how future changes in abiotic variables could affect the distribution of Sargassum species along the Western Atlantic Ocean, we applied Ecological Niche Models (ENM) for 12 widely recognized species. Another ENM, dubbed Sargassum WA, was calculated and represents 6 morphologically defined species that area likely to be lumped following recent phylogenetic data. We produced models from the present to contrasting representative concentration pathways scenarios (RCP 4.5 and 8.5) of future climate changes. Our results showed changes in niche suitability between 0.5° and 8.1° northward, and 0° and 5.5° southward. Six species are likely to reduce their suitability area from 10% to 80%, while other six species are likely to expand their suitability area from 4% to 168%. Overall, suitability area showed an increase at higher latitudes for most species but decrease in lower latitude for half of the species were observed. This pattern is consistent with the expected tropicalization of temperate latitudes following global warming. Such changes can produce considerable losses in ecosystem services provided by Sargassum beds, particularly in edge populations. Our findings highlight the need of Sargassum conservation policies and management strategies to avoid the negative effects of shifts in benthic Sargassum habitat suitability.

Keywords: algal ecology; benthic marine environment; ecological niche modelling; future climate change; global warming; *Sargassum;* species distribution model; Western Atlantic Ocean.

LISTA DE FIGURAS

Figure 1. Relation between maximum latitude with suitability area in the present		
scenario for each Western Atlantic Sargassum species and the latitudinal variation		
predicted in our model results81		
Figure 2. Habitat suitability model projections for 12 Sargassum species82		
Figure 3. Changes of future suitability area for Western Atlantic Sargassum species.		
Figure S1. Phylogenetic relationships of selected Sargassum species included in this		
study, following Camacho et al. (2015) and B. Barros-Barreto, M. T. Szechy and V.		
Cassano (unpublished data)99		
Figure S2. Variables histogram and their correlation used to fit ecological niche		
models for Sargassum species in the Western Atlantic Ocean		
Figure S3. Correlation between changes in suitability area and changes in latitudinal		
limits for 13 Sargassum species from the Western Atlantic Ocean		
Figure S4. Suitability map for Sargassum WA predictions for (A) present, (B) RCP		
4.5, and (C) RCP 8.5 scenarios102		

LISTA DE TABELAS

Table 1. Number of records used to produce niche ecological model for Sargassum species, values for mean test Area Under Receiver Operating Characteristic Curve (ROC-AUC), true skill statistics (TSS), fixed cumulative value 10 complementary loglog transformation threshold (T10), and the percentual relative contribution of the parameters temperature, salinity, depth, and ice to the model. We used TSS for those species with more than 35 records, for whom we used an ensemble approach. T10 was used for the species with few registers modeled with leave-one-out-crossvalidation approach in Maxent only. The more redder a cell, the greater the variable Table 2. Suitability area, suitability area variation (Δ Area), maximum North latitude (Max Lat North), variations northward (Δ North), maximum South latitude (Max Lat South), variation southward (Δ South), and overlap area between suitability occurrence in present and future scenarios for 13 benthic Sargassum species from the Western Atlantic Ocean. The percentage of the area and overlap area has the present projected scenario as reference. Latitudinal parameters are in degrees.78 Table S1. Values of Area Under Receiver Operating Characteristic Curve (AUC) and true skill statistics (TSS) for each algorithm tested for seven Sargassum species. Models tested: generalized linear model (GLM), generalized additive model (GAM), boosted regression trees (BRT), random forests (RF), Bioclim, classification and regression trees (CART), support vector machine (SVM), artificial neural networks (ANN), maximum likelihood (MAXLIKE), maximum entropy (MAXENT), and

SUMÁRIO

INTRODUÇÃO	16
ESPÉCIES SELECIONADAS	18
Sargassum bermudense	18
Sargassum buxifolium	18
Sargassum cymosum	19
Sargassum filipendula	20
Sargassum furcatum	21
Sargassum hystrix	21
Sargassum platycarpum	22
Sargassum polyceratium	22
Sargassum pteropleuron	23
Sargassum ramifolium	23
Sargassum rigidulum	23
Sargassum stenophyllum	24
Sargassum WA	25
OBJETIVOS	27
CAPÍTULO ÚNICO	28
DISTRIBUTIONAL RANGE SHIFTS OF BENTHIC SARGASSUM	SPECIES
(PHAEOPHYCEAE: FUCALES) UNDER FUTURE CLIMATE CHANGES	IN THE
WESTERN ATLANTIC OCEAN	29
Abstract	30
Introduction	32
Materials and Methods	35
Delimited area	35
Sargassum species	35
Occurrence data	
Environmental abiotic variables	
Prediction scenarios	38
Ecological niche modelling	
Species with small sample sizes	40
Analytical notes	41
Results	42

Changes in latitudinal limits	
Changes in suitability area	43
Changes in overlap area	43
Assessment metrics	
Predictors contributions	
Discussion	45
Latitudinal patterns	45
Suitability Area	46
Overlap	
Research Limitations	
Management and Conservation Strategies	51
Conclusions	52
Acknowledgements	53
References	54
List of Tables (Legends)	76
List of Figures (Legends)	80
Supplementary Material	96
Appendix S1 - Considerations of some unchosen model variab	les 103
Appendix S2 – Comments on research limitations	
Appendix S3 - R scripts used in this study	
CONCLUSÃO	
REFERÊNCIAS	

INTRODUÇÃO

Mudanças climáticas devem alterar a distribuição, fenologia e até mesmo a expressão do fenótipo de diversas espécies (Barry et al. 1995, Parmesan 2006, Thompson et al. 2002, Wernberg et al. 2012, Wiens 2016, Yan et al. 2017). Um dos processos associados com as mudanças climáticas nos ambientes marinhos é o aquecimento global (Levitus et al. 2000, Helmuth et al. 2002, Muller et al. 2009). O aquecimento das águas promove a tropicalização de comunidades bentônicas em ambientes marinhos temperados, representada tanto pelo desaparecimento de espécies de águas mais frias de ecossistemas temperados (i.e., florestas de kelps; Wernberg et al. 2010, 2016, Vergés et al. 2014, Franco et al. 2018, Jonsson et al. 2018) quanto pela sua substituição por espécies de águas mais quentes, como corais e peixes tropicais (Lima et al. 2007, Harley et al. 2012, Vergés et al. 2014). Evidência deste fenômeno tem sido reportada em sistemas marinhos temperados de locais como Austrália oriental (Vergés et al. 2016), Austrália Ocidental (Zarco-Perello et al. 2017, 2021), Atlântico Ocidental (Filbee-Dexter et al. 2016), Canal da Mancha (Hawkins et al. 2003, Southward et al. 1995), Península Ibérica (Casado-Amezúa et al. 2019) e Japão (Tanaka et al. 2012). Entretanto, os efeitos do aquecimento global sobre ambientes marinhos tropicais permanecem pouco conhecidos (Kleyplas 2019).

O gênero Sargassum é o gênero de algas pardas mais conspícuo nos bentos do infralitoral em latitudes tropicais (Tsuda 1972, Wanders 1976, Connor & Adey 1977, Morrissey 1980). Sargassum é responsável pela formação de várias agregações de macroalgas (chamadas de florestas ou bancos de sargaço ou *"Sargassum* beds") em ambientes costeiros tropicais e subtropicais (Guimaraens *et al.* 1994, Mafra Junior 2001, Pereira *et al.* 2006). Espécies de Sargassum atuam como espécies fundadoras, pioneiras, propiciando heterogeneidade ambiental, refúgio para diversos animais vertebrados e invertebrados bênticos e servindo de substrato para organismos epizoóticos e epifíticos (Earle 1969, Paula & Oliveira-Filho 1980, Farnham *et al.* 1981, Brayner-Barros *et al.* 2006, Raut *et al.* 2018). A heterogeneidade ambiental em substratos consolidados é ampliada pela grande complexidade estrutural tridimensional do talo de certas algas pardas, inclusive Sargassum (Airoldi *et al.* 2014).

Sargassum é um gênero multidiverso compreendendo 537 espécies descritas no mundo todo, das quais 356 são consideradas válidas (Guiry & Guiry Sargassum apresenta uma variação morfológica interespecífica e 2020). intraespecífica muito grande (Cordero 1981, Moreira & Suárez 2002, Schell et al. 2015, Dibner et al. 2021), levando à descrição de muitas espécies baseadas na morfologia (Mattio & Payri 2011). Entretanto, estudos de sistemática filogenética recentes mostraram que a diversidade atual de Sargassum parece ser ainda menor do que previamente considerada (Phillips & Fredericg 2000, Mattio et al. 2009, Yip et al. 2020). Por exemplo, revisões da flora de Sargassum do oceano Índico Ocidental (Ilhas Maurício e Reunião) reconheceu como válidos apenas 7 espécies entre as 44 anteriormente reportadas (Mattio et al. 2013). Outro estudo reconheceu apenas 6 morfotipos dos 41 anteriormente registrados para Cingapura (Yip et al. 2018). Revisões também encontraram uma menor diversidade real de Sargassum do que os registros históricos na Polinésia Francesa, Nova Caledônia e África do Sul, principalmente devido a identificações equivocadas e falta de delimitação clara interespecífica (Mattio et al. 2008, 2015, Mattio & Payri 2009).

No oceano Atlântico Ocidental, 24 espécies de *Sargassum* foram reportadas, sendo 22 espécies bênticas e 2 pelágicas, ocorrendo do Canadá ao sul do Brasil (Taylor 1969, Ugadim 1973, Richardson 1975, Paula 1988, Camacho *et al.* 2015, Wynne 2017, González-Nieto *et al.* 2020). Apesar de alguns esforços para compreender as dinâmicas das espécies pelágicas de *Sargassum*, bem como entender os fatos que influenciam a distribuição das marés marrom (por vezes chamadas de "marés douradas", Lapointe 1986, 1995, Brooks *et al.* 2018, Oviatt *et al.* 2019, Wang et al. 2019, Godínez-Ortega *et al.* 2021, Jouanno *et al.* 2021), nenhuma predição está disponível para descrever como as mudanças climáticas podem alterar a adequabilidade ambiental das espécies bênticas de *Sargassum* e, portanto, de como isso pode afetar a distribuição futura das espécies (mas veja Gouvêa *et al.* 2020).

Espécies de Sargassum são sensíveis a impactos locais (como poluição), regionais (como desenvolvimento urbano) e globais (Dawes & Tomasko 1988, Steen 2004, Engelen *et al.* 2005, Miki *et al.* 2016, Abe *et al.* 2020). Sargassum pode também atuar como espécies bioindicadoras de efluentes industriais enriquecidos com nutrientes e de poluição por metais pesados, sendo capazes de sequestrar e

acumular metais como cádmio, zinco e arsênio (Alquezar *et al.* 2013, Guitouni *et al.* 2016, Ho & Bantoto-Kinamot 2021). A presença de bancos de *Sargassum* saudáveis em determinados recifes é um indicador da saúde de toda a comunidade bêntica (Vroom & Braun 2010, Eggertsen *et al.* 2017). Portanto, entender como as mudanças climáticas afetarão a adequabilidade ambiental de *Sargassum* é uma etapa crucial para dar suporte a políticas de manejo, não apenas para preservar e proteger bancos de macroalgas, mas também para a conservação de comunidades bentônicas marinhas como um todo (Filbee-Dexter 2020).

ESPÉCIES SELECIONADAS

Sargassum bermudense

Espécie descrita a partir de exemplares de Bermuda e Cuba (Grunow 1916:147). Ocorre apenas nesses locais e nos Estados Unidos (Flórida e um registro em Massachusetts), alguns poucos locais do Mar do Caribe (Antígua e Barbuda, Bahamas, Porto Rico) e na Venezuela (Suárez 2005, Suárez *et al.* 2015, CRIA 2018, GBIF 2020, Guiry & Guiry 2020).

Dentre a lista de espécies com as quais *S. bermudense* já foi confundida, encontra-se *S. lendigerum*, *S. vulgare*, *S. cymosum* e *S. acinarium* (Schneider 2003). A variedade *S. bermudense* var. *stagnale*, cujos ramos podem chegar a 2m, foi encontrada somente em Walsingham Pond, nas Bermudas (Schneider 2003).

Sargassum buxifolium

Espécie descrita como variedade de *S. hystrix* por Chauvin em J. Agardh (1848:322). Foi elevado a espécie apenas recentemente (Wynne, 2011), baseado em diferenciações morfológicas em águas cubanas (Moreira & Cabrera 2007). A ocorrência da espécie está restrita à costa atlântica dos Estados Unidos (Massachusetts a Flórida), México, Belize e Caribe, chegando a águas sul-americanas na Colômbia e Venezuela (Earle 1969, GBIF 2020, Guiry & Guiry 2020). No Brasil, foi registrada em Fernando de Noronha e Abrolhos, no que parece ser o limite austral conhecido de sua distribuição (Figueiredo 2006, Oliveira *et al.* 2009).

Sargassum cymosum

O holótipo da espécie vem de águas brasileiras (C. Agardh, 1821). É a única espécie daqui selecionada que apresenta distribuição natural conhecida além do Atlântico. No Atlântico Ocidental, a espécie foi registrada da Carolina do Norte ao Brasil (Paula 1988, Paula & Oliveira 1982, Leite *et al.* 2007, Rover 2014, Camacho *et al.* 2015, Guiry & Guiry 2020,). A espécie também foi mencionada no Atlântico Central, nas Canárias, Açores, Cabo Verde e Ilhas Savage (Gallardo *et al.* 2016, Guiry & Guiry 2020).

A ocorrência da espécie no Atlântico Oriental está sujeita a maiores investigações taxonômicas. C. Agardh (1821, p.20) citou a espécie para os mares de Serra Leoa, como uma forma de *S. cymosum*, a saber forma β *Latifolium*. Com base nessa descrição, a ocorrência de *S. cymosum* na costa oeste africana foi ampliada para Gâmbia e Gabão e replicada em listas de macroalgas locais (Price *et al.* 1978). O material analisado por C. Agardh para Serra Leoa foi reconhecido como *Sargassum cheirifolium* por Kützing (1849:613). De Toni (1895) colocou o material do Gabão como sinônimo de *Sargassum vulgare*, inclusive *S. cheirifolium* e *S. cymosum* f. *Latifolium*. Atualmente, entretanto, *S. vulgare* está com status taxonômico não resolvido (veja Prud'homme van Reine 2011). Já *S. cheirifolium* está considerado hoje como *S. cymosum* var. *esperi* (Guiry & Guiry 2020).

Na África do Sul e em Madagascar, ocorre a variedade *S. cymosum* var. *scabriusculum* (Mattio *et al.* 2015, Vieira *et al.* 2021). Em revisão das espécies de *Sargassum* para a costa sul-africana, Mattio *et al.* (2015) encontrou que as citações na literatura para *S. cymosum* var. *gueinzii* e *S. cymosum* var. *lendigerum* tratam-se, na verdade, de *Sargassum* elegans. Já as citações de *S. cymosum* f. *dichocarpa* referiam-se a *S. cymosum* var. *scabriusculum*.

Menções a ilhas Galápagos pela Charles Darwin Foundation não constam mais de sua página na internet. Menção a ocorrência na Índia, China e Vietnã vêm de catálogos, sem consulta ao material de herbário para atualização taxonômica (Sahoo *et al.* 2001, Nguyen *et al.* 2013, Phang *et al.* 2016). A presença da espécie nas Ilhas Andaman ainda é incerta. A citação proveio de um catálogo (Silva *et al.* 1996) e a espécie não foi reencontrada em estudos posteriores (Karthick

et al. 2013). A ocorrência de *S. cymosum* nesses locais parece advir de equívocos de identificação e necessitam de mais estudos taxonômicos. Por fim, a forma *S. cymosum* f. *borbonica* é considerada endêmica da Ilha Reunião (Mattio *et al.* 2013).

Como a taxonomia da espécie está relativamente bem resolvida para o Atlântico Ocidental, optou-se por usar apenas os dados constantes das Américas para a geração dos modelos.

Sargassum filipendula

A espécie foi descrita para águas possivelmente do México (C. Agardh, 1824:300) e foi mencionada para o Atlântico Central (Canárias, Madeira e Ilhas Savage) e Ocidental (Guiry & Guiry 2020). Neste último, foi listado de Massachusetts até o Brasil, com limite sul em Santa Catarina, mais especificamente em Florianópolis, nas praias do Sambaqui e Canasvieiras (Fialho 2015, GBIF 2020, Guiry & Guiry 2020).

Menções para outros locais parecem advir de equívocos de identificação. Listas para o Atlântico Oriental mencionam Gabão, Gana e Mauritânia (Price *et al.* 1978:148, Marcot-Coqueugniot 1991, John *et al.* 2014:46). Em sua revisão de *Sargassum* para a África do Sul, Mattio *et al.* (2015) afirmou que nenhuma espécie similar foi coletada nesse país, sugerindo equívoco na identificação de exemplar do Cabo da Boa Esperança (GBIF 2020).

Menções a Sri Lanka, Mar da China, Indonésia e Malásia provém de catálogos (Phillips 1995, Silva *et al.* 1996, Atmadja & Prud'homme van Reine 2014, Phang *et al.* 2016) e exemplares de Tailândia, Seychelles e Índia (GBIF 2020) também parecem advir de equívocos prévios de identificação, situação similar à identificação fenotípica de exemplares da Arábia Saudita (Abdel-Kareem 2009, John & Al-Thani, 2014:366). Uma melhor identificação unindo taxonomia comparada e filogenética pode elucidar melhor a distribuição da espécie fora do Atlântico Ocidental. Entretanto, a presença dela em sua área de origem e os padrões que se desenham até agora que aliam processos evolutivos e oceanográficos nos asseguram uma robusta segurança nas áreas de ocorrência da espécie nas águas americanas.

Sargassum furcatum

Espécie descrita por Kützing (1843:362) para as Ilhas Virgens Americanas, no Caribe. A espécie foi encontrada em alguns outros países insulares, como Cuba, Antilhas Holandesas e Trinidad e Tobago, além da Venezuela e Brasil (Zayas *et al.* 2002, Solé & Pardo 2005, Menezes-Széchy & Paula 2010, Guiry & Guiry 2020). Segundo Robinson *et al.* (2012), a espécie parece estar expandindo a sua distribuição, tendo sido encontrada recentemente no México e na Costa Rica (Dreckmann 1996, Cabrera *et al.* 2019). No Brasil, foi listada apenas para Bahia, Rio de Janeiro e São Paulo (Széchy & Paula 2000, Oliveira *et al.* 2002), em localidades específicas como Baía de Todos os Santos (Marins *et al.* 2008, Nunes & Paula 2002), Região dos Lagos (Ornellas & Coutinho 1998, Pereira & Yoneshigue-Valentin 1999, Rocha 2003) e litoral norte de São Paulo, com limite austral em São Sebastião (Paula 1988, Siqueira 2012).

A espécie foi ainda encontrada no Atlântico Central, nas Canárias e Ilhas Savage (Haroun *et al.* 2002, Parente *et al.* 2000) e nas Ilhas Chafarinas, nos Marrocos (Flores-Moya & Conde 1998). A menção às Filipinas provém de catálogo (Silva *et al.* 1987) e provavelmente trata-se de erro de identificação.

A variedade *S. furcatum* var. *diversifolium* foi originalmente descrita como uma forma de *S. vulgare* por Grunow (1916:41) para as Ilhas Canárias, Açores e Madeira.

Sargassum hystrix

Espécie descrita para os bancos do Campeche, no México (J. Agardh 1847:7). Ocorre da Virgínia, Estados Unidos, até São Paulo, Brasil, incluindo Bermuda, América Central, Caribe, Colômbia e Venezuela (Camacho *et al.* 2015, GBIF 2020, Guiry & Guiry 2020). *S. hystrix* já foi encontrada a 137m de profundidade (Littler & Littler 2000)

As ocorrências mencionadas em catálogo para Senegal e Serra Leoa (John *et al.* 2004) foram consideradas duvidosas por Lawson & John (1987). A ocorrência na Indonésia provém de catálogo (Atmadja & Prud'homme van Reine 2014). Menção para o Mar Vermelho (Einav *at el.* 2021) advém de um trabalho sobre ecologia

bacteriana em macroalgas do Egito e não de taxonomia (Salem *et al.* 2011). Essas ocorrências, juntamente com a identificação de material de Seychelles como *S. hystrix* var. *spinulosum*, provavelmente são equívocos de identificação.

Sargassum platycarpum

Espécie descrita originalmente para a Martinica (Montagne 1848:248), ocorre em latitudes mais próximas do Equador, sendo listada do México, Caribe e Bermuda até o Brasil, onde foi encontrada na Laje de Santos e Ilhas de Alcatrazes e Queimada Grande (Rocha-Jorge 2015, GBIF 2020, Guiry & Guiry 2020).

No Atlântico Central, foi encontrado na Ilha da Madeira (Ferreira *et al.* 2018) e está presente em catálogos para Canárias e Cabo Verde (Price *et al.* 1978, John et al. 2004). A despeito da fragilidade e potencialidades apresentadas por essas últimas publicações, há registros da espécie em Abrolhos e no Recife Sebastião Gomes, mais próximos à costa (Torrano-Silva & Oliveira 2013). Já a menção da espécie no Golfo Arábico (Abdel-Kareem 2009) parece advir de equívoco de identificação.

Sargassum polyceratium

Espécie descrita para Cuba (Montagne 1837:356) com ocorrência a partir da Carolina do Norte, passando por Flórida e Bermuda, pegando o Golfo do México, Caribe, Colômbia, Venezuela e Brasil, onde ocorre até São Paulo (Venekey *et al.* 2008, Camacho *et al.* 2015, GBIF 2020, Guiry & Guiry 2020).

Menções ao Mar da China, Turquia e Indonésia vem de catálogos (Taskin *et al.* 2008, Atmadja & Prud'homme van Reine 2014, Phang *et al.* 2016) e na Filipinas foram citados em dois locais (Silva *et al.* 1987). Não há exemplares desses países nos catálogos de herbários verificados para este trabalho e a ocorrência da espécie nesses locais sem a ajuda humana é improvável.

Espécie típica de águas quentes, descrita para as Bahamas (Grunow 1868:55) e com ocorrência restrita aos Estados Unidos (Carolina do Norte, Flórida, Louisiana, Texas), Bermuda, América Central (Belize, Panamá), Caribe e Venezuela (Earle 1969, Prince & O'Neal 1979, Dawes *et al.* 1988, Dawes 1989, Camacho *et al.* 2015, GBIF 2020, Guiry & Guiry 2020).

Menções a Visayas Central, Filipinas (Silva *et al.* 1987), provavelmente provêm de equívocos de identificação.

Sargassum ramifolium

Espécie descrita para águas brasileiras (Kützing 1843:362, 1861:10) com distribuição basicamente do Golfo do México e Caribe ao Brasil (GBIF 2020, Guiry & Guiry 2020). Ocorre da Flórida e Bermudas até Bombinhas, Santa Catarina (Littler & Littler 1997, Rosado-Espinosa 2012, Semidey *et al.* 2013, Silva 2010, Carneiro 2017, Ballantine *et al.* 2016, Menezes-Széchy & Paula 2010).

Menções à presença da espécie na Mauritânia, Senegal e Guiné-Bissau na África e às Ilhas Canárias parecem ser verdadeiras (Welten *et al.* 2002, John *et al.* 2003, 2004).

Assim como todas as espécies, apenas utilizamos para a criação dos nossos modelos, os dados provenientes da costa ocidental do Atlântico.

Sargassum rigidulum

Espécie descrita para o Brasil a partir de exemplares de Pernambuco e Rio de Janeiro (Kützing 1849:615). Ocorre dos Estados Unidos (Flórida) e México até Santa Catarina, Brasil (Vroman 1968, Carela 1977, Oliveira Filho 1977, Moreira & Cabrera 2007, Bernecker 2009, Menezes-Széchy & Paula 2010, GBIF 2020, Guiry & Guiry 2020).

O aspecto geral da espécie assemelha-se a *S. cymosum*, com quem tem sido, por vezes, confundida (Oliveira Filho 1977). Alguns registros de *S. cymosum* foram baseados em exemplares de *S. rigidulum*, como ocorreu na Colômbia (Diaz-

Pulido & Díaz-Ruíz 2003), Venezuela (Cordero 2016), Barbados (Wynne *et al.* 2014) e, aparentemente, em Cabo Verde (Gabriel & Fredericq 2019). Camacho et al. 2015, por sua vez, coloca a espécie como sinônimo-júnior de *S. cymosum*, algo ainda não corroborado por Guiry & Guiry (2020).

A citação da espécie para o Japão (Yoshida *et al.* 2015) provavelmente se trata de um equívoco de identificação.

Sargassum stenophyllum

Martius é listado no AlgaeBase (Guiry & Guiry 2020) como o descritor da espécie, em livro escrito entre 1828 e 1834 de sua viagem pelo Brasil entre 1817 e 1820 (Martius 1834:8-9). Entretanto, nesse mesmo volume, o autor indica como referência o volume 1 (*"Algae, Lichenes, Hepaticae"*) de sua *"Florae Brasiliensis"* (Martius, 1833:47). De fato, o primeiro livro contém uma descrição extremamente simples da espécie, porém com ilustração, enquanto a segunda obra contém uma descrição muito mais detalhada e precisa da espécie. Em ambos os casos, também foi mencionada a citação prévia de Mertens para *Fucus stenophyllus*, referente à espécie, embora sem explicitar a literatura de referência.

A espécie foi descrita a partir de exemplares do Rio de Janeiro (*"Provinciarum Sebastianopolitanae"*), Bahia e São Paulo. O holótipo mencionado em Silva *et al.* (1996) como sendo de São Sebastião não leva em conta o antigo nome da então Província de São Sebastião do Rio de Janeiro.

A espécie está presente em águas da Colômbia, Venezuela e Brasil, tendo sido encontrada no Ceará (Caucaia) e da Bahia até o Rio Grande do Sul, onde foi encontrada até a Lagoa do Imbé (Eston & Bussab 1990, Haddad & Chiaverini 2000, Tanaka 2004, Scherner *et al.* 2012, Almeida 2014, Camacho *et al.* 2015, Oliveira *et al.* 2015, Machado *et al.* 2015, CRIA 2018, Razzera 2018, GBIF 2020).

Grunow (1916:138) considerou a espécie uma variedade de *S. cymosum*, citando-a para Açores, Senegal e Tenerife (nas Ilhas Canárias), locais sem registros recentes nem exemplares da espécie. A citação da espécie para a Nova Caledônia (Phillips 1995) proveio de exemplares que seriam de *S. aquifolium* (Mattio & Payri 2009). Já a ocorrência na Austrália e Nova Zelândia veio de um catálogo em que não consta nenhum voucher (Bostock & Holland 2010) e cuja identificação baseou-

se em *S. stenophyllum* J. Agardh, considerado "*nomen illegitimun*" - a mesma identificação usada para os exemplares da Nova Caledônia, devendo, portanto, tratar-se também de exemplares de *S. aquifolium*. *S. stenopphyllum* J. Agardh é considerado hoje sinônimo heterotípico de *S. novae-hollandiae* (Silva *et al.* 1996).

Sargassum WA

Trabalhos recentes e em andamento usando dados moleculares registraram uma diversidade genética muito pequena ou mesmo inexistente entre algumas espécies de *Sargassum* que ocorrem no Atlântico. As filogenias geradas mostraram uma politomia entre as espécies *S. cymosum, S. filipendula, S. furcatum, S. hystrix, S. polyceratium* and *S. stenophyllum*, sugerindo que elas podem ser conspecíficas. Elas formam um clado não resolvido dentro de árvores filogenéticas de máxima parcimônia dentro do subgênero *Sargassum* (Camacho *et al.* 2015; B. Barros-Barreto, M. T. Szechy & V. Cassano, unpub. data; Figura S1). Os registros dessas seis espécies foram unidos para modelá-las conjuntamente como uma única espécie filogenética, aqui chamada *Sargassum* WA.

Algumas das espécies aqui do estudo não constam ainda de análises filogenéticas, então sua posição evolutiva ainda não está clara com base nas melhores evidências hoje disponíveis. S. pteropleuron e S. platycarpum são espécies-irmãs que estão num clado diferente das espécies que compõem Sargassum WA. Já S. buxifolium foi originalmente descrito como uma variedade de S. hystrix (J. Agardh 1848:322) e apenas filogenias bastante recentes demonstraram sua relação muito próxima às espécies de Sargassum WA, apontando também para serem conspecíficas (Yip et al. 2020). A proximidade morfológica entre S. bermudense e S. cymosum também já foi registrada (Schneider 2003, Schneider & Flook, 2017) e novos dados moleculares incluem ambas espécies na mesma politomia de Sargassum WA (González-Nieto et al. 2020). Dados moleculares de S. rigidulum e S. ramifolium, bem como de várias outras espécies do Atlântico Ocidental, permanecem ausentes em filogenias. S. rigidulum já foi confundido com S. cymosum (Oliveira Filho 1977, Diaz-Pulido & Díaz-Ruíz 2003, Wynne et al. 2014, Cordero 2016, Gabriel & Fredericq 2019) e S. ramifolium já foi considerado uma variedade de S. cymosum (Grunow 1916:141, Paula 1988). Dessa forma, existe uma alta possibilidade de que essas duas

espécies venham a compor o complexo de espécies aqui considerado como Sargassum WA.

OBJETIVOS

O objetivo principal do estudo é testar se as mudanças climáticas globais, em especial o aquecimento dos oceanos, promoverá alterações na adequabilidade ambiental nas áreas de ocorrência de *Sargassum* bentônico no Atlântico ocidental em cenários futuros de aumento da concentração de dióxido de carbono atmosférico.

Mais especificamente, os objetivos deste trabalho são:

(1) Predizer as mudanças nas latitudes de adequabilidade ambiental de 13 espécies bentônicas de *Sargassum* (12 espécies morfológicas e 1 filogenética) do Atlântico ocidental nos cenários RCP 4.5 (cenário mais plausível) e RCP 8.5 (mais pessimista) do IPCC (2014);

(2) Estimar as alterações quantitativas e espaciais na área de adequabilidade das espécies selecionadas de Sargassum nos cenários RCP 4.5 e RCP 8.5.

Baseadas nessas premissas, elaboramos as seguintes hipóteses:

(1) Haverá um aumento da área de adequabilidade ambiental para as espécies em maiores latitudes nos cenários futuros em relação ao cenário presente;

(2) Haverá diminuição geral da área de adequabilidade ambiental das espécies de Sargassum, com redução da adequabilidade em locais mais próximos à Linha do Equador.

Adicionalmente, hipotetizamos que:

(3) Os efeitos de aumento da latitude e diminuição da área serão mais pronunciados no cenário RCP 8.5 do que no cenário RCP 4.5.

CAPÍTULO ÚNICO

Artigo formatado no padrão para submissão ao Journal of Phycology.

DISTRIBUTIONAL RANGE SHIFTS OF BENTHIC *SARGASSUM* SPECIES (PHAEOPHYCEAE: FUCALES) UNDER FUTURE CLIMATE CHANGES IN THE WESTERN ATLANTIC OCEAN

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Abstract

Climate changes have impacted phenology, morphology, genetic diversity and distribution of marine species at regional and global scales. One of the more pervasive changes in the marine environment refers to the tropicalization of temperate benthic communities. Recent climate warming has changed the global redistribution of marine species, whether by poleward pattern of expansions or regional contractions at lower latitude. However, the impacts of future global warming on tropical marine macroalgal communities have been underrepresented. Sargassum species are foundation species that play an important role for marine ecosystems by providing complexity and shelter for a wide range of other organisms. To understand how future changes in abiotic variables could affect the distribution of Sargassum species along the Western Atlantic Ocean, we applied Ecological Niche Models (ENM) for 12 widely recognized species. Another ENM, dubbed Sargassum WA, was calculated and represents 6 morphologically defined species that area likely to be lumped following recent phylogenetic data. We produced models from the present to contrasting representative concentration pathways scenarios (RCP 4.5 and 8.5) of future climate changes. Our results showed changes in niche suitability between 0.5° and 8.1° northward, and 0° and 5.5° southward. Six species are likely to reduce their suitability area from 10% to 80%, while other six species are likely to expand their suitability area from 4% to 168%. Overall, suitability area showed an increase at higher latitudes for most species but decrease in lower latitude for half of the species were observed. This pattern is consistent with the expected tropicalization of temperate latitudes following global warming. Such changes can produce considerable losses in ecosystem services provided by Sargassum beds, particularly in edge populations. Our findings highlight the need of Sargassum conservation policies and management strategies to avoid the negative effects of shifts in benthic Sargassum habitat suitability.

Keywords: algal ecology; benthic marine environment; ecological niche modelling; future climate change; global warming; *Sargassum;* species distribution model; Western Atlantic Ocean.

Introduction

32

Climate changes are expected to change the distribution, phenology and even the phenotypic expression of several species (Barry et al. 1995, Thompson et al. 2002, Parmesan 2006, Wernberg et al. 2012, Wiens 2016, Yan et al. 2017). One of the processes associated with climate change in the marine environment is global warming (Levitus et al. 2000, Helmuth et al. 2002, Muller et al. 2009). Warming waters promote the tropicalization of temperate marine benthic communities, which means the local disappearance of temperate species in temperate ecosystems (e.g. kelp forests; Wernberg et al. 2010, Vergés et al. 2014, Wernberg et al. 2016, Franco et al. 2018, Jonsson et al. 2018) and their replacement by tropical species (e.g., corals, tropical fishes, Lima et al. 2007, Harley et al. 2012, Vergés et al. 2014). Evidence of this phenomenon has already been reported in temperate marine systems such as Eastern Australia (Vergés et al. 2016), Western Australia (Zarco-Perello et al. 2017, 2021), Western Atlantic (Filbee-Dexter et al. 2016), English Channel (Hawkins et al. 2003, Southward et al. 1995), Iberian Peninsula (Casado-Amezúa et al. 2019) and Japan (Tanaka et al. 2012). However, the effects of global warming on tropical marine environments remain poorly known (Kleyplas 2019).

The genus *Sargassum* is the most conspicuous subtidal benthic marine brown alga in tropical latitudes (Tsuda 1972, Wanders 1976, Connor and Adey 1977, Morrissey 1980). *Sargassum* is responsible for the formation of several seaweed forests (or *Sargassum* beds) in tropical and subtropical subtidal coastal environments (Guimaraens et al. 1994, Mafra Junior 2001, Pereira et al. 2006). *Sargassum* spp. act as foundations species, providing environmental heterogeneity, refuge for a range of benthic invertebrate and vertebrate animals and acting themselves as substrate for several epizootic and epiphytic taxa (Earle 1969, Paula & Oliveira-Filho 1980, Farnham et al. 1981, Brayner-Barros et al. 2006, Raut et al. 2018).

Environmental heterogeneity along hard-bottom substrates is increased by the greater structural three-dimensional complexity presented by some brown algal thalli, including *Sargassum* (Airoldi et al. 2014).

Sargassum is a very diverse genus, comprising 537 described species worldwide, of which 356 are considered valid (Guiry & Guiry 2020). Sargassum has extensive intra and interspecific morphological variation (Cordero 1981, Moreira and Suárez 2002, Schell et al. 2015, Dibner et al. 2021), leading to the description of many morphology-based species and infra-species taxa (Mattio and Payri 2011). However, recent molecular-based systematic studies have shown the actual species diversity to be smaller than previously appreciated (Mattio et al. 2009, Phillips and Fredericq 2000, Yip et al. 2020). For example, revisions of the Sargassum flora from western Indian Ocean (Mauritius and Réunion) merged 44 taxa into 7 taxonomically defined species (Mattio et al. 2013). Another study conducted in Singapore recognized only 6 morphotypes against 41 previously recorded species (Yip et al. 2018). Revisions also found a smaller Sargassum diversity than historical records at French Polynesia, New Caledonia and South Africa, mainly due to misidentifications and unclear species delimitation (Mattio et al. 2008, 2015, Mattio and Payri 2009, Mattio et al. 2015).

In the Western Atlantic Ocean, 24 *Sargassum* species are reported (22 benthic and 2 pelagic) from Canada to southern Brazil (Taylor 1969, Ugadim 1973, Richardson 1975, Paula 1988, Camacho et al. 2015, Wynne 2017, González-Nieto et al. 2020). Despite some efforts to understand the dynamics of pelagic *Sargassum*, as well as the factors influencing the distribution of "brown tides" or "golden tides" (Lapointe 1986, 1995, Brooks et al. 2018, Oviatt et al. 2019, Wang et al. 2019, Godínez-Ortega et al. 2021, Jouanno et al. 2021), no forecast is available describing how climate change might shift benthic *Sargassum*'s habitat suitability, and hence future species distributions (but see Gouvêa et al. 2020).

Sargassum species are sensitive to local (e.g., pollution), regional (e.g., urban development), and global (e.g. climate change) impacts (Dawes and Tomasko 1988, Steen 2004, Engelen et al. 2005, Miki et al. 2016, Abe et al. 2020). *Sargassum* species are bioindicators of nutrient-enriched industrial effluents and heavy metals pollution, being capable of sequestration of metals like cadmium, zinc and arsenic (Alquezar et al. 2013, Guitouni et al. 2016, Ho and Bantoto-Kinamot 2021). The presence of healthy *Sargassum* beds in a particular reef indicates the healthiness of the entire benthic community (Eggertsen et al. 2017, Vroom and Braun 2010). Therefore, to understand how climate change affects *Sargassum* suitability, in terms of extent and range, is a crucial step to support management policies not only to protect and preserve algal beds but also marine benthic communities in general (Filbee-Dexter 2020).

The main objective of this study is to test whether global climate change in the form of ocean warming under future CO₂ emission scenarios will promote changes in Western Atlantic benthic *Sargassum* suitability areas. We hypothesize that *Sargassum* spp. suitability areas will: (1) increase toward higher latitudes; and (2) reduce in Equatorial latitudes. To test these hypotheses, we produce niche distribution models (or habitat suitability models) to estimate present and future suitability habitat areas under future global warming scenarios for 12 of the most common benthic *Sargassum* morpho-species in the tropical Western Atlantic Ocean and for one phylogenetic-defined species formed by the merge of 6 morpho-species, following recently published and unpublished molecular datasets. We also expected that future changes in distribution and suitability area will be more pronounced in the worst-case RCP 8.5 scenario than in the RCP 4.5 lower emission scenario.

Materials and Methods

Delimited area

Western Atlantic (WA) ocean raster was created with QGIS version 3.4.2 software (QGIS Development Team 2018), extending from 70.34845° on the North to -58.14374° southwards between -102.9249° and -17.35459° longitudes. WA raster was used as a mask for the worldwide abiotic variable rasters. Raster cells without marine environments (i.e., island) were excluded. Only areas with minimum depth lower than 265 m were maintained, since it is the deepest known record of a macroalgae (Littler et al. 1985).

Sargassum species

We selected 12 of the 24 WA *Sargassum* species. The species were selected based on the following criteria: (i) the species must be described from WA specimens; (ii) the species must be restricted to the WA or almost all known specimens must be from WA waters; (iii) the species must inhabit benthic hard-bottom environments (pelagic and drifting taxa were excluded); (iv) the species must be morphologically distinguishable; (v) the species must have minimal taxonomy concerns, at least morphology-wise (see Prud'homme van Reine 2011). Additionally, the species needed to have at least 5 different occurrence records along the WA. The species selected under these criteria were: *S. bermudense, S. buxifolium, S. cymosum, S. filipendula, S. furcatum, S. hystrix, S. platycarpum, S. polyceratium, S. pteropleuron, S. ramifolium, S. rigidulum*, and *S. stenophyllum*. While there is still a debate underlying the boundaries between some Atlantic *Sargassum* species, recent published and unpublished molecular phylogenetic results reported very low to no genetic divergence between *S.* *cymosum, S. filipendula, S. furcatum, S. hystrix, S. polyceratium* and *S. stenophyllum*, suggesting that they could be conspecific. They form an unresolved clade in maximum likelihood *Sargassum* subgenus *Sargassum* trees (Camacho et al. 2015; B. Barros-Barreto, M. T. Szechy & V. Cassano, unpub. data; Figure S1). Consequently, we also produced ecological niche modeling for a putative single species comprising the records of these six *Sargassum* species, herein named *Sargassum* WA.

Occurrence data

We searched for *Sargassum* species occurrence data in the Global Biodiversity Information Facility (GBIF, 2020), the speciesLink Network (CRIA 2018), references listed by AlgaeBase (Guiry and Guiry 2020), herbaria websites, and data provided by peer-reviewed indexed articles. Grey literature was also checked, including monographs, and annals of congress. For species with small sample sizes, we enhanced our literature survey effort to identify and assign correct coordinates. We excluded occurrence data without GPS coordinates and with coordinates in mainland or outside our study area, as well as probable misidentifications. With this dataset and WA raster, we remove duplicated coordinates, and only one occurrence per cell was considered. Detail of sorted species names, occurrence records and their latitudes and longitudes can be provided by request from the authors.

Environmental abiotic variables

We used abiotic variables rasters from Bio-ORACLE v.2.0 (Tyberghein et al. 2012; Assis et al. 2017) assisted by the "*leaflet*" (Cheng at al. 2019) and "*smdpredictors*" (Bosch 2018) packages for R (R Core Team 2019). We follow three criteria to select variables. First,
variables must be available in both present and future scenarios. Second, the variables must represent *Sargassum* biology and physiology needs, limits and constraints. Third, the correlation between the variables must be < 0.70 (see Figure S2). According to these criteria, we selected four variables: the minimum depth of the seafloor, maximum sea water temperature at minimum depth, mean sea water salinity at minimum depth, and mean sea ice thickness (see also Appendix S1).

Temperature is one of the main parameters that will change in climate change scenarios. Different studies ENMs with aquatic organisms show the importance of mean sea surface temperature (Jueterbock et al. 2013, Riul 2015, Jayathilake and Costello 2020), seabed temperature (Simon-Nutbrown et al. 2020), maximum and minimum sea surface temperature (Jueterbock et al. 2013, Riul 2015, Franco et al. 2018, Gouvêa et al. 2020, Jayathilake and Costello 2020, Prinz 2020), mean sea surface at summer and/or winter seasons (Assis et al. 2014, Martínez et al. 2018) and seasonal variation in sea surface temperatures (Sudo et al. 2019) to the models prediction. We choose only mean sea surface temperature between them for the following reasons: (i) our organisms are tropical perennial species; (ii) we take account for suitable areas where the species can be found, regardless their reproductive or germinative status; (iii) some *Sargassum* species are stenothermic (Earle 1969); and (iv) to avoid highly correlated variables in the delimited study area.

Salinity can also help to better understand the *Sargassum* distribution in the Atlantic Ocean. Some species are stenohaline and are not found nearby greater river discharges or mangroves (Earle 1969). For other macroalgae, salinity contributed for ENM models (Jueterbock et al. 2013, Sandman et al. 2013, Franco et al. 2018, Jonsson et al. 2018). Despite Assis et al. (2014) did not find any importance of salinity for the temperate Sargassaceae *Fucus vesiculosus*, minimum sea salinity met with *Sargassum horneri* important variables (Li

et al. 2020), and salinity was an important variable for benthic *Sargassum* globally distribution (Gouvêa et al. 2020).

Depth is variable that will not significatively change over the century in the Atlantic basin. Despite this aspect, bathymetry contributed to ENM models of shallow-water organisms (Simon-Nutbrown et al. 2020), including other Sargassaceae (Sandman et al. 2013, Jonsson et al. 2018). Otherwise, some species with specific thermic requirements may find in tropical deeper waters adequate environmental conditions, with sites eventually serving as refugia and contributing for populations genetic connectivity (Graham et al. 2007, Davis et al 2021).

Ice cover and distance of glacier are parameters commonly used to predict Arctic and Antarctic macroalgae suitability areas (Jerosch et al. 2019, Assis et al. 2022). Ice affects light input to above waters and consequently, photosynthetic rates (Runcie and Riddle 2006). Also, the ice crystals formation may lead to greater mechanical cell wall damage, even causing cell rupture (Sun and Li 2003, Bayer-Giraldi et al. 2014).

Prediction scenarios

We choose the following scenarios to predict suitability areas of *Sargassum* species: (i) present scenario (data range from 2000 to 2016); (ii) Representative Concentration Pathway (RCP) 4.5, a realistic scenario of greenhouse emissions, for the year 2100; and (iii) RCP 8.5, a "business-as-usual" pessimistic scenario of high carbon dioxide atmospheric concentrations at 2100 (IPCC 2014). We provided an ensemble of models for all 13 selected Sargassum species. For the species with more than 35 independent occurrence records, we provided an averaging ensemble of 11 modelling algorithms. For species with less than 35 records, we used the "leave-one-out-cross-validation approach" (see below). The species with >35 records were: *S. cymosum, S. filipendula, S. hystrix, S. polyceratium, S. platycarpum, S. pteropleuron*, and the *Sargassum* WA species described above (Table 1).

We used profile, regression and machine learning methods. We generated and ran ten times the following ENM algorithms for each morpho-species, plus the Sargassum WA species: generalized linear model (GLM; McCullagh and Nelder 1989), generalized additive model (GAM; Hastie and Tibshirani 1990), classification and regression trees (CART; Breiman et al. 1984), boosted regression trees (BRT; Friedman 2001), multivariate adaptive regression spline (MARS; Friedman 1991), random forests (RF; Breiman 2001), support vector machine (SVM; Vapnik 1995), artificial neural networks (ANN; Rosenblatt 1958), maximum entropy (Maxent; Phillips et al. 2006), maxlike (Royle et al. 2012), and bioclim (Busby 1991). Some ENM algorithms require presence and absence data, while other algorithms need presence and background coordinates. For both algorithm categories we provided pseudo-absences points as either absence or background data. Pseudo-absences represent points in geographical space where the species probably do not occur (Jayathilake and Costello 2020). We randomly set one group of pseudo-absences points into our study area in an equal number of occurrences registered for each species (Phillips et al. 2009). We visually checked the pseudo-absences points to make sure they were reasonable. Pseudoabsence coordinates for each species can be found in Supplementary Table S2.

As a result of our selection criteria, we were left with 4 environmental variables (see above) to fit the models. We utilized 75% of randomly selected presence and pseudo-absence points as train data, and the other 25% of the points for testing the models. This procedure was repeated for each algorithm, species, and scenario. After each model was generated, the "sdm" program automatically changed the points used for train and test (Naimi and Araujo 2016). To predict the models in present and futures scenarios, we use only models with Area Under Receiver Operating Characteristic Curve (ROC-AUC) ≥ 0.70 and true skill statistics $(TSS) \ge 0.60$. The mean parameters of each algorithm for each species can be assessed in Supplementary Table S1. We used "sdm" package to identify the suitability limits that maximizes TSS for each model. TSS provide a good balance between sensitivity and specificity and is independent of species prevalence (Allouche et al. 2006). Each model, from each algorithm outputted a map that was binarized following the limit maximization of TSS of each model. Values above TSS maximization limits were considered suitable niche values for a particular species. Values below the TSS maximization limits were considered unsuitable habitats. An averaging ensemble map for current and future scenarios was built by summing all generated suitability maps. For the final ensemble, suitability areas were considered true for areas recognized as suitable in at least 50% of generated maps.

Species with small sample sizes

For species with less than 35 occurrence records, we modeled ecological niche using leave-one-out-cross-validation (LOOCV; "jackknife approach") in the Maxent program, version 3.6 (Pearson et al. 2007, Shcheglovitova and Anderson 2013, see Table 1). This approach leads to more realistic analyses for species with small sample sizes. We used default parameters for convergence threshold (10⁵), with random sample of 10000 background points

to fit present environmental conditions and 500 iterations. We choose "crossvalidation" in "replicate run type", using the number of occurrence records as the number of replicates. We binarized each map using T10 (fixed cumulative value 10 complementary log-log transformation threshold, which considers an omission rate of 10%) as a minimum value, following Pearson et al. (2007). An ensemble was then prepared using only the cells present in at least 50% of the maps (Buckley et al. 2011, Principe et al. 2021).

Analytical notes

We compared the maximum northward and southward latitudes and the total suitability areas for each species, the change in the area occupied (present and two future scenarios), and the overlapping between present and future scenarios in the ensemble maps. Non-parametric Spearman correlations were calculated between percentage change in suitability area and percentage changes in overlap area, as well as between percentual changes in suitability area and latitudinal variation.

All the analyses were done in R using the packages *dismo* (Hijmans et al. 2017), *plyr* (Wickham 2011), *rgeos* (Bivand and Rundel 2019), *raster* (Hijmans 2020), *rgdal* (Bivand et al. 2019), *sdm* (Naimi and Araujo 2016), *SDMTools* (VanDerWal et al. 2014) and *usdm* (Naimi et al. 2013). Complete scripts used in this study are available in Appendix S3.

Results

Changes in latitudinal limits

Latitudinal limits of all species increased southward 1.08° in average under the RCP 4.5 scenario (range: from 0.17° for *S. filipendula* and *Sargassum* WA to 4.08° for *S. buxifolium*) and 2.92° under the RCP 8.5 scenario (range: from 0.25° for *Sargassum* WA to 5.5° for *S. rigidulum*). The only exception was *S. furcatum* who did not change its current south latitudinal limit under the RCP 4.5 scenario (Table 2). Species with the highest values in terms of their southernmost latitude limits in the present day, like *S. bermudense*, *S. filipendula* and *S. polyceratium*, showed the smallest values of future change in southern latitudinal limits under both RCP scenarios (0.7° to 1.0° southward shifts) (Figure 1B).

Northern latitudinal limits of all species increased northward by 2.14° in average under the RCP 4.5 scenario (from 0.50° for *S. cymosum* to 5.42° for *S. bermudense*) and 4.24° under the RCP 8.5 scenario (range: 1.67° for *Sargassum hystrix* to 8.02° for *S. bermudense*), without exceptions (Table 2). Species with the highest values in northernmost latitude limits in the present day (*S. bermudense, S. filipendula, S. buxifolium* and *Sargassum* WA) showed the greatest values of northward changes in latitude distributional limits under both RCP scenarios (Figure 1A).

There was no correlation between change in suitability area and latitudinal range, except for northward variation in RCP 8.5 scenario (Spearman rank correlation: rho = 0.58, S = 152, df = 11, p = 0.02, Figure S3). We found two different groups of species, according to the magnitude of their change in area. For S. bermudense, S. filipendula, S. hystrix, S. platycarpum, S. polyceratium and S. pteropleuron the models suggest an increase in suitability area in both scenarios, RCP 4.5 from 4.2% to 188%, and RCP 8.5 from 9.7% to 167.8%, respectively (Table 2, Figure 1). However, for the other six species (S. buxifolium, S. cymosum, S. furcatum, S. ramifolium, S. rigidulum and S. stenophyllum) the models showed a reduction in suitability area in both scenarios, RCP 4.5 from -10.1 to -60.4%, and RCP 8.5 from -23.8% to -80.2%. Greater changes in future suitability areas were found for species with the smallest present-day predicted suitability areas (S. pteropleuron with + 167.8%, and S. stenophyllum with -80.2%, Figure 3). For Sargassum WA, we found an 1% increase in suitability area in RCP 4.5 scenario and a -13.1% decrease in RCP 8.5 prediction (Table 2, Figure S4).

Changes in overlap area

Species that increased their suitability area in future RCP scenarios are expected to maintain their current suitability area, with 100% overlap between present and futures scenarios (Table 2). Conversely, species which showed reductions in future suitability areas, also reduced in the same magnitude their overlap areas. A high positive correlation between areas of overlap and changes in suitability area between present and future scenarios were observed (Spearman rank correlation: RCP 4.5, rho = 0.92, S = 29.2, df = 11, p < 0.001; RCP 8.5, rho = 0.95, S = 17.9, df = 11, p = 3×10^{-7}). Reduction in overlap areas occurred mainly in the Caribbean region, despite an increase in their poleward range suitability.

The mean AUC for the test data ranged between 0.82 and 0.97 (Table 1). To compose the ensemble forecasting of the predictions, we excluded algorithms that do not run or fit well (AUC < 0.8). So, we don't consider Bioclim's predictions for *S. filipendula*, GAM's models for *S. platycarpum*, Bioclim, GAM, CART and MAXLIKE predictions for *S. pteropleuron* and Bioclim, GLM, ANN and MAXLIKE algorithms for *Sargassum* WA (see Table S1).

Predictors contributions

The most important abiotic variable for all species was maximum sea water temperature at minimum depth, which contribution varied between 56.1% (in *S. hystrix*) and 99.5% (in *S. bermudense*) for the models (Table 1). Temperature contributed for more than 90% of the models in *S. bermudense* and *S. buxifolium*. Temperature was followed by salinity, depth and ice cover. Mean annual salinity contribution ranged between 0.2% for *S. bermudense* to 36.2% for *S. furcatum*. For all species but *S. bermudense* and *S. buxifolium*, salinity contribution was greater than 5%. For seven species (*S. cymosum, S. filipendula, S. hystrix, S. platycarpum, S. polyceratium* and *S. petropleuron,* and *Sargassum* WA) depth contribution was greater than 5%. Mean sea ice thickness contribution ranged between 0%, for species modeled only with Maxent, to 10% for *Sargassum* WA, ice thickness contributed > 5%.

Discussion

Latitudinal patterns

Our niche distribution models predicted that all 12 tropical *Sargassum* morphospecies and also *Sargassum* WA will expand their potential distribution poleward in both north and southward directions. This prediction evidences the ongoing process of tropicalization of temperate marine benthic ecosystems which has already been observed in other parts of the world (Lima et al. 2007, Vérges et al. 2014, Thibaut et al. 2015, Vérges et al. 2016, Smale 2020). For example, local kelp and temperate *Sargassum* forests have been replaced by tropical Phaeophycean species, including dense tropical *Sargassum* beds, in southern Japan (Tanaka et al. 2012). Significant changes in community composition are welldocumented in Western Australian warm temperate kelp communities after marine heat waves, with the decrease in the abundance and cover of temperate kelp and temperate animals' species, followed by their replacement by tropical species (Wernberg et al. 2016). Niche distribution models for a range of temperate seaweed species also forecast range contractions and further poleward shifts (Martínez et al. 2018). The absence of canopyforming species in areas where dominant temperate species become absent are expected to be occupied by tropical canopy-forming species.

The shift poleward is greater in RCP 8.5 scenario than in RCP 4.5 for all species, following the observed pattern of other macroalgae (Martínez et al. 2018, Li et al. 2020, Prinz 2020, Simon-Nutbrown et al. 2020). For 10 of the 13 *Sargassum* species here studied, habitat suitabilities were forecasted to expand northward more than 3° of latitude under the RCP 8.5 scenario, while only 5 species had a southward expansion predicted for the same magnitude. Our results suggest that potential tropicalization of high latitude habitats due to poleward range shifts in *Sargassum* species distribution might be more prevalent in the Northern

Hemisphere. *S. bermudense*, for example, showed the highest predicted northward shift value of habitat suitability range (8.08°), which is equivalent to a potential 1700 km increase in its northward distributional range (i.e., from Cape Cod, Massachusetts, USA, to north Newfoundland, Canada). This prediction could appear extreme, however, a northward ranged shift of 593 km in ~50 years has already been reported for *Sargassum flavifolium* along Portugal's coastline (Lima et al. 2007).

Local and regional impacts caused by pollution, urbanization, heavy balneability use, and environmental degradation can reduce the abundance of *Sargassum* beds in lower latitudes (Barradas et al. 2022). Empirical evidence of local and regional *Sargassum* bed losses, particularly those associated to highly urbanized coastal areas along the Western Atlantic Ocean has been documented (Scherner et al. 2013). Gorman et al. (2020) for example, reported decline in *Sargassum* spp. cover in the order of 2.6% per year, in the last five decades across 48 sites along 1000 km of the subtropical Brazilian coast. An average total loss of 52% cover across sites was reported, ranging from 20% to 89% cover loss depending on the studied site. Loss of climate suitability overlap herein predicted for six *Sargassum* morpho-species and *Sargassum* WA, predicted to occur mostly in the Caribbean region, will probably be aggravated by local and regional stressors, inducing greater losses of niche overlap.

Suitability Area

Different from our initial expectations, not all *Sargassum* species will suffer future reduction in their suitability area under both RCP 4.5 and RCP 8.5 scenarios. Half of *Sargassum* species will increase their suitability areas, while the other half will decrease their suitability areas in tropical latitudes, mainly in Central America, Gulf of Mexico and Caribe. A similar pattern was observed for 5 out of 6 tropical to warm-temperate species evaluated in

Southwestern Atlantic by Riul (2015). In these cases, species may develop new or exacerbate ongoing disjunct distributions, potentially leading to a decrease in genetic connectivity between southern and northern populations and isolate or reduced populations raising extinction probability (*sensu* MacArthur and Wilson 1963).

Our results suggest *S. stenophyllum* will be the most negatively affected by future climate change (in both RCPs scenarios). *S. stenophyllum* has the second smallest predicted present day suitability area and model forecasts show future disjunct antitropical distributions. Under future RCPs scenarios, low latitude *S. stenophyllum populations* may find suitable environmental conditions only offshore, in deeper, possibly mesophotic, habitats. Compared to other Western Atlantic *Sargassum* species, *S. stenophyllum* has relatively slower growth rates (Paula and Eston 1987) which may jeopardize even further its occurrence in coastal tropical and equatorial latitudes under future climate scenarios.

Sargassum pteropleuron turned out to be the species with the smallest present day suitability area but showing the biggest expansion in future suitability area. *S. pteropleuron* more than doubled its suitability area under RCP 4.5 and RCP 8.5 scenarios. Interestingly, *S. pteropleuron* is the species with the smallest number of distributional records. Nowadays, *S. pteropleuron* is restricted to the Caribbean and North America coasts (Prince and O'Neal 1979, Díaz-Piferrer 1981, Camacho et al. 2015), despite being an eurythermal and euryhaline species (Earle 1969). *S. pteropleuron* along with his sister-species, *S. platycarpum*, are genetically distinct from the younger Sargassum WA clade and belong to an independent lineage that dispersed throughout Atlantic less than 1.5 Mya, after the closure of Panama Isthmus (Phillips and Fredericq 2000, Camacho et al. 2015, Yip et al. 2020).

Except for *Sargassum* WA, the RCP 8.5 scenario indicated larger changes in suitability area for all species than those observed under the RCP 4.5 scenario. Species with present day reduced suitability area are expected to lose more area under the RCP 8.5 scenario

than under the RCP 4.5 scenario. Species that increase their suitability area, show further increase under the RCP 8.5 scenario. This pattern of continuous and proportional increased in suitability area as the atmospheric carbon dioxide concentration increase has been reported for other marine species, including phaeophycean (Sudo et al. 2019, Prinz 2020, Li et al. 2020, Simon-Nutbrown et al. 2020, Principe et al. 2021). For the phylogenetic species *Sargassum* WA, the 1% raise in suitability area at RCP 4.5 scenario and 13.1% reduction at RCP 8.5 are similar to predictions observed for the warm-temperate *Sargassum horneri* (Li et al. 2020). *S. horneri* projections made by 2100 show a 2.3% increase in suitability area at RCP 2.6 scenario followed by a 87.6% decrease at RCP 8.5 (Li et al. 2020). The *Sargassum* WA differences in both scenarios led us to think that a little enhance in seawater temperature may have a very discrete positive effect on the specie, but in RCP 8.5 scenario, the seawater temperature may be hotter than its physiological requirements for survival (see Bozinovic and Pörtner 2015).

According to recent phylogenetic studies, morphological data and classic taxonomy (Mattio and Payri 2011, Camacho *et al.* 2015, Yip *et al.* 2020), all species that show reduction in their suitability areas are included in *Sargassum* Subgenus *Sargassum* Section *Sargassum*. These species form a morphological complex with recent evolutionary origin (≤ 1 Mya) whose ancestors arrived recently, ~200-400 thousand years ago, in the Atlantic Ocean (Yip et al. 2020). These species simply might not yet be evolutionarily adapted to future warmer ocean conditions and must migrate towards the pole to find temperatures similar to present day conditions or be extinct (Wiens 2016).

Overlap

For all species herein studied, very little divergence between future suitability areas and the overlap areas were observed. *Sargassum* species that showed an increase in future suitability area will keep their current suitability area (as probably their current actual area of occurrence) and just add further range extensions poleward. A reduction in overlap areas was only observed in *Sargassum* species showing a decrease in their future suitability areas. The main pattern observed was the maintenance of the present species suitability area - entirely or partially - with a little enhancement of the suitability area toward the poles. Riul (2015) found a similar pattern only for one tropical Brazilian waters macroalga, *Dictyopteris jolyana*. The overlap areas for 5 subtropical or "warm-temperate species" (*Gelidium coarctatum, Ceramium brasiliense, Cryptonemia delicatula, Levringea brasiliensis* and *Plocamium brasiliense*) did not match their present suitable habitats and the local maintenance of some species will be only possible with spatially disjunct populations (Riul 2015).

Further advances in molecular and genetic techniques to improve species delimitation and phylogenetic resolution among putative *Sargassum* species within the *Sargassum* WA species complex will help us better forecast future scenarios for potentially different *Sargassum* species, subspecies, or phenotypes. To model *Sargassum* WA, we used only *Sargassum* species for which molecular data is available. Thus, our *Sargassum* WA species comprises data for only 6 WA species. Molecular data from Atlantic *Sargassum* morpho-species included or excluded in this study might turn out to belong to the *Sargassum* WA model results remain unknown. For example, *S. buxifolium* was described as a variety of *S. hystrix* (J. Agardh 1848) and only very recently new phylogenetic data demonstrate their very close phylogenetic relationship, pointing towards conspecificity (Yip et al. 2020). The morphological proximity between *S. bernudense* and *S. cymosum* has been reported elsewhere (Schneider 2003, Schneider and Flook, 2017). Very recently, González-Nieto et al. (2020) using molecular techniques considered these two species into the same polytomy as *Sargassum* WA species. Molecular-based phylogenetic data for *S. rigidulum* and *S.*

ramifolium, including other Atlantic *Sargassum* species, remain absent. In the past, *S. rigidulum* has been confused with *S. cymosum* (Oliveira Filho 1977, Diaz-Pulido & Díaz-Ruíz 2003, Wynne et al. 2014, Cordero 2016, Gabriel & Fredericq 2019) and *S. ramifolium* was considered as a *S. cymosum* variety (Grunow 1916, Paula 1988). Therefore, there is a high likelihood that these two species might belong to the *Sargassum* WA species complex as well.

Research Limitations

Our work considers only 4 environmental variables that cover the entire study area for all considered scenarios. Other abiotic and climatic variables (e.g., specific nutrients and co-factors) could help improve the fit of our models but unfortunately data availability is spatially or temporarily limited, or non-existent. An environmental variable that could substantially enhance our predictions are the presence of hard substrata. Benthic *Sargassum* are stenoancyric species, only fixing their holdfast in consolidated subtract, like rocks, corals, and anthropogenic facilities, rarely on wood (Earle 1969). The main difference in our suitability area projected for the present in relation to the known occurrence are the presence of suitability areas in places with mud or sand substrata. A detailed raster with substrate information could exclude areas with only soft-bottom seabed. Nevertheless, shipwrecks and oil platforms may have led to a *Sargassum* colonization of these new available substrata.

An increase in the number of collections of species with few records (i.e., *S. bermudense*) and in undersampled locations can increase the number of occurrences, improving the output of the most sensitive algorithms. Likewise, advances in modeling, with new and better algorithms, will increase prediction accuracy. Other considerations for improving the models and the predictions are discussed in Appendix S2.

Management and Conservation Strategies

Our results highlight the possibility of tropical WA *Sargassum* becoming part of the macroalgal component in current temperate environments. The tropicalization of temperate communities may be more evident in the Northern Hemisphere, where tropical *Sargassum* showed greater latitudinal displacement. Also, northern boundaries of the present suitability habitats have a greater area with shallow waters than the southern boundaries.

Habitats today occupied by species showing future reduction in suitability areas (i.e., *S. cymosum, S. stenophyllum, S buxifolium*) may be occupied by other *Sargassum* species (i.e., *S. hystrix, S. polyceratium*). The impact of *Sargassum* species' replacement over the algal community is currently unknown and hard to be determined. Depending on the velocity of ocean warming, luxuriant *Sargassum* beds may be replaced by habitats of less structural complexity, such as barrens or turfs (Airoldi et al. 2014, Gorman et al. 2020). The importance of management policies, including Marine Protected Areas and local habitat restoration, may warrant species conservation (Gianni et al. 2013, Schneider and Flook 2017). Research focused on mapping, monitoring and conservation are needed and must be encouraged.

Conclusions

Our results on the effect of climate change on WA *Sargassum* species distributions showed us an increase in latitudinal range toward the poles for all 13 species here considered, with broader effects on RCP 8.5 scenario than in RCP 4.5. Considering that *Sargassum* spp represent foundation species, this suggests a move of tropical systems into warm temperate habitats at higher latitudes. Half of the species showed an increase in its suitability area, while the other half showed a reduction in suitability area. Whenever there was an increase in the suitability area, the future projection incorporated the entire suitability area predicted for the present scenario. Species with reduced suitability don't present a major shift in relation to the present distribution but show a decrease in the suitability of tropical regions. Attention should be paid to an eventual increase in the area of occurrence of *S. pteropleuron* and to a potential drastic reduction in the Tropical populations of *S. stenophyllum, S. cymosum* and *S. buxifolium*. Efforts for conservation and management initiatives of these species must be maintained and expanded.

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References

Abe, H., Suzuki, H., Kumagai, N. H. & Hiroya Yamano, H. 2020. Distribution gradient of fucoid algae (Sargassaceae, Phaeophyta) along the coastline of Okinawa Island, southern Japan: Relationship to environmental factors. *Regional Studies in Marine Science*, 40, 101526.

Agardh, J. G. 1848. Species genera et ordines algarum, seu descriptiones succinctae specierum, generum et ordinum, quibus algarum regnum constituitur. Volumen Primum. Algas fucoideas complectens. I. Lundae [Lund]: C.W.K. Gleerup., 363p.

Airoldi, L., Ballesteros, E., Buonomo, R., Van Belzen, J., Bouma, T. J., Cebrian, E., De Clerk, O., Engelen, A. H., Ferrario, F., Fraschetti, S., Gianni, F., Guidetti, P., Ivesa, L., Mancuso, F. P., Micheli, F., Perkol-Finkel, S., Serrão, E. A., Strain, E. M., Mangialajo, L. 2015. Marine forests at risk: solutions to halt the loss and promote the recovery of Mediterranean canopy-forming seaweeds. *Proceedings of the 5th Mediterranean Symposium on Marine Vegetation (Portorož, Slovénie, 27-28 octobre 2014)*, pp. 28–33.

Allouche, O., Tsoar, A. & Kadmon, R. 2006. Assessing the accuracy of species distribution models: prevalence, kappa and the true skill statistic (TSS). *Journal of Applied Ecology* 43:1223–1232.

Alquezar, R., Glendenning, L. & Costanzo, S. 2013. The use of the brown macroalgae, *Sargassum flavicans*, as a potential bioindicator of industrial nutrient enrichment. *Marine Pollution Bulletin*. 77:140-146.

Assis, J., Serrão, E., Claro, B., Perrin, C. & Pearson, G. 2014. Climate-driven range shifts explain the distribution of extant gene pools and predict future loss of unique lineages in a marine brown alga. *Molecular Ecology* 23(11):2797-2810.

Assis, J., Tyberghein, L., Bosh, S., Verbruggen, H., Serrão, E. A. & De Clerck, O. 2017. Bio-ORACLE v2.0: Extending marine data layers for bioclimatic modelling. *Global Ecology and Biogeography* 27(3):277-284.

Assis, J., Serrão, E. A., Duarte, C. M., Fragkopoulou, E. & Krause-Jensen, D. 2022. Major Expansion of Marine Forests in a Warmer Arctic. *Frontiers in Marine Science* 9:850368.

Barradas, J. I., Chow, F., Dias, G. M., & Ghilardi-Lopes, N. P. 2022. Response of *Sargassum* Beds (Fucales) to Human Trampling: Reproductive, Morphological, and Biochemical Aspects. *Estuaries and Coasts* 45(2):501-509.

Barry, J. P., Baxter, C.H., Sagarin, R.D. & Gilman, S.E. 1995 Climate related, long-term faunal changes in a California rocky intertidal community. *Science* 267:672–675.

Bayer-Giraldi, M., Jin, E. S. & Wilson, P. 2014. Characterization of Ice Binding Proteins from Sea Ice Algae. *Methods in molecular biology* 1166:241-53.

Bivand, R., Keitt, T. & Rowlingson, B. 2019. rgdal: Bindings for the 'Geospatial' Data Abstraction Library. R package version 1.4-8. Available at: <u>https://CRAN.R-project.org/package=rgdal</u> (last accessed 4 June 2022).

Bivand, R. & Rundel, C. 2019 . rgeos: Interface to Geometry Engine - Open Source ('GEOS'). R package version 0.5-2. Available at: <u>https://CRAN.R-project.org/package=rgeos</u> (last accessed 4 June 2022).

Bosch, S. 2018. sdmpredictors: Species Distribution Modelling Predictor Datasets. R package version 0.2.8. Available at: <u>https://CRAN.R-project.org/package=sdmpredictors</u> (last accessed 4 June 2022).

Bozinovic, F. & Pörtner, H.-O. 2015. Physiological ecology meets climate change. *Ecology and Evolution* 5:1025–1030.

Brayner-Barros, S. G., Eskinazi-Leça, E., Oliveira, N. M. B. 2006, Diatomáceas epífitas em *Sargassum polyceratium* (Phaeophyta) coletada no litoral de Pernambuco. *Anais do XI Congresso Brasileiro de Ficologia, Itajaí (SC)*. pp.113-121.

Breiman, L., Friedman, J., Stone, C. J. & Olshen, R. A. 1984. Classification and Regression Trees. Chapman & Hall/CRC, New York, 368pp.

Breiman, L. 2001. Random Forests. Machine Learning 45: 5-32.

Brooks, M. T., Coles, V. J., Hood, R. R. & Gower, J. F. 2018 Factors controlling the seasonal distribution of pelagic *Sargassum*. Mar. Ecol.-Prog. Ser., 599:1–18.

Buckley, L. B., Waaser, S.A., MacLean, H.J. & Fox, R. 2011. Does including physiology improve species distribution model predictions of responses to recent climate change? *Ecology* 92(12):2214-21.

Buonomo, R., Chefaoui, R. M., Lacida, R. B., Engelen, A. H., Serrão, E. A. & Airoldi, L. 2018. Predicted extinction of unique genetic diversity in marine forests of *Cystoseira* spp. *Mar. Environ. Res.* 138:119–128.

Busby, J. R. 1991. BIOCLIM – a bioclimate analysis and prediction system. Plant Protection Quarterly (Australia) 6(1):8-9.

Camacho, O., Mattio, L., Draisma, S., Fredericq, S. & Diaz-Pulido, G. 2015. Morphological and molecular assessment of *Sargassum* (Fucales, Phaeophyceae) from Caribbean Colombia, including the proposal of *Sargassum giganteum* sp. nov., *Sargassum schnetteri* comb. nov. and *Sargassum* section *Cladophyllum* sect. nov. *Systematics and Biodiversity* 13(2):105-130.

Casado-Amezúa, P., Araújo, R., Bárbara, I., Bermejo, R., Borja, Á., Díez, I., Fernández, C., Gorostiaga, J. M., Guinda, X., Hernández, I., Juanes, J. A., Peña, V., Peteiro, C., Puente, A., Quintana, I., Tuya, F., Viejo, R. M., Altamirano, M., Gallardo, T. & Martínez, B. 2019. Distributional shifts of canopy-forming seaweeds from the Atlantic coast of Southern Europe. *Biodiversity and Conservation* 28:1151–1172.

Cheminée, A., Sala, E., Pastor, J., Bodilis, P., Thiriet, P., Mangialajo, L., Cottalorda, J.-M. & Francour, P. 2013. Nursery value of *Cystoseira* forests for Mediterranean rocky reef fishes. *J. Exp. Mar. Biol. Ecol.* 442:70–79.

Cheng, J., Karambelkar, B. & Xie, Y. 2019. leaflet: Create Interactive Web Maps with the JavaScript 'Leaflet' Library. R package version 2.0.3. Available at: <u>https://CRAN.R-project.org/package=leaflet</u> (last accessed 4 June 2022).

CRIA - Centro de Referência em Informação Ambiental. 2018. SpeciesLink Network. Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP). Available at <u>http://www.splink.org.br</u> (last accessed 4 June 2022).

Connor, J. & Adey, W. 1977. The benthic algal composition, standing crop, and productivity of a Caribbean algal ridge. *Atoll Res. Bull.* 211:1-40.

Cordero, M. K. 2016. Caracterización ficológica del litoral rocoso y manglares de Yapascua, Parque Nacional San Esteban, Estado de Carabobo. Trabajo Especial de Grado en Licenciado en Biología. Universidad de Carabobo, Valencia, Venezuela. 137pp.

Cordero, P. A., Jr. 1981. Eco-morphological observation of the genus *Sargassum* in central Philippines, including notes on their biomass and bed determination. *Proc. Fourth Int. Coral Reef Symp.* 2:399-409.

Davis, T. R., Champion, C., & Coleman, M. A. 2021. Climate refugia for kelp within an ocean warming hotspot revealed by stacked species distribution modelling. *Marine Environmental Research* 166:105267.

Dawes, C. J. & Tomasko, D. A. 1988. Physiological responses of perennial bases of *Sargassum filipendula* from three sites on the west coast of Florida. *Bull Mar Sci* 42:166–173.

Díaz-Piferrer, M. 1981. The genus *Sargassum* in western Atlantic: a biogeographical approach. *Proc. Int. Seaweed Symp.* 1:307-312.

Diaz-Pulido, G. & Diaz-Ruiz, M. 2003. Diversity of Benthic Marine Algae of the Colombian Atlantic. *Biota Colombiana* 4(2):203-246.

Dibner, S., Martin, L., Thibaut, T., Aurelle, D., Blanfuné, A., Whittaker, K., Cooney, L., Schell, J., Goodwin, D. & Siuda, A. 2021. Consistent genetic divergence observed among pelagic *Sargassum* morphotypes in the western North Atlantic. *Marine Ecology* 43(1): e12691.

Earle, S. A. 1969. Phaeophyta of the eastern Gulf of Mexico. Phycologia 7:71-254.

Engelen, A. H., Aberg, P., Olsen, J. L., Stam, W. T. & Breeman, A. M. 2005. Effects of wave exposure and depth on biomass, density and fertility of the fucoid seaweed *Sargassum polyceratium* (Phaeophyta, Sargassaceae). *European Journal of Phycology* 40:149-158.

Eggertsen, L., Ferreira, C., Fontoura, L., Kautsky, N., Gullström, M. & Berkström, C. 2017. Seaweed beds support more juvenile reef fish than seagrass beds: Carrying capacity in a south-western Atlantic tropical seascape. *Estuarine, Coastal and Shelf Science* 196:97-108.

Farnham, W. F., Murfin, C., Critchley, A. T. & Morrell, S. L. 1981. Distribution and control of the brown alga *Sargassum muticum*. *Proc. Int. Seaweed Symp.* 10:277-82.

Filbee-Dexter, K. 2020. Ocean Forests Hold Unique Solutions to Our Current Environmental Crisis. *One Earth*. 2:398-401.

Filbee-Dexter, K., Feehan, C. J. & Scheibling, R. E. 2016. Large-scale degradation of a kelp ecosystem in an ocean warming hotspot. *Marine Ecology Progress Series* 543:141–152.

Franco, J., Bertocci, I., Rodriguez, L., Martinez, B., Sousa Pinto, I. & Arenas, F. 2018. The 'golden kelp' *Laminaria ochroleuca* under global change: Integrating multiple eco-physiological responses with species distribution models. *Journal of Ecology* 106:47-58.

Friedman, J. H. 1991. Multivariate adaptive regression splines. Ann. Stat. 19:1-67.

Friedman, J. H. 2001. Greedy function approximation: a gradient boosting machine. *The Annals of Statistics* 29:1189-1232.

Gabriel, D. & Fredericq, S. 2019. The marine macroalgae of Cabo Verde archipelago: an updated checklist. *Arquipelago - Life and Marine Sciences* 36:39 -60.

GBIF - The Global Biodiversity Information Facility. 2020, What is GBIF? Available at: https://www.gbif.org/what-is-gbif (last accessed 1 June 2020).

Gianni, F., Bartolini, F., Airoldi, L., Ballesteros, E., Francour, P., Guidetti, P., Meinesz, A., Thibaut, T. & Mangialajo, L. 2013. Conservation and restoration of marine forests in the Mediterranean sea and the potential role of marine protected areas. *Adv. Oceanogr. Limnol.* 4:83–101.

Godínez-Ortega, J. L., Cuatlán-Cortés, J. V., López-Bautista, J. M. & Tussenbroek, B. I. v. 2021. A Natural History of Floating *Sargassum* Species (Sargasso) from Mexico. *In* Hufnagel, L. [Ed.]. *Natural History and Ecology of Mexico and Central America*. IntechOpen, London. Available at: <u>https://www.intechopen.com/chapters/76196</u> (last accessed 4 June 2022).

González-Nieto, D., Oliveira, M., Núñez Resendiz, M., Dreckmann, K., Mateo-Cid, L. & Sentíes, A. 2020. Molecular assessment of the genus *Sargassum* (Fucales, Phaeophyceae) from the Mexican coasts of the Gulf of Mexico and Caribbean, with the description of *S. xochitlae* sp. nov. *Phytotaxa* 461:254-274.

Gorman, D., Horta, P., Flores, A. A. V., Turra, A., Berchez, F. A. S., Batista, M. B., Lopes Filho, E. S., Melo, M S., Ignacio, B. L., Carneiro, I. M., Villaça, R. C. & Széchy, M. T. M. 2020. Decadal losses of canopy-forming algae along the warm temperate coastline of Brazil. *Global Change Biology* 26(3):1446-1457.

Gouvêa, L. P., Assis, J., Gurgel, C. F. D., Serrão, E. A., Silveira, T. C. L., Santos, R., Duarte, C. M., Peres, L. M. C., Carvalho, V. F., Batista, M., Bastos, E., Sissini, M. N. & Horta, P. A.

2020. Golden carbon of *Sargassum* forests revealed as an opportunity for climate change mitigation. *Science of The Total Environment* 729:138745 pp.1-10.

Graham, M. G., Kinlan, B. P., Druehl, L. D., Garske, L. E. & Banks, S. 2007. Deep-water kelp refugia as potential hotspots of tropical marine diversity and productivity. *Proceedings of the National Academy of Sciences of the United States of America* 104(42):16576-16580.

Grunow, A. 1916. Additamenta ad cognitionem Sargassorum. Verhandlungen der Kaiserlich-Koniglichen Zoologisch-Botanischen Gesellschaft in Wien. 66 ed. 185p.

Guimaraens, M. A., Combells, C. & Corbett, C. 1994. Species diversity and richness of reef building corals and macroalgae of reef communities in Discovery Bay, Jamaica. A*cta Biologica Leopoldensia* 16:41-50.

Guiry, M.D. & Guiry, G.M. 2020. AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. Available at: <u>https://www.algaebase.org</u> (last accessed 4 June 2022).

Guitouni, M., Madan, A. A. & Al-Shabeeb, S. S. 2016. Seaweeds as bioindicators of heavy metals pollution in Tarut Bay, Saudi Arabia. *International Journal of Advanced Research*. 4(10):1095-1105.

Harley, C. D. G., Anderson, K. M., Demes, K. W., Jorve, J. P., Kordas, R. L., Coyle, T. A. & Graham, M. H. 2012. Effects of climate change on global seaweed communities. *Journal of Phycology* 48:1064–1078.

Hastie, T. J. & Tibshirani, R. J. 1990. Generalized Additive Models. Chapman & Hall/CRC, New York/Boca Raton, 352pp. Hawkins, S. J., Southward, A., Genner, M. J. 2003. Detection of environmental change in a marine ecosystem – evidence from the western English Channel. *Science of the Total Environment* 310:245–256.

Helmuth, B., Harley, C. D., Halpin, P. M., O'Donnell, M., Hofmann, G. E. & Blanchette, C. A. 2002. Climate change and latitudinal patterns of intertidal thermal stress. *Science* 298:1015–1017.

Hijmans, R. J., Phillips, S., Leathwick, J. & Elith, J. 2017. dismo: Species Distribution
Modeling. R package version 1.1-4. Available at: <u>https://CRAN.R-</u>project.org/package=dismo (last accessed 4 June 2022).

Hijmans, R. J. 2020. raster: Geographic Data Analysis and Modeling. R package version 3.012. Available at: <u>https://CRAN.R-project.org/package=raster</u> (last accessed 4 June 2022).

Ho, M. T. G. & Bantoto-Kinamot, V. 2021. *Sargassum, Padina* and *Turbinaria* as bioindicators of cadmium in Bais Bay, Negros Oriental. *The Palawan Scientist* 13(1):90-98.

IPCC - Intergovernmental Panel on Climate Change. 2014. *Climate Change 2014 Synthesis Report: Summary for Policymakers*. IPCC, Geneva, Switzerland, 31pp.

Jayathilake, R. M. D. & Costello, M. J. 2020. A modelled global distribution of the kelp biome. *Biological Conservation* 252:108815 pp.1-10.

Jerosch, K., Scharf, F. K., Deregibus, D., Campana, G. L. Zacher, K., Pehlke, H., Falk, U., Hass, H. C., Quartino, M. L. & Abele, D. 2019. Ensemble Modeling of Antarctic Macroalgal Habitats Exposed to Glacial Melt in a Polar Fjord. *Frontiers in Ecology and Evolution* 7(207):1-16.

Jonsson, P. R., Kotta, J., Andersson, H. C., Herkül, K., Virtanen, E., Sandman, A. N. & Johannesson, K. 2018. High climate velocity and population fragmentation may constrain climate-driven range shift of the key habitat former *Fucus vesiculosus*. *Diversity and Distributions* 24:892–905.

Jouanno, J., Benshila, R., Berline, L., Soulié, A., Radenac, M.-E., Morvan, G., Diaz, F., Sheinbaum, J., Chevalier, C., Thibaut, T., Changeux, T., Ménard, F., Berthet, S., Aumont, O., Ethé, C., Nabat, P. & Mallet, M. 2021. A NEMO-based model of *Sargassum* distribution in the tropical Atlantic: Description of the model and sensitivity analysis (NEMO-Sarg1.0). *Geoscientific Model Development* 14:4069-4086.

Jueterbock, A., Tyberghein, L., Verbruggen, H., Coyer, J. A, Olsen, J. L. & Hoarau, G. 2013. Climate change impact on seaweed meadow distribution in the North Atlantic rocky intertidal. *Ecology and Evolution* 3(5):1356-1373.

Kleypas, J. A. 2019. Climate change and tropical marine ecosystems: A review with an emphasis on coral reefs. *UNED Research Journal* 11(1) Núm. especial: S24-S35.

Lapointe, B. E. 1986. Phosphorus-limited photosynthesis and growth of *Sargassum natans* and *Sargassum fluitans* (Phaeophyceae) in the western North Atlantic. *Deep-Sea Res. Pt. A* 33:391–399.

Lapointe, B. E. 1995. A comparison of nutrient-limited productivity in *Sargassum natans* from neritic vs. oceanic waters of the western North Atlantic Ocean. *Limnol. Oceanogr.* 40:625–633.

Levitus, S., Antonov, J. I., Boyer, T. P. & Stephens, C. 2000. Warming of the World Ocean. *Science* 287:2225–2229.

Li, J.-J., Huang, S.-H., Liu, Z.-Y. & Bi, Y.-X. 2020. Climate-Driven Range Shifts of Brown Seaweed *Sargassum horneri* in the Northwest Pacific. *Frontiers in Marine Science* 7:570881.

Lima, F. P. 2007. Biogeography of Benthic Invertebrate Assemblages on the Portuguese Rocky Coast: Relation with Climatic and Oceanographic Patterns. Ph.D. thesis, Faculdade de Ciências da Universidade do Porto, Porto, Portugal, 235 pp.

Lima, F. P., Ribeiro, P. A., Queiroz, N., Hawkins, S. J., & Santos, A. M. 2007. Do distributional shifts of northern and southern species of algae match the warming pattern? *Global Change Biology* 13:2592–2604.

Littler, M. M., Littler, D. S., Blair, S. M. & Norris, J. N. 1985. Deepest known plant life discovered on an uncharted seamount. *Science* 227(4682):57-59.

Lopes, R. & Videira, N. 2013. Valuing marine and coastal ecosystem services: An integrated participatory framework. *Ocean Coast. Manage.* 84:53-162.

MacArthur, R. H. & Wilson, E. O. 1963. An equilibrium theory of insular zoogeography. *Evolution* 17(4):373-387.

Mafra Junior, L. L. 2001. Bases para o manejo de *Sargassum cymosum* (Phaeophyta-Fucales) na enseada de Armação do Itapocoroy, Penha, SC. Oceanography monography. Universidade do Vale do Itajaí, Itajaí, Brasil, 51 pp.

Marcelino, V. R. & Verbruggen, H. 2015. Ecological niche models of invasive seaweeds. *Journal of Phycology* 51:606-620. Martínez, B., Radford, B., Thomsen, M. S., Connell, S. D., Carreño, F., Bradshaw, C. J. A., Fordham, D. A., Russell, B. D., Gurgel, C. F. D. & Wernberg, T. 2018. Distribution models predict large contractions of habitat-forming seaweeds in response to ocean warming. *Diversity and Distributions* 24(10):1350-1366.

Mattio, L. & Payri, C. E. 2009. Taxonomic revision of *Sargassum* species (Fucales, Phaeophyceae) from New Caledonia based on morphological and molecular analyses. *J. Phycol.* 45:1374–88.

Mattio, L. & Payri, C. E. 2011. 190 years of *Sargassum* taxonomy, facing the advent of DNA phylogenies. *Bot. Rev.* 77:31–70.

Mattio, L., Payri, C. E. & Stiger-Pouvreau, V. 2008. Taxonomic revision of *Sargassum* (Fucales, Phaeophyceae) from French Polynesia based on morphological and molecular analyses. *J. Phycol.* 44:1541–55.

Mattio, L., Payri, C. E. & Verlaque, M. 2009. Taxonomic revision and geographic distribution of the subgenus *Sargassum* (Fucales, Phaeophyceae) in the Western and Central Pacific islands based on morphological and molecular analyses. *J. Phycol.* 45:1213–27.

Mattio, L., Zubia, M., Loveday, B., Crochelet, E., Duong, N., Payri, C. E., Bhagooli, R. & Bolton, J. J. 2013. *Sargassum* (Fucales, Phaeophyceae) in Mauritius and Reunion, western Indian Ocean: taxonomic revision and biogeography using hydrodynamic dispersal models. *Phycologia* 52(6):578–94.

Mattio, L., Anderson, R. J. & Bolton, J. J. 2015. A revision of the genus *Sargassum* (Fucales, Phaeophyceae) in South Africa. *S. Afr. J. Bot.* 98:95–107.

McCullagh, P. & Nelder, J. A. 1989. Generalized linear models. Chapman and Hall, London/New York, 511 pp.

Miki, O., Nagai, T., Marzuki, M., Okumura, C., Kosugi, C. & Kato, T. 2016. Effects of Fe fertilizer eluate on the growth of *Sargassum horneri* at the germling and immature stages. *J. Appl. Phycol.* 28:1775–1782.

Moreira, L., & Suárez, A. M. 2002. Estudio del género *Sargassum* C. Agardh, 1820 (Phaeophyta, Fucales, Sargassaceae) en aguas cubanas. 3. Variaciones morfológicas de *Sargassum filipendula* C. Agardh. *Revista de Investigaciones Marinas* 23:59 -62

Morrissey, J. 1980. Community structure and zonation of macroalgae and hermatypic corals on a fringing reef flat of Magnetic Island (Queensland, Australia). *Aquat. Bot.* 8:91-139.

Muller, R., Laepple, T., Bartsch, I. & Wiencke, C. 2009. Impact of oceanic warming on the distribution of seaweeds in polar and cold-temperate waters. *Bot. Mar.* 52:617–638.

Naimi, B. & Araújo, M. B. 2016. sdm: a reproducible and extensible R platform for species distribution modelling. *Ecography* 39:368-375.

Naimi, B., Hamm, N. A. S., Groen, T. A., Skidmore, A. K. & Toxopeus A. G. 2013. Where is positional uncertainty a problem for species distribution modelling. *Ecography* 37(2):191-203.

Oliveira-Filho, E. C. 1977. Algas Marinhas Bentônicas do Brasil. Thesis in Phycology, Universidade de São Paulo, São Paulo, Brazil, 406 pp.

Oviatt, C. A., Huizenga, K., Rogers, C. S. & Miller, W. J. 2019. What nutrient sources support anomalous growth and the recent *Sargassum* mass stranding on Caribbean beaches? A review. *Mar. Pollut. Bull.* 145:517–525.

Parmesan, C. 2006. Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology, Evolution, and Systematics* 37:637–669.

Paula, E. J. 1988. O Gênero Sargassum C. Ag. (Phaeophyta - Fucales) no Litoral do Estado deSão Paulo, Brasil. Boletim de Botânica da Universidade de São Paulo 10:65-118.

Paula, E. & Eston, V. 1987. Are There Other *Sargassum* Species Potentially as Invasive as *S. muticum? Botanica Marina* 30(5):405-410.

Paula, E. J. & Oliveira-Filho, E. C.. 1980. Aspectos fenológicos de duas populações de Sargassum cymosum (Phaeophyta – Fucales) do litoral de São Paulo, Brasil. Bolm. Botânica, Univ. S. Paulo. 8:21-39.

Pearson, R. G., Raxworthy, C. J., Nakamura, M. & Peterson, A. T. 2007. Predicting species distributions from small numbers of occurrence records: a test case using cryptic geckos in Madagascar. *J. Biogeogr.* 34:102–117.

Pereira, S. M. B., Oliveira-Carvalho, M. F., Burgos, D. C. & Araújo, E. L. 2006. Caracterização estrutural das macroalgas do ambiente recifal da praia de Enseada dos Corais -Pernambuco - Brasil. *Anais do XI Congresso Brasileiro de Ficologia, Itajaí (SC)*. Itajaí, Brazil, pp.231-242. Phillips, N. & Fredericq, S. 2000. Biogeographic and phylogenetic investigations of the pantropical genus *Sargassum* (Fucales, Phaeophyceae) with respect to Gulf of Mexico Species. *Gulf of Mexico Science* 2:77-87.

Phillips, S. J., Anderson, R. P. & Schapire, R. E. 2006. Maximum entropy modeling of species geographic distributions. *Ecological Modelling* 190:231-259.

Phillips, S. J., Dudík, M., Elith, J., Graham, C. H., Lehmann, A., Leathwick, J. & Ferrier, S. 2009. Sample selection bias and presence-only distribution models: implications for background and pseudo-absence data. *Ecological Applications* 19(1):181–197.

Prince, J. S. & O'Neal, S. W. 1979. The ecology of *Sargassum pteropleuron* Grunow (Phaeophyceae, Fucales) in the waters off South Florida. I. Growth, reproduction and population structure. *Phycologia* 18(2):109-114.

Principe, S. C., Acosta, A. L., Andrade, J. E. & Lotufo, T. M. C. 2021. Predicted Shifts in the Distributions of Atlantic Reef-Building Corals in the Face of Climate Change. *Frontiers in Marine Science* 8:673086.

Prinz, A. 2020 Evolutionary History of the *Fucus spiralis / Fucus guiryi* Complex Large-scale Analysis of the Distribution Range and Genetic Structure across the Northern Hemisphere. Master thesis in Marine Biology, Universidade do Algarve. Portugal, 132 pp.

Prud'homme Van Reine, W. F. 2011. Report of the Nomenclature Committee for Algae: 11. *Taxon* 60(3):898-899.

QGIS Development Team. 2018. QGIS 3.22. Geographic Information System User Guide.QGISAssociation.Electronicdocument.Availableat: https://docs.qgis.org/3.4/en/docs/user_manual/index.html (last accessed 4 June 2022).

R Core Team. 2019. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Available at: <u>https://www.R-project.org/</u> (last accessed 4 June 2022).

Raut, Y., Morando, M. & Capone, D. G. 2018. Diazotrophic Macroalgal Associations with Living and Decomposing *Sargassum*. *Front. Microbiol.* 9:3127.

Richardson, W. D. 1975. The marine algae of Trinidad, West Indies. *Bulletin of the British Museum (Natural History) Botany* 5(3):1-143, 27 pls.

Riul, P. 2015. Modelagem de distribuição de espécies bênticas marinhas na costa do Brasil: bioinvasão, conservação e efeito das mudanças climáticas. Ph.D. thesis in Ecology, Universidade Federal de Santa Catarina, Florianópolis, Brazil, 132p.

Rosenblatt, F. 1958. The perceptron: a probabilistic model for information storage and organization in the brain. *Psychol. Rev.* 65(6):386-408.

Royle, J. A., Chandler, R. B., Yackulic, C. & Nichols, J. D. 2012. Likelihood analysis of species occurrence probability from presence-only data for modelling species distributions. *Methods Ecol. Evol.* 3:545–554.

Runcie, J. W. & Riddle, M. J. 2006. Photosynthesis of marine macroalgae in ice-covered and ice-free environments in East Antarctica. *European Journal of Phycology* 41(2):223-233.

Sandman, A. N., Wikström, S. A., Blomqvist, M., Kautsky, H. & Isaeus, M. 2013. Scaledependent influence of environmental variables on species distribution: A case study on five coastal benthic species in the Baltic Sea. *Ecography* 35:354-363.

Schell, J. M., Goodwin, D. S. & Siuda, A. N. 2015. Recent *Sargassum* inundation events in the Caribbean: shipboard observations reveal dominance of a previously rare form. *Oceanography* 28:8–11.

Scherner, F., Horta, P. A., Oliveira, E. C., Simonassi, J. C., Hall-Spencer, J. M., Chow, F., Nunes, J. M. C. & Pereira, S. M. B. 2013. Coastal urbanization leads to remarkable seaweed species loss and community shifts along the SW Atlantic. *Marine Pollution Bulletin* 76(1-2):106-115.

Schneider, C. W. 2003. An annotated checklist and bibliography of the marine macroalgae of the Bermuda Islands. *Nova Hedwigia* 76(3-4):275-361.

Schneider, C. W. & Flook, C. T. 2017. Could marine animal conservation laws be responsible for the decline or extirpation of macroalgal populations in Bermuda over the past century? *Botanica Marina* 60(6):591-602.

Simon-Nutbrown, C., Hollingsworth, P. M., Fernandes, T. F., Kamphausen, L., Baxter, J. M. & Burdett, H. L. 2020. Species Distribution Modeling Predicts Significant Declines in Coralline Algae Populations Under Projected Climate Change With Implications for Conservation Policy. *Frontiers in Marine Science* 7:575825, 14 pp.

Shcheglovitova, M. & Anderson, R. P. 2013. Estimating optimal complexity for ecological niche models: A jackknife approach for species with small sample sizes. *Ecological Modelling* 269:9-17.

Smale, D. A. 2020. Impacts of ocean warming on kelp forest ecosystems. *New Phytologist* 225(4):1447-1454.

Southward, A. J., Hawkins, S. J. & Burrows, M. T. 1995 Seventy years' observations of changes in distributions and abundance of zooplankton and intertidal organisms in the Western English Channel in relation to rising sea temperature. *Journal of Thermal Biology* 20:127–155.

Steen, H. 2004. Effects of reduced salinity on reproduction and germling development in *Sargassum muticum* (Phaeophyceae, Fucales). *Eur. J. Phycol.* 39:293–299.

Steneck, R. S., Graham, M. H., Bourque, B. J., Corbett, D., Erlandson, J. M., Estes, J. A. & Tegner, M. J. 2002. Kelp forest ecosystems: biodiversity, stability, resilience and future. *Environ. Conserv.* 29(4):436–459.

Sudo, K., Watanabe, K., Yotsukura, N. & Nakaoka, M. 2019. Predictions of kelp distribution shifts along the northern coast of Japan. *Ecological Research*. 35(1):47-60.

Sun, D.-W. & Li, B. 2003. Microstructural change of potato tissues frozen by ultrasoundassisted immersion freezing. *Journal of Food Engineering* 57(4):337-345.

Tanaka, K., Taino, S., Haraguchi, H., Prendergast, G. & Hiraoka, M. 2012. Warming off southwestern Japan linked to distributional shifts of subtidal canopy-forming seaweeds. *Ecology and Evolution* 2:2854-2865.

Taylor, W. R. 1969. Notes on the distribution of West Indian marine algae particularly in the Lesser Antilles with a bibliography of recent works on Eastern American tropical algae. *Contr. Univ. Mich. Herb.* 9:125-203.

Thibaut, T., Blanfuné, A., Boudouresque, C. F. & Verlaque, M. 2015. Decline and local extinction of Fucales in the French riviera: the harbinger of future extinctions? *Mediterr. Mar. Sci.* 16(1):206–224.

Thompson, R. C., Crowe, T. P. & Hawkins, S. J. 2002. Rocky intertidal communities: past environmental changes, present status and predictions for the next 25 years. *Environmental Conservation* 29:168–191.

Tsuda, R., 1972. Morphological, zonational, and seasonal studies on two species of *Sargassum* on the reefs of Guam. *Proc. Int. Seaweed Symp.* 7:40-44.

Tyberghein, L., Verbruggen, H., Pauly, K., Troupin, C., Mineur, F. & De Clerck, O. 2012 Bio-ORACLE: A global environmental dataset for marine species distribution modelling. *Global Ecology and Biogeography* 21:272–281.

Ueki, C., Murakami, A., Kato, T., Saga, N. & Motomura, T. 2010 Effects of nutrient deprivation on photosynthetic pigments and ultrastructure of chloroplasts in *Porphyra yezoensis*. *Nippon Suisan Gakkaishi* 76:375–382

Ugadim, Y. 1973. Algas marinhas bentônicas do litoral sul do estado de São Paulo e do litoral do estado do Paraná, I Divisão Chlorophyta. *Boletim de Botânica da Universidade de São Paulo* 1:11-77.

Vapnik, V. N. 1995. The nature of statistical learning theory. Springer, New York, 188 pp.

VanDerWal, J., Falconi, L., Januchowski, S., Shoo, L. & Storlie, C. 2014. SDMTools: Species Distribution Modelling Tools: Tools for processing data associated with species
distribution modelling exercises. R package version 1.1-221. Available at: <u>https://CRAN.R-</u>project.org/package=SDMTools (last accessed 4 June 2022).

Vásquez, J. A., Zuñiga, S., Tala, F., Piage, N., Rodríguez, D. C. & Alonso Vega, J. M. 2014. Economic valuation of kelp forests in northern Chile: values of goods and services of the ecosystem. *J. Appl. Phycol.* 26(2):1081–1088.

Vergés, A., Steinberg, P. D., Hay, M. E., Poore, A. G., Campbell, A. H., Ballesteros, E., Heck Jr., K. L., David J. Booth, D. J., Coleman, M. A., Feary, D. A., Figueira, W., Langlois, T., Marzinelli, E. M., Mizerek, T., Mumby, P. J., Nakamura, Y., Roughan, M., van Sebille, E. Gupta, A. S., Smale, D. A., Tomas, F., Wernberg, T & Wilson, S. K. 2014. The tropicalization of temperate marine ecosystems: climate-mediated changes in herbivory and community phase shifts. *Proceedings of the Royal Society B: Biological Sciences* 281(1789):20140846.

Vergés, A., Doropoulos, C., Malcolm, H. A., Skye, M., Garcia-Pizá, M., Marzinelli, E. M., Campbell, A. H., Ballesteros, E., Hoey, A. S., Vila-Concejo, A., Bozec, Y.-M. & Steinberg, P. D. 2016. Long-term empirical evidence of ocean warming leading to tropicalization of fish communities, increased herbivory, and loss of kelp. *Proceedings of the National Academy of Sciences* 113(48):13791-13796.

Vroom, P. S. & Braun, C. L. 2010. Benthic Composition of a Healthy Subtropical Reef: Baseline Species-Level Cover, with an Emphasis on Algae, in the Northwestern Hawaiian Islands. *PLoS ONE* 5(3):e9733. Wanders, J. B. W., 1976. The role of benthic algae in the shallow reef of Curaçao (Netherlands Antilles). II. Primary productivity of the *Sargassum* beds on the north-east coast submarine plateau. *Aquat. Bot.* 2:327-335.

Wang, M., Hu, C., Barnes, B. B., Mitchum, G., Lapointe, B. & Montoya, J. P. 2019. The great Atlantic *Sargassum* belt. *Science* 365:83–87.

Wernberg, T., Thomsen, M. S., Tuya, F., Kendrick, G. A., Staehr, P. A., & Toohey, B. D. 2010. Decreasing resilience of kelp beds along a latitudinal temperature gradient: Potential implications for a warmer future. *Ecology Letters* 13:685–694.

Wernberg, T., Smale, D. A., & Thomsen, M. S. 2012. A decade of climate change experiments on marine organisms: Procedures, patterns and problems. *Global Change Biology* 18:1491–1498.

Wernberg, T., Bennett, S., Babcock, R., de Bettignies, T., Cure, K., Depczynski, M., Dufois,
F., Fromont, J., Fulton, C., Hovey, R., Harvey, E., Holmes, T., Kendrick, G., Radford, B.,
Santana-Garcon, J., Saunders, B., Smale, D., Thomsen, M., Tuckett, C. & Wilson, S. 2016.
Climate-driven regime shift of a temperate marine ecosystem. *Science* 353:169-172.

Wickham, H. 2011. The Split-Apply-Combine Strategy for Data Analysis. *Journal of Statistical Software* 40(1):1-29.

Wiens, J. J. 2016. Climate-related local extinctions are already widespread among plant and animal species. *PLOS Biology* 14:1–18.

Wynne, M. J., Bradshaw, T. & Carrington, C. M. S. 2014. A checklist of the benthic marine algae of Barbados, West Indies. *Botanica Marina* 57(3):167-184.

Wynne, M. J. 2017. A checklist of benthic marine algae of the tropical and subtropical western Atlantic: fourth revision. *Nova Hedwigia Beiheft* 145:1-202.

Yan, W., Zhong, Y. & Shangguan, Z. 2017. Contrasting responses of leaf stomatal characteristics to climate change: a considerable challenge to predict carbon and water cycles. *Glob. Chang. Biol.* 23, 3781–3793.

Yip, Z. T., Quek, Z. B. R., Low, J. K. Y., Wilson, B., Bauman, A. G., Chou, L. M., Todd, P.A. & Huang, D. 2018. Diversity and phylogeny of *Sargassum* (Fucales, Phaeophyceae) inSingapore. *Phytotaxa* 369:200–10.

Yip, Z. T., Quek, Z. B. R. & Huang, D. 2020. Historical biogeography of the widespread macroalga *Sargassum* (Fucales, Phaeophyceae). *Journal of Phycology* 56:300-309.

Zarco-Perello, S., Wernberg, T., Langlois, T. J. & Vanderklift, M. A. 2017. Tropicalization strengthens consumer pressure on habitat-forming seaweeds. *Scientific Reports* 7(1):1-8.

Zarco-Perello, S., Bosch, N. E., Bennett, S., Vanderklift, M. A. & Wernberg, T. 2021. Persistence of tropical herbivores in temperate reefs constrains kelp resilience to cryptic habitats. *Journal of Ecology* 109(5):2081–2094.

List of Tables (Legends)

Table 1. Number of records used to produce niche distribution model for *Sargassum* species, values for mean test Area Under Receiver Operating Characteristic Curve (ROC-AUC), true skill statistics (TSS), fixed cumulative value 10 complementary log-log transformation threshold (T10), and the percentual relative contribution of the parameters temperature, salinity, depth, and ice to the model. We used TSS for those species with more than 35 records, for whom we used an ensemble approach. T10 was used for the species with few registers modeled with leave-one-out-cross-validation approach in Maxent only. The redder a cell, the greater the variable importance to the final model for each species.

Table 2. Suitability area, maximum North latitude (Max Lat North), variations northward (Δ North), maximum South latitude (Max Lat South), variation southward (Δ South), and overlap area between suitability occurrence in present and future scenarios for 13 benthic *Sargassum* species from the Western Atlantic Ocean. The percentage of the area and overlap area has the present projected scenario as reference. Latitudinal parameters are in degrees.

Table 1. Number of records used to produce niche distribution model for *Sargassum* species, values for mean test Area Under Receiver Operating Characteristic Curve (ROC-AUC), true skill statistics (TSS), fixed cumulative value 10 complementary log-log transformation threshold (T10), and the percentual relative contribution of the parameters temperature, salinity, depth, and ice to the model. We used TSS for those species with more than 35 records, for whom we used an ensemble approach. T10 was used for the species with few registers modeled with leave-one-out-cross-validation approach in Maxent only. The redder a cell, the greater the variable importance to the final model for each species.

Species	Records	Mean AUC (range)	Mean TSS/T10 (range)	Temperature	Salinity	Depth	lce
S. bermudense	10	0.83 (0.62 - 0.99)	0.36 (0.32 - 0.44)	99,5%	0,2%	0,1%	0,1%
S. buxifolium	21	0.86 (0.59 - 1.00)	0.33 (0.27 - 0.36)	95,0%	4,5%	0,2%	0,3%
S. cymosum	62	0.82 (0.78 - 0.85)	0.64 (0.61 - 0.67)	68,9%	14,0%	6,7%	6,2%
S. filipendula	183	0.86 (0.83 - 0.91)	0.69 (0.66 - 0.76)	66,4%	10,6%	11,1%	8,6%
S. furcatum	33	0.89 (0.70 - 1.00)	0.34 (0.31 - 0.35)	61,7%	36,2%	2,1%	0,0%
S. hystrix	142	0.83 (0.79 - 0.87)	0.66 (0.62 - 0.68)	56,2%	25,1%	6,8%	7,2%
S. platycarpum	43	0.85 (0.82 - 0.89)	0.71 (0.63 - 0.77)	62,1%	19,2%	8,1%	7,0%
S. polyceratium	90	0.90 (0.85 - 0.92)	0.76 (0.70 - 0.80)	61,3%	24,9%	5,3%	4,7%
S. pteropleuron	52	0.86 (0.84 - 0.89)	0.73 (0.65 - 0.77)	73,1%	12,3%	12,6%	1,2%
S. ramifolium	24	0.89 (0.70 - 1.00)	0.37 (0.33 - 0.40)	78,4%	21,4%	0,2%	0,0%
S. rigidulum	29	0.93 (0.74 - 1.00)	0.35 (0.31 - 0.37)	70,7%	26,3%	3,0%	0,0%
S. stenophyllum	29	0.97 (0.92 - 1.00)	0.17 (0.15 - 0.18)	61,6%	34,6%	3,8%	0,0%
Sargassum WA	404	0.86 (0.84 - 0.88)	0.63 (0.61 - 0.64)	59,8%	15,5%	5,1%	10,0%

Table 2. Suitability area, suitability area variation (Δ Area), maximum North latitude (Max Lat North), variations northward (Δ North), maximum South latitude (Max Lat South), variation southward (Δ South), and overlap area between suitability occurrence in present and future scenarios for 13 benthic *Sargassum* species from the Western Atlantic Ocean. The percentage of the area and overlap area has the present projected scenario as reference. Latitudinal parameters are in degrees.

Species	Scenario	Suitability Area (km²)	ΔArea (%)	Max Lat North	ΔNorth	Max Lat South	ΔSouth	Overlap Area (km²)	Overlap Area (%)	
Sargassum bermudense	present	2225197	0	41.84118	0	-40.22967	0	2225197	100	
	RCP 4.5	2341813	+ 5.4	47.25702	5.41584	-40.47963	-0.24996	2225197	100	
	RCP 8.5	2533400	+ 13.9	49.92328	8.08210	-41.22951	-0.99984	2225197	100	
Sargassum buxifolium	present	1678061	0	39.84148	0	-31.81428	0	1678061	100	
	RCP 4.5	1507977	- 10.1	41.34126	1.49978	-35.89699	-4.08271	1482665	88.4	
	RCP 8.5	743374	- 55.7	47.25702	7.41554	-36.89684	-5.08256	629083	37.5	
Sargassum cymosum	present	2017188	0	41.35021	0	-36.81171	0	2017188	100	
	RCP 4.5	1521753	- 24.6	41.85018	0.49997	-37.06169	-0.24998	1517677	75.2	
	RCP 8.5	699781	- 54.0	44.93333	3.58312	-40.06151	-3.24980	673056	44.5	
Sargassum filipendula	present	2082507	0	43.93339	0	-40.47815	0	2082507	100	
	RCP 4.5	2169444	+ 4.2	47.76649	3.83310	-40.64481	-0.16666	2082507	100	
	RCP 8.5	2284614	+ 9.7	50.09968	6.16629	-41.22810	-0.74995	2082507	100	
Sargassum furcatum	present	1463296	0	37.75847	0	-33.14741	0	1463296	100	
	RCP 4.5	1050691	- 28.2	40.59137	2.83290	-33.14741	0	1046927	71.5	
	RCP 8.5	1057763	- 37.7	41.42458	3.66611	-34.51864	-1.37123	1045386	71.4	
Sargassum hystrix	present	1612724	0	38.51705	0	-31.56202	0	1612724	100	
	RCP 4.5	1763806	+ 9.4	39.35033	0.83328	-33.14526	-1.58324	1612724	100	

	RCP 8.5	1858465	+ 15.2	40.18362	1.66657	-34.31186	-2.74984	1612724	100
Sargassum platycarpum	present	1503744	0	36.35052	0	-29.14551	0	1503744	100
	RCP 4.5	1696117	+ 12.8	38.68371	2.33319	-30.47876	-1.33325	1503744	100
	RCP 8.5	1826365	+ 21.5	39.76698	3.41646	-31.39537	-2.24986	1503744	100
Sargassum polyceratium	present	1313469	0	35.76722	0	-26.89564	0	1313469	100
	RCP 4.5	1607232	+ 22.4	37.68377	1.91655	-27.64560	-0.74996	1313469	100
	RCP 8.5	1721550	+ 31.1	38.93369	3.16647	-30.72874	-3.83310	1313469	100
Sargassum pteropleuron	present	663327	0	35.43391	0	-25.56239	0	663327	100
	RCP 4.5	1449037	+ 118.4	39.43366	3.99975	-26.56233	-0.99994	663327	100
	RCP 8.5	1776542	+ 167.8	40.10029	4.66638	-30.56209	-4.99970	663327	100
Sargassum ramifolium	present	1565947	0	39.34156	0	-31.39768	0	1565947	100
	RCP 4.5	1310462	- 16.3	40.75801	1.41645	-33.14741	-1.74973	1299848	83
	RCP 8.5	1030171	- 34.2	41.34126	1.99970	-34.31390	-2.91622	1001235	63.9
Sargassum rigidulum	present	1456138	0	40.59137	0	-31.39768	0	1456138	100
	RCP 4.5	1128950	- 22.5	41.42458	0.83321	-33.14741	-1.74973	1128087	77.5
	RCP 8.5	1109425	- 23.8	43.67423	3.08286	-36.89684	-5.49916	1102225	75.7
Sargassum stenophyllum	present	795816	0	39.42488	0	-32.23088	0	795816	100
	RCP 4.5	315013	- 60.4	40.17477	0.74989	-33.14741	-0.91653	305970	38.4
	RCP 8.5	157738	- 80.2	41.34126	1.91638	-36.23027	-3.99939	129060	18.5
Sargassum WA	present	1768857	0	42.43348	0	-37.64499	0	1768857	100
	RCP 4.5	1786798	+ 1.0	44.10005	1.66657	-37.81165	-0.16666	1768857	100
	RCP 8.5	1536880	- 13.1	48.76643	6.33295	-37.89497	-0.24998	1523227	86.1

List of Figures (Legends)

Figure 1. Relation between maximum latitude with suitability area in the present scenario for each Western Atlantic *Sargassum* species and the latitudinal variation predicted in our model results. Similar dots represent the same species in different projected scenarios. Leftmost dot represents the present scenario, the middle dot represents the RCP 4.5 scenario, and the farright dot represents RCP 8.5 scenario. (A) North latitude. (B) South latitude.

Figure 2. Habitat suitability model projections for 12 *Sargassum* species. Red areas refer to the suitability areas while blue areas refer to unsuitability areas for present (A, D, G, J, M, P, S, V, Y, B1, E1, H1), RCP 4.5 (B, E, H, K, N, Q, T, W, Z, C1, F1, I1) and RCP 8.5 (C, F, I, L, O, R, U, X, A1, D1, G1, J1) scenarios. Species tested: *S. bermudense* (A-C), *S. buxifolium* (D-F), *S. cymosum* (G-I), *S. filipendula* (J-L), *S. furcatum* (M-O), *S. hystrix* (P-R), *S. platycarpum* (S-U), *S. polyceratium* (V-X), *S. pteropleuron* (Y-A1), *S. ramifolium* (B1-D1), *S. rigidulum* (E1-G1), and *S. stenophyllum* (H1-J1).

Figure 3. Changes of future suitability area for Western Atlantic *Sargassum* species. Similar dots represent the same species in different projected scenarios. Dashed line represents the projected suitability area for the present scenario. Dots on the left side of the line represent a decrease in suitability area. Dots on the right side highlight an increase in the suitability area. Dots on the dashed line show the present scenario for each species. Dots next to the dashed line show the RCP 4.5 scenario and points further away represent the RCP 8.5 scenario, with exception of *S. furcatum* whose most distant point represents the RCP 4.5 scenario, and *Sargassum* WA whose the dot on the right side of the dashed line represents RCP 4.5 scenario and the dot on the left side represent the RCP 8.5 scenario.



A)

Maximum present North latitude (degrees)



Figure 1. Relation between maximum latitude with suitability area in the present scenario for each Western Atlantic *Sargassum* species and the latitudinal variation predicted in our model results. Similar dots represent the same species in different projected scenarios. Leftmost dot represents the present scenario, the middle dot represents the RCP 4.5 scenario, and the far-right dot represents RCP 8.5 scenario. (A) North latitude. (B) South latitude.

Figure 2. Habitat suitability model projections for 12 Sargassum species. Red areas refer to the suitability areas while blue areas refer to unsuitability areas for present (A, D, G, J, M, P, S, V, Y, B1, E1, H1), RCP 4.5 (B, E, H, K, N, Q, T, W, Z, C1, F1, I1) and RCP 8.5 (C, F, I, L, O, R, U, X, A1, D1, G1, J1) scenarios. Species tested: *S. bermudense* (A-C), *S. buxifolium* (D-F), *S. cymosum* (G-I), *S. filipendula* (J-L), *S. furcatum* (M-O), *S. hystrix* (P-R), *S. platycarpum* (S-U), *S. polyceratium* (V-X), *S. pteropleuron* (Y-A1), *S. ramifolium* (B1-D1), *S. rigidulum* (E1-G1), and *S. stenophyllum* (H1-J1).



























Figure 3. Changes of future suitability area for Western Atlantic *Sargassum* species. Similar dots represent the same species in different projected scenarios. Dashed line represents the projected suitability area for the present scenario. Dots on the left side of the line represent a decrease in suitability area. Dots on the right side highlight an increase in the suitability area. Dots on the dashed line show the present scenario for each species. Dots next to the dashed line show the RCP 4.5 scenario and points further away represent the RCP 8.5 scenario, with exception of *S. furcatum* whose most distant point represents the RCP 4.5 scenario, and *Sargassum* WA whose the dot on the right side of the dashed line represents RCP 4.5 scenario and the dot on the left side represent the RCP 8.5 scenario.

Supplementary Material

Table S1. Values of Area Under Receiver Operating Characteristic Curve (AUC) and true skill statistics (TSS) for each algorithm tested for seven Sargassum species. Models tested: generalized linear model (GLM), generalized additive model (GAM), boosted regression trees (BRT), random forests (RF), Bioclim, classification and regression trees (CART), support vector machine (SVM), artificial neural networks (ANN), maximum likelihood (Maxlike), maximum entropy (Maxent), multivariate adaptive regression spline (MARS).

Figure S1. Phylogenetic relationships of selected *Sargassum* species included in this study, following Camacho *et al.* (2015) and B. Barros-Barreto, M. T. Szechy & V. Cassano (unpublished data).

Figure S2. Variables histogram and their correlation used to fit ecological niche models for *Sargassum* species in the Western Atlantic Ocean.

Figure S3. Correlation between changes in suitability area and changes in latitudinal limits for 13 *Sargassum* species from the Western Atlantic Ocean. A & B: data from the northern hemisphere. C & D: data from the southern hemisphere. A & C: results from RCP 4.5. B & D: results from RCP 8.5. Dashed lines represent regression lines for better pattern visualization. Dots correspond to the same species as in Fig.3. Suitability area corresponds to the future suitability area in relation to the suitability area predicted for the present scenario for each species.

Figure S4. Suitability map for *Sargassum* WA predictions for (A) present, (B) RCP 4.5, and (C) RCP 8.5 scenarios.

Appendix S1 - Considerations of some unchosen model variables

Appendix S2 – Comments on research limitations

Appendix S3 – R scripts used in this study

Table S1. Values of Area Under Receiver Operating Characteristic Curve (AUC) and true skill statistics (TSS) for each algorithm tested for seven *Sargassum* species. Models tested: generalized linear model (GLM), generalized additive model (GAM), boosted regression trees (BRT), random forests (RF), Bioclim, classification and regression trees (CART), support vector machine (SVM), artificial neural networks (ANN), maximum likelihood (MAXLIKE), maximum entropy (MAXENT), and multivariate adaptive regression spline (MARS).

Species/Algorithms	GLM		GAM		BRT		RF		Bioc	lim	CAR	Т	SVM		ANN		MAX	LIKE	MAX	ENT	MAR	S
	AUC	TSS																				
S. cymosum	0.78	0.67	0.82	0.63	0.81	0.67	0.85	0.62	0.84	0.66	0.81	0.63	0.84	0.63	0.81	0.61	0.81	0.62	0.82	0.67	0.81	0.66
S. filipendula	0.83	0.68	0.87	0.72	0.86	0.67	0.91	0.73	0.79	0.60	0.86	0.67	0.89	0.76	0.86	0.66	0.83	0.67	0.86	0.68	0.85	0.66
S. hystrix	0.84	0.67	0.84	0.66	0.81	0.66	0.87	0.68	0.79	0.62	0.84	0.64	0.83	0.68	0.84	0.63	0.82	0.65	0.82	0.67	0.86	0.67
S. platycarpum	0.84	0.73	0.82	0.72	0.89	0.77	0.87	0.77	0.82	0.63	0.84	0.65	0.87	0.75	0.84	0.65	0,84	0.69	0.85	0.77	0.78	0.59
S. polyceratium	0.90	0.77	0.92	0.77	0.90	0.76	0.92	0.80	0.87	0.73	0.90	0.74	0.93	0.80	0.89	0.72	0.85	0.70	0.90	0.78	0.88	0.77
S. pteropleuron	0.88	0.77	NA	NA	0.87	0.75	0.89	0.76	0.74	0.50	0.81	0.60	0.83	0.68	0.82	0.66	0.77	0.55	0.89	0.77	0.84	0.65
Sargassum WA	0.79	0.57	0.85	0.62	0.85	0.60	0.88	0.63	0.74	0.55	NA	NA	0.86	0.64	0.84	0.58	0.78	0.57	0.84	0.61	0.85	0.63



Figure S1. Phylogenetic relationships of selected *Sargassum* species included in this study, following Camacho et al. (2015) and B. Barros-Barreto, M. T. Szechy and V. Cassano (unpublished data)



Figure S2. Variables histogram and their correlation used to fit ecological niche models for *Sargassum* species in the Western Atlantic Ocean.



Figure S3. Correlation between changes in suitability area and changes in latitudinal limits for 13 *Sargassum* species from the Western Atlantic Ocean. A & B: data from the northern hemisphere. C & D: data from the southern hemisphere. A & C: results from RCP 4.5. B & D: results from RCP 8.5. Dashed lines represent regression lines for better pattern visualization. Dots correspond to the same species as in Figure 3. Suitability area corresponds to the future suitability area in relation to the suitability area predicted for the present scenario for each species.



Figure S4. Suitability map for *Sargassum* WA predictions for (A) present, (B) RCP 4.5, and (C) RCP 8.5 scenarios.

Appendix S1 - Considerations of some unchosen model variables

We choose not to use other variables, such as light and nutrient measures. Light is usually correlated with cloud cover and, indirectly, depth. But we aren't able to predict future scenarios for light intensity. We know photon flux density (PFD) requirements for WA benthic *Sargassum* are usually low, and light compensation occurred at reduced intensities when the temperature dropped (Carpenter and Cox 1974, Hanisak and Samuel 1987), possibly not being a limiting parameter in lower depths. However, it must be considered that light is an important factor to consider when we search for deep-water refugia (Graham et al. 2007). Refugia could aid species with lower temperatures requirements to survive in tropical latitudes at higher depths (Wiens 2016). *Sargassum obtusifolium* was found in 200 m depth in Hawaii (De Wreed and Jones 1973, as *S. hawaiensis*).

Also, we cannot predict future scenarios for nitrogen nor phosphorus concentrations in coastal areas, as urban development can rapidly change these nutrients input. Physiological studies with pelagic *Sargassum natans* and *S. fluitans* species found an increase in growth and photosynthetic rate with phosphate enrichment, but not with nitrate or ammonium addiction (Lapointe 1986). Nitrogen and phosphorus concentrations in the North Atlantic Ocean were low to indetectable. However, nitrogen-fixer epiphite *Dichothrix fucicola* usually were found in pelagic *Sargassum* (Carpenter 1972) and *Sargassum* patches interstitial water has 2-3 times more reactive phosphorus concentrations than in surrounding areas (Culliney 1970). Otherwise, recent studies with brown algae have found that neither nitrogen nor phosphorus are important variables in SDM models when one think in the distribution limits of a species. The algae can grow with low nutrient concentrations, but at a slower rate (Franco et al. 2018). Nutrients, however, can increase the upper limit temperature of *Sargassum polycystum* (Zou

et al. 2017). Nutrients are also important variables for predicting abundance and biomass, helping to forecast algae blooms (Lapointe 1986, Brooks et al. 2018, Jouanno et al. 2021).

References

Brooks, M. T., Coles, V. J., Hood, R. R. & Gower, J. F. 2018. Factors controlling the seasonal distribution of pelagic *Sargassum*. *Mar. Ecol.-Prog. Ser.*, 599:1–18.

Carpenter, E. J., 1972. Nitrogen fixation by a blue-green epiphyte on pelagic *Sargassum*. *Science* 178:1207-1209.

Carpenter, E. J. & Cox, J. L. 1974. Production of pelagic *Sargassum* and a blue-green epiphyte in the western Sargasso Sea. Limnology and Oceanography. 19(3):429-436.

Culliney, J. L. 1970. Measurements of reactive phosphorus associated with pelagic *Sargassum* the north Sargasso Sea. *Limnol. Oceonogr.* 15:304-306.

De Wreede, R. E. & Jones, E. C. 1973. New records of *Sargassum hawaiiensis* Doty and Newhouse (Sargassaceae, Phaeophyta), a deep water species. *Phycologia* 12(1/2):59-62.

Franco, J., Bertocci, I., Rodriguez, L., Martinez, B., Sousa Pinto, I. & Arenas, F. 2018. The 'golden kelp' *Laminaria ochroleuca* under global change: Integrating multiple eco-physiological responses with species distribution models. *Journal of Ecology* 106:47-58.

Graham, M. G., Kinlan, B. P., Druehl, L. D., Garske, L. E. & Banks, S. 2007. Deep-water kelp refugia as potential hotspots of tropical marine diversity and productivity. *Proceedings of the National Academy of Sciences of the United States of America* 104(42):16576-16580.

Hanisak, M. D. & Samuel, M. A. 1987. Growth rates in culture of several species of *Sargassum* from Florida, USA. *Hydrobiologia* 151/152:399-404.

Jouanno, J., Benshila, R., Berline, L., Soulié, A., Radenac, M.-E., Morvan, G., Diaz, F., Sheinbaum, J., Chevalier, C., Thibaut, T., Changeux, T., Ménard, F., Berthet, S., Aumont, O., Ethé, C., Nabat, P. & Mallet, M. 2021. A NEMO-based model of *Sargassum* distribution in the tropical Atlantic: Description of the model and sensitivity analysis (NEMO-Sarg1.0). *Geoscientific Model Development* 14:4069-4086.

Lapointe, B. E. 1986. Phosphorus-limited photosynthesis and growth of *Sargassum natans* and *Sargassum fluitans* (Phaeophyceae) in the western North Atlantic. *Deep-Sea Res. Pt. A* 33:391–399.

Wiens, J. J. 2016. Climate-related local extinctions are already widespread among plant and animal species. *PLOS Biology* 14:1–18.

Zou, X. -X., Xing, S.-S., Su, X., Zhu, J., Huang, H.-Q. & Bao, S.-X. 2018. The effects of temperature, salinity and irradiance upon the growth of *Sargassum polycystum* C. Agardh (Phaeophyceae). *J. Appl. Phycol.* 30:1207–1215.

Appendix S2 – Comments on research limitations

In temperate regions, temperature seasonal variations are more pronounced and directly influence benthic composition. The perennial species in temperate environments – or at least species with perennial holdfast – have been adapted to seasonal physical changes (Hällfors et al. 2016). These variations fit the species' realized niche and must be considered when modeling temperate species. Tropical species, however, evolved in an environment that is climatically more stable, and are hence strongly driven by ecological interactions, microclimate and non-climate driven environmental heterogeneity such as changes in substrate characteristics (Brown 2014).

Nutrients have synergistic effects with CO₂ concentration on seaweed growth, settlement and photosynthesis (Russell et al. 2009). The influence of different nutrient availability and concentration on habitat modeling, such as iron, nitrogen, and phosphorus average concentration have been studied in *Sargassum* and other algae (Miki et al. 2016, Wenke 2019). Iron as a fertilizer eluate, and iron-binding ethylenediaminetetraacetic acid (Fe-EDTA), promote the growth of germlings and immature stages of *Sargassum horneri* in Japan (Miki et al. 2016). Iron depositions are also related to recent accumulation as of drifting pelagic *Sargassum* on Caribbean beaches (Oviatt et al. 2019). Nitrogenase protein contains iron, so iron concentrations act in nitrogen fixation regulation (Wenke 2019). The development of present day and future nutrient rasters could improve *Sargassum* niche distribution models.

Dispersion is an aspect we cannot predict using the ENM approach. Changes in macroalgae suitability area may be faster than dispersion or migration rates towards new suitable habitats, potentially leading to local extinctions (Wiens 2016). Local extinctions due to the inability of a species to move into newly suitable areas has been documented for *Fucus*

vesiculosus in the Baltic Sea (Jonsson et al. 2018). Local extinctions for other large Fucales have also been observed with *Sargassum* and *Cystoseira* species in the Mediterranean Sea (Thibaut et al. 2005, 2015). WA *Sargassum* usually have air bladders or aerocysts (Paula & Eston 1987, Camacho et al. 2015), and some species like *S. hystrix* and *S. platycarpum* were found floating along the Brazil Current, 100 km offshore (Oliveira-Filho et al. 1979). The effects of the Amazon discharge as a major dispersal barrier are also unknown for *Sargassum* species and it may hinder or even interrupt dispersion from and to the South Atlantic (Floeter et al. 2008). In fact, our present suitability areas suggest the influence of the Amazon River as a barrier to dispersal. *Sargassum* species only known to occur in Northern latitudes (e.g., *S. bermudense* and *S. pteropleuron*) have similar environments in the Southern Hemisphere. Without dispersion, macroalgae survival will depend solely on their capacity to acclimate or to shift their realized niche (Wiens 2016). To work around this issue, physiological experiments can improve the ENM predictions (Ready et al. 2010).

Ecological data like density and abundance may be helpful to foresee some ecosystems services (Gouvêa et al. 2020), and density of predators, epibionts colonization, assemblage resilience, and reproductive seasonality may improve predictions for species and whole communities (Lima 2007, Ling & Johnson 2009, Peck et al. 2018, Kvile et al. 2022).

References

Brown, J. H. 2014. Why are there so many species in the tropics? *Journal of Biogeography* 41:8–22.

Camacho, O., Mattio, L., Draisma, S., Fredericq, S. & Diaz-Pulido, G. 2015. Morphological and molecular assessment of *Sargassum* (Fucales, Phaeophyceae) from Caribbean Colombia,

including the proposal of *Sargassum giganteum* sp. nov., *Sargassum schnetteri* comb. nov. and *Sargassum* section *Cladophyllum* sect. nov. *Systematics and Biodiversity* 13(2):105-130.

Floeter, S. R., Rocha, L. A., Robertson, D. R., Joyeux, J. C., Smith-Vaniz, W. F., Wirtz, P., Edwards, A. J., Barreiros, J. P., Ferreira, C. E. L., Gasparini, J. L., Brito, A., Falcón, J. M., Bowen, B. W. & Bernardi, G. 2008. Atlantic reef fish biogeography and evolution. *Journal of Biogeography* 35:22–47.

Gouvêa, L. P., Assis, J., Gurgel, C. F. D., Serrão, E. A., Silveira, T. C. L., Santos, R., Duarte,
C. M., Peres, L. M. C., Carvalho, V. F., Batista, M., Bastos, E., Sissini, M. N. & Horta, P. A.
2020. Golden carbon of *Sargassum* forests revealed as an opportunity for climate change mitigation. *Science of The Total Environment* 729:138745 pp.1-10.

Hällfors, M. H., Liao, J., Dzurisin, J., Grundel, R., Hyvärinen, M., Towle, K., Wu, G. & Hellmann, J. J. 2016. Addressing potential local adaptation in species distribution models: Implications for conservation under climate change. *Ecological Applications* 26:1154–1169.

Jonsson, P. R., Kotta, J., Andersson, H. C., Herkül, K., Virtanen, E., Sandman, A. N. & Johannesson, K. 2018. High climate velocity and population fragmentation may constrain climate-driven range shift of the key habitat former *Fucus vesiculosus*. *Diversity and Distributions* 24:892–905.

Kvile, K. Ø., Andersen, G. S., Baden, S. P., Bekkby, T., Bruhn, A. Geertz-Hansen, O., Hancke, K. Hansen, J. L. S., Krause-Jensen, D., Rinde, E., Steen, H., Wegeberg, S. &
Gundersen, H. 2022. Kelp Forest Distribution in the Nordic Region. *Frontiers in Marine Science* 9:850359.

Lima, F. P. 2007. Biogeography of Benthic Invertebrate Assemblages on the Portuguese Rocky Coast: Relation with Climatic and Oceanographic Patterns. Ph.D. thesis, Faculdade de Ciências da Universidade do Porto, Porto, Portugal, 235 pp.

Ling, S. D. & Johnson, C. R. 2009. Population dynamics of an ecologically important rangeextender: kelp beds versus sea urchin barrens. *Mar. Ecol. Prog. Ser.* 374: 113–125.

Miki, O., Nagai, T., Marzuki, M., Okumura, C., Kosugi, C. & Kato, T. 2016. Effects of Fe fertilizer eluate on the growth of *Sargassum horneri* at the germling and immature stages. *J. Appl. Phycol.* 28:1775–1782.

Oliveira-Filho, E. C., Ugadim, Y. & Paula, E. J. 1979. Comunidades Associadas a Plantas de *Sargassum* Flutuantes em Águas da Corrente do Brasil - Considerações Biogeográficas. *Boletim de Botânica da Universidade de São Paulo* 7:5-9.

Oviatt, C. A., Huizenga, K., Rogers, C. S. & Miller, W. J. 2019. What nutrient sources support anomalous growth and the recent *Sargassum* mass stranding on Caribbean beaches? A review. *Mar. Pollut. Bull.* 145:517–525.

Paula, E. & Eston, V. 1987. Are There Other *Sargassum* Species Potentially as Invasive as *S. muticum? Botanica Marina* 30(5):405-410.

Peck, M. A., Arvanitidis, C., Butenschön, M., Canu, D. M., Chatzinikolaou, E., Cucco, A., Domenici, P., Fernandes, J. A., Gasche, L., Huebert, K. B., Hufnagl, M., Jones, M. C., Kempf, A., Keyl, F., Maar, M., Mahévas, S., Marchal, P., Nicolas, D., Pinnegar, J. K., Rivot, E., Rochette, S., Sell, A. F., Sinerchia, M., Solidoro, C., Somerfield, P. J., Teal, L. R., Travers-Trolet, M. & Van De Wolfshaar, K. E. (2018). Projecting changes in the distribution and productivity of living marine resources: A critical review of the suite of modelling approaches used in the large European project VECTORS. *Estuarine Coastal and Shelf Science* 201:40-55

Ready, J., Kaschner, K., South, A. B., Eastwood, P. D., Rees, T., Rius, J., Agbayani, E., Kullander, S. & Froese, R. 2010. Predicting the distributions of marine organisms at the global scale. *Ecological Modelling* 221(3):467–478.

Russell, B. D., Thompson, J. A. I., Falkenberg, L. J. & Connell, S. D. 2009. Synergistic effects of climate change and local stressors: CO2, and nutrient-driven change in subtidal rocky habitats. *Global Change Biol.* 15:2153–2162.

Thibaut, T., Pinedo, S., Torras, X. & Ballesteros, E. 2005. Long-term decline of the populations of Fucales (*Cystoseira* spp. and *Sargassum* spp.) in the Alberes coast (France, North-western Mediterranean). Marine Pollution Bulletin 50:1472-1489.

Thibaut, T., Blanfuné, A., Boudouresque, C. F. & Verlaque, M. 2015. Decline and local extinction of Fucales in the French riviera: the harbinger of future extinctions? *Mediterr. Mar. Sci.* 16(1):206–224.

Wenke, B. B. 2019 The Many Roles of the Nitrogenase Iron Protein. Ph.D. dissertation. California Institute of Technology, Pasadena, California, 108pp.

Wiens, J. J. 2016. Climate-related local extinctions are already widespread among plant and animal species. *PLOS Biology* 14:1–18.

Appendix S3 - R scripts used in this study

Calling packages

if(!require(dismo)){install.packages('dismo'); library(dismo)} if(!require(rgeos)){install.packages('rgeos'); library(rgeos)} if(!require(raster)){install.packages('raster'); library(raster)} if(!require(rgdal)){install.packages('rgdal'); library(rgdal)} if(!require(leaflet)){install.packages('leaflet'); library(leaflet)} if(!require(sdmpredictors)){install.packages('sdmpredictors'); library(sdmpredictors)} if(!require(SDMTools)){install.packages('SDMTools'); library(SDMTools)} if(!require(sdm)){install.packages('sdm'); library(sdm)} if(!require(usdm)){install.packages('usdm'); library(usdm)} if(!require(dplyr)){install.packages('dplyr'); library(dplyr)} my.colors = colorRampPalette(c("#5E85B8","#EDF0C0","#C13127"))

Downloading layers of interest
Present
prof <- load_layers("BO_bathymin") ## depth
temp <- load_layers("BO2_tempmax_bdmin") ## temperature
salinidade <- load_layers("BO2_salinitymean_bdmin") ## salinity
gelo <- load_layers("BO2_icethickmean_ss") ## ice</pre>

RCP 4.5 2100
temp.rcp45.2100 <- load_layers("BO2_RCP45_2100_tempmax_bdmin")
sal.rcp45.2100 <- load_layers("BO2_RCP45_2100_salinitymean_bdmin")
gelo.rcp45.2100 <- load_layers("BO2_RCP45_2100_icethickmean_ss")</pre>

RCP 8.5 2100
temp.rcp85.2100 <- load_layers("BO2_RCP85_2100_tempmax_bdmin")
sal.rcp85.2100 <- load_layers("BO2_RCP85_2100_salinitymean_bdmin")
gelo.rcp85.2100 <- load_layers("BO2_RCP85_2100_icethickmean_ss")</pre>

Writing rasters on pc
writeRaster (prof, "prof.tif") ## do the same with the others

Calling rasters after mask with WA extent provided in QGIS software
Present
prof <- raster("prof mask.tif") #depth</pre>

temp <- raster("temp_mask.tif")
salinidade <- raster("salinidade_mask.tif")
gelo <- raster("gelo_mask.tif")</pre>

#temperature #salinity #ice

RCP 4.5 2100 temp.rcp45.2100 <- raster("temp.rcp45.2100_mask.tif") sal.rcp45.2100 <- raster("sal.rcp45.2100_mask.tif") gelo.rcp45.2100 <- raster("gelo.rcp45.2100_mask.tif")

RCP 8.5 2100
temp.rcp85.2100 <- raster("temp.rcp85.2100_mask.tif")
sal.rcp85.2100 <- raster("sal.rcp85.2100_mask.tif")
gelo.rcp85.2100 <- raster("gelo.rcp85.2100_mask.tif")</pre>

Creating extent of 26m depth WA = prof WA [WA>=(-265)] =1 WA [WA<(-265)]= NA plot (WA, col="blue", legend=F)

Creating stack with environmental layers
varbio.stack.presente <- stack (gelo, prof, salinidade, temp)
Masking environmental layers with depth 265m layer
varbio.m <- mask (varbio.stack.presente, WA)
Excluding NA's cells (depth > 265m)
varbio.m <- trim (varbio.m)</pre>

```
varbio.stack.rcp45.2100 <- stack (gelo.rcp45.2100, prof, sal.rcp45.2100,
temp.rcp45.2100)
varbio.m.rcp45.2100 <- mask (varbio.stack.rcp45.2100, WA)
varbio.m.rcp45.2100 <- trim (varbio.m.rcp45.2100)
names(varbio.m.rcp45.2100) =c("gelo", "profundidade", "salinidade", "temperatura")
```

varbio.stack.rcp85.2100 <- stack (gelo.rcp85.2100, prof, sal.rcp85.2100, temp.rcp85.2100)

```
varbio.m.rcp85.2100 <- mask (varbio.stack.rcp85.2100 , WA)
varbio.m.rcp85.2100 <- trim (varbio.m.rcp85.2100)
names(varbio.m.rcp85.2100) = c("gelo", "profundidade", "salinidade", "temperatura")</pre>
```

```
### Checking correlation between variables
varbio.val <- getValues(varbio.m)</pre>
                                                ## extracting values
varbio.coord <- coordinates (varbio.m)
                                                ## extraindo coordinate
varbio.j <- cbind (varbio.coord, varbio.val)</pre>
varbio.df <- na.omit (varbio.j)</pre>
write.table(varbio.df.presente, "bioclimatica.csv")
var.cor <- cor (varbio.df)</pre>
var.cor
### Potting (Figure S2)
names (varbio.m) = c ("ice", "depth", "salinity", "temperature")
panel.hist <- function(x, ...)
 usr <- par("usr"); on.exit(par(usr))</pre>
 par(usr = c(usr[1:2], 0, 1.5))
 h \le hist(x, plot = FALSE)
 breaks <- h$breaks;
 nB <- length(breaks)
 y \le h$counts; y \le y/max(y)
 rect(breaks[-nB], 0, breaks[-1], y, col="cyan", ...)
}
panel.cor <- function(x, y, digits=2, prefix="", cex.cor, ...)</pre>
 usr <- par("usr"); on.exit(par(usr))
 par(usr = c(0, 1, 0, 1))
 r \le abs(cor(x, y))
 txt \leq format(c(r, 0.2), digits = digits)[1]
 txt <- paste(prefix, txt, sep="")
 if(missing(cex.cor)) cex.cor <- 0.8/strwidth(txt)
 text(0.5, 0.5, txt, cex = 1)
}
```

Figure S2 pairs(varbio.m, panel=panel.smooth, diag.panel=panel.hist, lower.panel=panel.cor)

```
### Calling table (without duplicate registers)
o <- read.table ("Scymosum_limpo_preview.csv", head=T, dec=".", sep=",")
lat.lon01 <- cbind (o$lon, o$lat)  ## latitude and longitude coordinates
pts01 <-SpatialPoints(lat.lon01)</pre>
```

```
### Extracting values
varbio.model <- varbio.df #[, c("x", "y", "gelo", "prof", "sal", "temp")]</pre>
```

Checking if the points generate makes biological features
plot (varbio.m[[2]], legend=F, main="Presença e background")
points (backtotal[,1:2], add=T, col="firebrick", cex=0.5, pch=16)
points (clim3.pts01, add=T, pch=16, cex=0.5, col="blue")

write.table(backtotal, "background.txt")

```
### Saving background and presence in the same object
### Add 1 for presence and 0 for pseudo-absence
pts.presente <- clim2.pts01[,1:2]  # presence
pts.presente <- as.data.frame(pts.presente)
colnames(pts.presente) <- c("x", "y")
pts.presente$pb <- rep(1, nrow(pts.presente))</pre>
```

```
backtotal.presente <- backtotal[,1:2]  # pseudo-absence
backtotal.presente <- as.data.frame(backtotal.presente)
backtotal.presente$pb <- rep(0,nrow(pts.presente))
str(backtotal.presente)
```

```
data.presente <- rbind(pts.presente, backtotal.presente)
r2 <- SpatialPointsDataFrame(data.presente[,1:2], data.presente)
```

```
### Preparing objects
preds = varbio.m
dados1 = clim2.pts01
names(preds) <- c("gelo", "profundidade", "salinidade", "temperatura")
profundidade = prof
temperatura = temp
### ENM
d \le sdm::sdmData(formula=pb \sim gelo + profundidade + salinidade + temperatura,
      train=r2,
      predictors=preds)
d
model1<-sdm(pb~gelo+profundidade+salinidade+temperatura,
      data=d.
      #methods=c('glm', 'gam', "brt", "rf", "bioclim", "cart", "fda", nnetMLP
      #"svm","rpart", "mda", "maxlike", "maxent", "mars", "glmnet", "bioclimD"),
      methods=c('glm', 'gam', "brt", "rf", "bioclimD", "cart",
              "svm","rpart", "maxlike", "maxent", "mars"),
      replications='sub', test.percent=25, n=10)
model1
roc(model1, smooth=T)
write.sdm(model1,"model.presente Scymosum.sdm")
### Viewing model informations
getModelInfo(model1)
v1<- getVarImp(model1, id=1, wtest='test.dep')
                                                        ## variables importance
v1
plot(v1)
### Predicting
p2.presente <- predict(model1,newdata=preds, type='response',
              overwrite=T)
outfile2 <- writeRaster(p2.presente, filename='pred.presente.scymosum.tif',
                      format="GTiff", overwrite=TRUE,
```

```
options=c("INTERLEAVE=BAND","COMPRESS=LZW"))
str(p2.presente)
```

Accessing AUC and TSS AUCS_model1<-getEvaluation(model1,stat='AUC',wtest='test.dep', opt=1) AUCS_model1 TSS_model1 <- getEvaluation(model1,stat='tss',wtest='test.dep', opt=1) TSS_model1

```
### Checking specific algorithm parameter
p1<- p2.presente
sd.glm<-calc(p1[[1:10]], fun=sd)
sd.glm<-scale(sd.glm)
plot(sd.glm,col=my.colors(1000),main="GLM - VARIAÇÃO")
```

```
### Ensemble forecasting using AUC as parameter (example)
w<-AUCS_model2[1:10,2]
w<-w/sum(w)
ens.glm<- p1[[1]]*w[1]+p1[[2]]*w[2]+p1[[3]]*w[3]+p1[[4]]*w[4]+p1[[5]]*w[5]+
p1[[6]]*w[6]+p1[[7]]*w[7]+p1[[8]]*w[8]+p1[[9]]*w[9]+p1[[10]]*w[10]
plot(ens.glm,col=my.colors(1000), main = "ENSEMBLE - GLM")</pre>
```

```
plot (ensemble.tss, col=my.colors(1000))
points(clim2.pts01, col="dark green", pch=16, cex=0.5)
```

Binarizing

```
ensemble.tss.presente <- ensemble.tss
ensemble.tss.presente[ensemble.tss.presente>=0.5]=1
ensemble.tss.presente[ensemble.tss.presente<0.5]=0
plot (ensemble.tss.presente, col=my.colors(1000))
points(clim2.pts01, col="dark green", pch=16, cex=0.5)
```

writeRaster (ensemble.tss.presente, "ensemble.tss.presente_Scymosum.tif")

```
### Accessing coordinates of binarized ensemble
ensemble.bin.1 <- ensemble.tss.presente
ensemble.bin.1 [ensemble.bin.1>=1]=1
ensemble.bin.1 [ensemble.bin.1<0.5]=NA
ensemble.bin.1 <- trim(ensemble.bin.1)
ensemble.bin.1
table (getValues(ensemble.bin.1))  # number of cells occupied</pre>
```

Area cell size1<-area(ensemble.bin.1, na.rm=TRUE, weights=F) cell size1<-cell size1[!is.na(cell size1)] raster area.presente<- sum (cell size1[1:length(cell size1)]) raster area.presente ### Intersection with present occurrence (overlap) overlap.pres <- intersect(ensemble.bin.1, ensemble.tss.presente) $x \leq brick(resample(ensemble.bin.1, overlap.pres, method = "ngb"),$ resample(ensemble.tss.presente, overlap.pres, method = "ngb")) plot(ensemble.tss.presente, col = "blue") image(ensemble.bin.1, add = TRUE, col = "green") image(x, add = TRUE, col = "red")sum(values(area(ensemble.bin.1))) ## suitability area ##### Prediction for RCP 4.5 2010 scenario p.rcp.45.2100 <- predict(model1,newdata=varbio.m.rcp45.2100, type='response', overwrite=T) outfile3 <- writeRaster(p.rcp.45.2100, filename='p.rcp.45.2100 Scymosum.tif', format="GTiff", overwrite=TRUE, options=c("INTERLEAVE=BAND","COMPRESS=LZW")) str(p.rcp.45.2100) ensemble.tss.rcp45 <- sdm::ensemble (model1, newdata=varbio.m.rcp45.2100, filename='ensemble.full.tss.rcp45.Scymosum.img', setting=list(method='weighted',stat='tss'), overwrite=T) writeRaster(ensemble.tss.rcp45,"ensemble.rcp45 Scymosum.tif") ### Binarize ensemble ensemble.tss.rcp.45.bin <- ensemble.tss.rcp45 ensemble.tss.rcp.45.bin[ensemble.tss.rcp.45.bin>=0.5]=1 ensemble.tss.rcp.45.bin[ensemble.tss.rcp.45.bin<0.5]=0 writeRaster (ensemble.tss.rcp.45.bin, "ensemble.tss.bin.rcp45 Scymosum.tif")

Accessing coordinates of binarized ensemble ensemble.bin.2 <- ensemble.tss.rcp.45.bin ensemble.bin.2 [ensemble.bin.2>=1]=1 ensemble.bin.2 [ensemble.bin.2<0.5]=NA ensemble.bin.2 <- trim(ensemble.bin.2) ensemble.bin.2 table (getValues(ensemble.bin.2))

plot(ensemble.tss.presente, col = "blue", legend=F) image(ensemble.bin.2, add = TRUE, col = "red") image(overlap.pres, add = TRUE, col = "green")

```
### Area - get sizes of all cells in raster [km2]
cell_size.rcp45<-area(ensemble.bin.2, na.rm=TRUE, weights=F)
### delete NAs from vector of all raster cells
cell_size.rcp45<-cell_size.rcp45[!is.na(cell_size.rcp45)]
### compute area [km2] of all cells in geo_raster
raster_area.rcp.45<- sum (cell_size.rcp45[1:length(cell_size.rcp45)])
raster_area.rcp.45
```

```
### Overlapping Area
cell_size3<-area(overlap.pres, na.rm=TRUE, weights=F)
cell_size3<-cell_size3[!is.na(cell_size3)]
raster_area.overlap.rcp45 <- sum (cell_size3[1:length(cell_size3)])
raster_area.overlap.rcp45</pre>
```

overwrite=T)

writeRaster(ensemble.tss.rcp85,"ensemble.rcp85 Scymosum.tif")

```
### Binarize ensemble
ensemble.tss.rcp.85.bin <- ensemble.tss.rcp85
ensemble.tss.rcp.85.bin[ensemble.tss.rcp.85.bin>=0.5]=1
ensemble.tss.rcp.85.bin[ensemble.tss.rcp.85.bin<0.5]=0
plot (ensemble.tss.rcp.85.bin, legend=F, col=my.colors(1000))
#ensemble.bin.tss.rcp85_Scymosum.png
```

writeRaster (ensemble.tss.rcp.85.bin, "ensemble.tss.bin.rcp85_Scymosum.tif")

```
### Accessing coordinates of binarized ensemble
ensemble.bin.3 <- ensemble.tss.rcp.85.bin
ensemble.bin.3 [ensemble.bin.3>=1]=1
ensemble.bin.3 [ensemble.bin.3<0.5]=NA
ensemble.bin.3 <- trim(ensemble.bin.3)
ensemble.bin.3
table (getValues(ensemble.bin.3))</pre>
```

```
plot(ensemble.tss.presente, col = "blue", legend=F)
image(ensemble.bin.3, add = TRUE, col = "red")
image (overlap.rcp85, col="green", add=T)
```

```
### Area - get sizes of all cells in raster [km2]
cell_size.rcp85<-area(ensemble.bin.3, na.rm=TRUE, weights=F)
cell_size.rcp85<-cell_size.rcp85[!is.na(cell_size.rcp85)]
raster_area.rcp.85 <- sum (cell_size.rcp85[1:length(cell_size.rcp85)])
raster_area.rcp.85
```

```
### Overlapping percentage
overlap.total.rcp85 = raster_area.overlap.rcp85 / raster_area.presente
overlap.total.rcp85
```

Reading object with Fixed cumulative value 10 Cloglog threshold values t10 <- read.table ("t10.csv", sep=";", head=T) t10 <- t10[,2]</pre>

Calling rasters created with Maxent.jar ### Numbers of rasters equal number of observations/presence {r01.bin <- raster("Sargassum bermudense 0 layers presente 4v.asc") $r01.bin[r01.bin \le t10[1]] = 0$ r01.bin[r01.bin > t10[1]] = 1{r02.bin <- raster("Sargassum bermudense 1 layers presente 4v.asc") $r02.bin[r01.bin \le t10[2]] = 0$ r02.bin[r01.bin > t10[2]] = 1{r03.bin <- raster("Sargassum bermudense 2 layers presente 4v.asc") $r03.bin[r03.bin \le t10[3]] = 0$ r03.bin[r03.bin > t10[3]] = 1{r04.bin <- raster("Sargassum bermudense 3 layers presente 4v.asc") $r04.bin[r04.bin \le t10[4]] = 0$ r04.bin[r04.bin > t10[4]] = 1{r05.bin <- raster("Sargassum bermudense 4 layers presente 4v.asc") $r05.bin[r05.bin \le t10[5]] = 0$ r05.bin[r05.bin > t10[5]] = 1{r06.bin <- raster("Sargassum bermudense 5 layers presente 4v.asc") $r06.bin[r06.bin \le t10[6]] = 0$ r06.bin[r06.bin > t10[6]] = 1{r07.bin <- raster("Sargassum bermudense 6 layers presente 4v.asc") $r07.bin[r07.bin \le t10[7]] = 0$ r07.bin[r07.bin > t10[7]] = 1{r08.bin <- raster("Sargassum bermudense 7 layers presente 4v.asc") $r08.bin[r08.bin \le t10[8]] = 0$ r08.bin[r08.bin > t10[8]] = 1{r09.bin <- raster("Sargassum bermudense 8 layers presente 4v.asc") $r09.bin[r09.bin \le t10[9]] = 0$ r09.bin[r09.bin > t10[9]] = 1{r10.bin <- raster("Sargassum bermudense 9 layers presente 4v.asc") $r10.bin[r10.bin \le t10[10]] = 0$ r10.bin[r10.bin > t10[10]] = 1{r11.bin <- raster("Sargassum bermudense 10 layers presente 4v.asc") $r11.bin[r11.bin \le t10[11]] = 0$ r11.bin[r11.bin > t10[11]] = 1

Ensemble ensemble.p4v <- sum (r01.bin, r02.bin, r03.bin, r04.bin, r05.bin, r06.bin, r07.bin, r08.bin, r09.bin, r10.bin, r11.bin)

```
### Binarize ensemble
ensemble.p4v [ensemble.p4v < nrow(points)/2] = 0
ensemble.p4v [ensemble.p4v >= nrow(points)/2] = 1
```

writeRaster (ensemble.p4v, "ensemble.bin.t10.presente_Sbermudense_maxent_default.tif")

```
### Read results table
    results <- read.table ("Resultados.csv", head=T, dec=".", sep=",")
    colnames(results) = c("Species", "Scenario", "Area", "Perc.Area", "S", "dS", "N",
"dN", "Overlap Area", "Perc.Overlap")</pre>
```

```
results <- arrange (results, Species)
attach(results)
spp = (Species[Scenario == "present"])
```

```
###### Figure 3 - Changes in future suitability area
       windows()
       legcolors = c("black", "darkblue", "darkmagenta", "aquamarine4", "firebrick",
"brown4", "darkslategrey", "blue", "green", "red", "orange", "purple", "yellow4")
       plot ((Area[Scenario == "present"]/10^6) ~ Perc.Area[Scenario == "present"],
              ylab = "Present suitability area (10^{6} \text{ km}^2)",
              xlab= "Percentage of present suitability area (%)",
              pch=c(1:2,4:11, 3, 12:13),
              col=legcolors,
              xlim=c(0,300))
       par(new=TRUE)
       plot ((Area[Scenario == "present"]/10^6) ~ Perc.Area[Scenario == "RCP 4.5"],
              pch=c(1:2,4:11, 3, 12:13),
              col=legcolors,
              xlim=c(0.300))
       par(new=TRUE)
       plot ((Area[Scenario == "present"])/10^6 ~ Perc.Area[Scenario == "RCP 8.5"],
              pch=c(1:2,4:11, 3, 12:13),
              col=legcolors,
              xlim=c(0,300))
       abline(v = c(100), col="black", lwd=1, lty=2)
       legend ("topright", legend=spp,
              col=legcolors,
              pch=c(1:2,4:11, 3, 12:13),
              cex=0.8, text.font=3)
```

Figure 1. Relation between maximum latitude with suitability area in the # present scenario for each Western Atlantic Sargassum species and the latitudinal # variation predicted in our model results

```
windows()
par(mfrow=c(1,2))
### North
plot (N[Scenario == "present"] ~ dN [Scenario == "present"], #main="present",
       col=legcolors,
      pch=c(1:2,4:11, 3, 12:13),
      xlim=c(0,10),
      xlab="North latitudinal variation (degrees)",
      ylab="Maximum present North latitude (degrees)")
par(new=TRUE)
plot (N[Scenario == "present"] ~ dN [Scenario == "RCP 4.5"], #main="RCP 4.5",
       col=legcolors,
      pch=c(1:2,4:11, 3, 12:13),
      xlim=c(0,10),
      xlab="North latitudinal variation (degrees)",
      ylab="Maximum present North latitude (degrees)")
par(new=TRUE)
plot (N[Scenario == "present"] ~ dN [Scenario == "RCP 8.5"], #main="RCP 8.5",
       col=legcolors,
      pch=c(1:2,4:11, 3, 12:13),
       xlim=c(0,10), xlab="", ylab="")
legend ("bottomright", legend=spp,
       col=legcolors,
      pch=c(1:2,4:11, 3, 12:13),
       cex=0.5, text.font=3)
mtext("A)", side=3, line=2, at=-2.5, cex=1, family="serif")
### South
#S[Scenario == "present"] = S[Scenario == "present"]*(-1)
plot ((S[Scenario == "present"]) ~ dS [Scenario == "present"],
      ylab="Maximum present South Latitude (degrees)",
      xlab="South latitudinal variation (degrees)",
      col=legcolors,
      pch=c(1:2,4:11, 3, 12:13),
      xlim=c(0, 10))
par(new=TRUE)
plot ((S[Scenario == "present"]) ~ dS[Scenario == "RCP 4.5"],
      ylab="Maximum present South Latitude (degrees)",
      xlab="South latitudinal variation (degrees)",
       col=legcolors,
      pch=c(1:2,4:11, 3, 12:13),
      xlim=c(0, 10))
par(new=TRUE)
```

Figure S3. Correlation between changes in suitability area and changes in # latitudinal limits for 13 *Sargassum* species from the Western Atlantic Ocean.

```
par(mfrow=c(2,2))
plot (Perc.Area[Scenario == "RCP 4.5"] ~ dN[Scenario == "RCP 4.5"],
       col=legcolors,
      xlim=c(0,8),
      ylim=c(0,300),
      cex.axis =0.8,
      ylab="Suitabiliy area (%)",
      xlab = "North latitudinal variation (degrees)",
       pch=c(1:2,4:11, 3, 12:13))
abline (lm (Perc.Area[Scenario == "RCP 4.5"] ~ dN[Scenario == "RCP 4.5"]), lty=2)
mtext("A)", side=3, line=2, at=-2.2, cex=1, family="serif", font=2)
mtext("RCP 4.5", side=3, line=2, at=4, cex=1, family="serif", font=2)
plot (Perc.Area[Scenario == "RCP 8.5"] ~ dN[Scenario == "RCP 8.5"],
       col=legcolors,
      xlim=c(0,8),
      ylim=c(0,300),
      ylab="Suitabiliy area (%)",
      xlab = "North latitudinal variation (degrees)",
       cex.axis =0.8.
       pch=c(1:2,4:11, 3, 12:13))
abline (lm (Perc.Area[Scenario == "RCP 8.5"] ~ dN[Scenario == "RCP 8.5"]), lty=2)
mtext("B)", side=3, line=2, at=-2.2, cex=1, family="serif", font=2)
mtext("RCP 8.5", side=3, line=2, at=4, cex=1, family="serif", font=2)
plot (Perc.Area[Scenario == "RCP 4.5"] ~ dS[Scenario == "RCP 4.5"],
       col=legcolors,
      xlim=c(0,8),
      ylim=c(0,300),
      ylab="Suitability area (%)",
      xlab = "South latitudinal variation (degrees)",
       cex.axis = 0.8,
       pch=c(1:2,4:11, 3, 12:13))
```

abline (lm (Perc.Area[Scenario == "RCP 4.5"] ~ dS[Scenario == "RCP 4.5"]), lty=2) mtext("C)", side=3, line=2, at=-2.2, cex=1, family="serif", font=2)

mtext("D)", side=3, line=2, at=-2.2, cex=1, family="serif", font=2)

Spearman correlation tests

Correlation between latitudinal variation and change in percentual suitability area cor.test (dS[Scenario == "RCP 4.5"], Perc.Area[Scenario == "RCP 4.5"], method ="spearman", alt="greater") # non significant cor.test (dS[Scenario == "RCP 8.5"], Perc.Area[Scenario == "RCP 8.5"], method ="spearman", alt="greater") # non significant cor.test (dN[Scenario == "RCP 4.5"], Perc.Area[Scenario == "RCP 4.5"], method ="spearman", alt="greater") # significant cor.test (dN[Scenario == "RCP 8.5"], Perc.Area[Scenario == "RCP 4.5"], method ="spearman", alt="greater") # significant cor.test (dN[Scenario == "RCP 8.5"], Perc.Area[Scenario == "RCP 8.5"], method ="spearman", alt="greater") # significant cor.test (dN[Scenario == "RCP 8.5"], Perc.Area[Scenario == "RCP 8.5"], method ="spearman", alt="greater") # non significant

Correlation between overlap areas and changes in percentual suitability area
cor.test (Perc.Area[Scenario == "RCP 4.5"], Perc.Overlap[Scenario == "RCP 4.5"],
 method ="spearman", alt="greater") # significant
cor.test (Perc.Area[Scenario == "RCP 8.5"], Perc.Overlap[Scenario == "RCP 8.5"],
 method ="spearman", alt="greater") # significant

model.SWA = read.sdm("model.presente_SSargassumWA.sdm")
teste = function (i) {
 var.imp <- sdm::getVarImp(model.SWA, id=i)
 var.imp
 imp <- var.imp@varImportance
 ice = imp\$corTest[imp\$variables== "gelo"]
 temperature = imp\$corTest[imp\$variables== "temperatura"]
 depth = imp\$corTest[imp\$variables== "profundidade"]
 salinity= imp\$corTest[imp\$variables== "salinidade"]
 imp <- cbind (ice, depth, salinity, temperature)</pre>

```
colnames (imp) = c("ice", "depth", "salinity", "temperature")
 return (imp)
f \leq function(i)
 s<- rbind(
       # GLM
       \#teste (i+1),teste (i+2),teste (i+3),teste (i+4),teste (i+5),
       \#teste (i+6),teste (i+7),teste (i+8),teste (i+9),teste (i+10),
       #GAM
       teste (i+11),teste (i+12),teste (i+13),teste (i+14),teste (i+15),
       teste (i+16),teste (i+17),teste (i+18),teste (i+19),teste (i+20),
       #BRT
       teste (i+21), teste (i+22), teste (i+23), teste (i+24), teste (i+25),
       teste (i+26),teste (i+27),teste (i+28),teste (i+29),teste (i+10),
       # RF
       teste (i+31), teste (i+32), teste (i+33), teste (i+34), teste (i+35),
       teste (i+36),teste (i+37),teste (i+38),teste (i+39),teste (i+40),
       # BC
       #teste (i+41),teste (i+42),teste (i+43),teste (i+44),teste (i+45),
       #teste (i+46),teste (i+47),teste (i+48),teste (i+49),teste (i+50),
       #CART
       teste (i+51),teste (i+52),teste (i+53),teste (i+54),teste (i+55),
       teste (i+56),teste (i+57),teste (i+58),teste (i+59),teste (i+60),
       # SVM
       teste (i+61), teste (i+62), teste (i+63), teste (i+64), teste (i+65),
       teste (i+66),teste (i+67),teste (i+68),teste (i+69),teste (i+70),
       # RPART / ANN
       teste (i+71),teste (i+72),teste (i+73),teste (i+74),teste (i+75),
       teste (i+76),teste (i+77),teste (i+78),teste (i+79),teste (i+80),
       # MAXLIKE
       teste (i+81),teste (i+82),teste (i+83),teste (i+84),teste (i+85),
       teste (i+86),teste (i+87),teste (i+88),teste (i+89),teste (i+10),
       # MAXENT
       teste (i+91),teste (i+92),teste (i+93),teste (i+94),teste (i+95),
       teste (i+96),teste (i+97),teste (i+98),teste (i+99),teste (i+100)
       # MARS
       teste (i+101),teste (i+102),teste (i+103),teste (i+104),teste (i+105),
       teste (i+106),teste (i+107),teste (i+108),teste (i+109),teste (i+110)
 colnames (s) = c("ice", "depth", "salinity", "temperature")
 return (s)
Ş
f(0)
im \leq -cbind (
 mean (f(0)[,1]), #ice
 mean (f(0)[,2]), #depth
 mean (f(0)[,3]), #salinity
 mean (f(0)[,4]) #temperature
colnames (im) = c("ice", "depth", "salinity", "temperature")
```

```
### Calling ensemble rasters
Sbermudense.present <- raster
("Sbermudense/ensemble.bin.t10.presente_Sbermudense_maxent.default.tif")
Sbermudense.rcp45 <- raster
("Sbermudense/ensemble.bin.t10.rcp45_Sbermudense_maxent.default.tif")
Sbermudense.rcp85 <- raster
("Sbermudense/ensemble.bin.t10.rcp85_Sbermudense_maxent.default.tif")
```

```
### Plotting maps
windows()
par (mfrow=c(1,3))
{
    plot (Sbermudense.present, legend=F, col=my.colors(1000))
    mtext("A)", side=3, line=2, at=-120, cex=1, family="serif", font=2)
    mtext("Present", side=3, line=2, at=-60, cex=1, family="serif", font=2)
    mtext("S. bermudense", side=2, line=6, at=0, cex=1, family="serif", font=3)
```

```
plot (Sbermudense.rcp45, legend=F, col=my.colors(1000))
mtext("B)", side=3, line=2, at=-120, cex=1, family="serif", font=2)
mtext("RCP 4.5", side=3, line=2, at=-60, cex=1, family="serif", font=2)
```

plot (Sbermudense.rcp85, legend=F, col=my.colors(1000))
mtext("C)", side=3, line=2, at=-120, cex=1, family="serif", font=2)
mtext("RCP 8.5", side=3, line=2, at=-60, cex=1, family="serif", font=2)
}

im

CONCLUSÃO

Os resultados obtidos com a modelagem de nicho ecológico corroboram parcialmente nossas hipóteses iniciais. Todas as 13 espécies (12 morfoespécies e 1 espécie filogenética) de *Sargassum* analisadas tiveram manutenção ou aumento do limite latitudinal com adequabilidade ambiental propícia para a espécie. Esse aumento foi mais pronunciado no cenário RCP 8.5 do que no cenário RCP 4.5 e em direção ao norte do que em direção ao sul. Considerando que as espécies de *Sargassum* são fundadoras, essas predições sugerem que pode ocorrer um movimento de deslocamento de comunidades marinhas tropicais em direção a hábitats temperados quentes em maiores latitudes.

Contrariamente ao hipotetizado, apenas metade das morfoespécies analisadas apresentou diminuição da sua área de adequabilidade ambiental, em especial na região Equatorial e no Caribe. A outra metade apresentou aumento generalizado na sua área de adequabilidade ambiental, com aumento dessa área tanto nos pólos como em latitudes tropicais. Todas as morfoespécies tiveram o efeito em sua área de adequabilidade - seja aumento ou diminuição da área - com maior magnitude no cenário RCP 8.5 do que no cenário RCP 4.5. A espécie filogenética apresentou um ligeiro aumento de sua área de adequabilidade no cenário RCP 4.5 e uma diminuição no cenário RCP 8.5. Todas as espécies que apresentaram um aumento da área de adequabilidade ambiental incorporaram, nas predições para os cenários futuros, toda a área de adequabilidade contida no cenário presente. As espécies que tiveram sua adequabilidade ambiental diminuída não apresentaram mudanças bruscas de área em relação à predição para o presente, ou seja, suas áreas de adequabilidade futura estão dentro, em grande parte, das áreas de adequabilidade atuais.

Atenção deve ser dada a um eventual aumento das áreas de ocorrência de *S. pteropleuron* e a uma drástica redução de populações tropicais das morfoespécies *S. stenophyllum*, *S. cymosum* e *S. buxifolium*. Esforços para conservação e iniciativas de manejo e, principalmente, de pesquisas e monitoramento destas espécies devem ser mantidas e ampliadas.

REFERÊNCIAS

- Abdel-Kareem, M. S. M. 2009. Phenetic Studies and New Records of Sargassum Species (Fucales, Phaeophyceae) from the Arabian Gulf Coast of Saudi Arabia. Academic Journal of Plant Sciences 2(3):173-181.
- Abe, H., Suzuki, H., Kumagai, N. H. & Hiroya Yamano, H. 2020. Distribution gradient of fucoid algae (Sargassaceae, Phaeophyta) along the coastline of Okinawa Island, southern Japan: Relationship to environmental factors. *Regional Studies in Marine Science*, 40, 101526.
- Agardh, C. A. 1821. Species algarum rite cognitae, cum synonymis, differentiis specificis et descriptionibus succinctis. Volumen primum. Pars prima. Fucoideae. ex officina Berlingiana, Lundae [Lund], 168 pp.
- Agardh, C. A. 1824. Systema algarum. Literis Berlingianis [Berling], Lundae [Lund], 312 pp.
- Agardh, J. G. 1847. Nya alger från Mexico. Öfversigt af Kongl. Vetenskaps-Adademiens Förhandlingar 4:5-17.
- Agardh, J. G. 1848. Species genera et ordines algarum, seu descriptiones succinctae specierum, generum et ordinum, quibus algarum regnum constituitur. Volumen Primum. Algas fucoideas complectens. I. Lundae [Lund]: C.W.K. Gleerup. 363p.
- Airoldi, L., Ballesteros, E., Buonomo, R., Van Belzen, J., Bouma, T. J., Cebrian, E., De Clerk, O., Engelen, A. H., Ferrario, F., Fraschetti, S., Gianni, F., Guidetti, P., Ivesa, L., Mancuso, F. P., Micheli, F., Perkol-Finkel, S., Serrão, E. A., Strain, E. M., Mangialajo, L. 2014. Marine forests at risk: solutions to halt the loss and promote the recovery of Mediterranean canopy-forming seaweeds. *Proceedings of the 5th Mediterranean Symposium on Marine Vegetation (Portorož, Slovénie, 27-28 octobre 2014)*, pp. 28–33.
- Almeida, R. S. 2014. Avaliação do efeito da acidificação e da temperatura na fisiologia de Sargassum stenophyllum (Phaeophyceae) – Subsídios aos estudos relacionados ao aquecimento global e acidificação dos oceanos. Biology monograph. Universidade Federal de Santa Catarina, Florianópolis, Brazil, 42 pp.

- Alquezar, R., Glendenning, L. & Costanzo, S. 2013. The use of the brown macroalgae, Sargassum flavicans, as a potential bioindicator of industrial nutrient enrichment. Marine Pollution Bulletin. 77:140-146.
- Atmadja, W. S. & Prud'homme van Reine, W. F. 2014. Checklist of the seaweed species biodiversity of Indonesia with their distribution and classification: green algae (Chlorophyta) and brown algae (Phaeophyceae, Ochrophyta). Naturalis Biodiversity Centre, Indonesian Institute of Sciences (LIPI), Leiden/Indonesia, 59 pp.
- Ballantine, D. L., Torres, H. R. & Aponte, N. E. 2016. The Mesophotic, Coral Reef–Associated, Marine Algal Flora of Puerto Rico, Caribbean Sea. *Smithsonian Contributions to Botany* 105:1-41.
- Barry, J. P., Baxter, C.H., Sagarin, R.D. & Gilman, S.E. 1995 Climate related, long-term faunal changes in a California rocky intertidal community. *Science* 267:672–675.
- Bernecker, A. 2009. Marine benthic algae and Cyanobacteria reported from the Caribbean coast of Costa Rica. In Wehrtmann, I. S. & Cortés, J. [Eds.] Marine Biodiversity of Costa Rica, Central America. Monographiae Biologicae 86, Springer, pp. 17-52.
- Bostock, P. D. & Holland, A. E. 2010. Census of the Queensland Flora. Queensland Herbarium Biodiversity and Ecosystem Sciences, Department of Environment and Resource Management, Brisbane, Australia, 320 pp.
- Brayner-Barros, S. G., Eskinazi-Leça, E., Oliveira, N. M. B. 2006, Diatomáceas epífitas em Sargassum polyceratium (Phaeophyta) coletada no litoral de Pernambuco. Anais do XI Congresso Brasileiro de Ficologia, Itajaí (SC). pp.113-121.
- Brooks, M. T., Coles, V. J., Hood, R. R. & Gower, J. F. 2018 Factors controlling the seasonal distribution of pelagic *Sargassum*. Mar. Ecol.-Prog. Ser., 599:1–18.
- Cabrera, R., Díaz-Larrea, J. & Umanzor, S. 2019. New records of marine macroalgae on the Caribbean on coast of Costa Rica. *American Journal of Plant Sciences* 10:1708-1728

- Camacho, O., Mattio, L., Draisma, S., Fredericq, S. & Diaz-Pulido, G. 2015. Morphological and molecular assessment of *Sargassum* (Fucales, Phaeophyceae) from Caribbean Colombia, including the proposal of *Sargassum giganteum* sp. nov., *Sargassum schnetteri* comb. nov. and *Sargassum* section *Cladophyllum* sect. nov. *Systematics and Biodiversity* 13(2):105-130.
- Carela, C. D. D. 1977. Contribución al Estudio de los Moluscos en el Litoral de la República Dominicana. Thesis. Universidad Autónoma de Santo Domingo, Santo Domingo, República, Dominicana, 212 pp.
- Carneiro, P. B. M. 2017. Escalas Espaciais e Biodiversidade de Organismos Bentônicos no Atlântico Sudoeste. Ph.D. thesis. Universidade Federal do Ceará, Fortaleza, Brazil, 124pp.
- Casado-Amezúa, P., Araújo, R., Bárbara, I., Bermejo, R., Borja, Á., Díez, I., Fernández, C., Gorostiaga, J. M., Guinda, X., Hernández, I., Juanes, J. A., Peña, V., Peteiro, C., Puente, A., Quintana, I., Tuya, F., Viejo, R. M., Altamirano, M., Gallardo, T. & Martínez, B. 2019. Distributional shifts of canopy-forming seaweeds from the Atlantic coast of Southern Europe. *Biodiversity and Conservation* 28:1151–1172.
- Cordero, M. K. 2016. Caracterización ficológica del litoral rocoso y manglares de Yapascua, Parque Nacional San Esteban, Estado de Carabobo. Trabajo Especial de Grado en Licenciado en Biología. Universidad de Carabobo, Valencia, Venezuela. 137pp.
- CRIA Centro de Referência em Informação Ambiental. 2018. SpeciesLink Network. Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP). Available at <u>http://www.splink.org.br</u> (last accessed 4 June 2022).
- Connor, J. & Adey, W. 1977. The benthic algal composition, standing crop, and productivity of a Caribbean algal ridge. *Atoll Res. Bull.* 211:1-40.
- Cordero, P. A., Jr. 1981. Eco-morphological observation of the genus Sargassum in central Philippines, including notes on their biomass and bed determination. Proc. Fourth Int. Coral Reef Symp. 2:399-409.

- Dawes, C. J. 1989. Physiological Responses of Brown Seaweeds Sargassum filipendula and S. pteropleuron Before and After Transplanting on the West coast of Florida. Journal of Coastal Research 5(4):693-700.
- Dawes, C. J. & Tomasko, D. A. 1988. Physiological responses of perennial bases of *Sargassum filipendula* from three sites on the west coast of Florida. *Bull Mar Sci* 42:166–173.
- Dawes, C. J., Bird, K. & Hanisak, M. D. 1988. Physiological Responses of Transplanted Populations of *Sargassum pteropleuron* Grunow in Florida. *Aquatic Botany* 31:107-123.
- De Toni, G. B. 1895. Sylloge algarum omnium hucusque cognitarum. III. Sylloge Fucoidearum. Patavii, 638 pp.
- Diaz-Pulido, G. & Diaz-Ruiz, M. 2003. Diversity of Benthic Marine Algae of the Colombian Atlantic. *Biota Colombiana* 4(2):203-246.
- Dibner, S., Martin, L., Thibaut, T., Aurelle, D., Blanfuné, A., Whittaker, K., Cooney, L., Schell, J., Goodwin, D. & Siuda, A. 2021. Consistent genetic divergence observed among pelagic *Sargassum* morphotypes in the western North Atlantic. *Marine Ecology* 43(1): e12691.
- Dreckmann, K. M., Stout, I. & Granados, A. B. 1996. Lista actualizada de las algas marinas bentónicas de Puerto Morelos, Quintana Roo, Caribe Mexicano. *Polibotánica* 3:1-17.
- Earle, S. A. 1969. Phaeophyta of the eastern Gulf of Mexico. Phycologia 7:71-254.
- Engelen, A. H., Aberg, P., Olsen, J. L., Stam, W. T. & Breeman, A. M. 2005. Effects of wave exposure and depth on biomass, density and fertility of the fucoid seaweed *Sargassum polyceratium* (Phaeophyta, Sargassaceae). *European Journal of Phycology* 40:149-158.
- Eggertsen, L., Ferreira, C., Fontoura, L., Kautsky, N., Gullström, M. & Berkström, C. 2017. Seaweed beds support more juvenile reef fish than seagrass beds: Carrying capacity in a south-western Atlantic tropical seascape. *Estuarine, Coastal and Shelf Science* 196:97-108.

- Einav, R., Guiry, M. D. & Israel, A. 2021. A revised list of seaweeds from the Red Sea (1756–2020). *Israel Journal of Plant Sciences* 67:1-73.
- Eston, V. R. & Bussab, W. O. 1990. An experimental analysis of ecological dominance in a rocky subtidal macroalgal community. *J. Exp. Mar. Biol. Ecol.* 136:170-195.
- Farnham, W. F., Murfin, C., Critchley, A. T. & Morrell, S. L. 1981. Distribution and control of the brown alga Sargassum muticum. Proc. Int. Seaweed Symp. 10:277-82.
- Ferreira, S. J., Gonçalves Silva, J. J. & Araújo, R. 2018. Marine algae collection in the Herbarium of the Funchal Natural History Museum (MADM) with new records from the archipelago of Madeira. *Boletim do Museu Municipal do Funchal* 68(352):31-52.
- Fialho, F. A. N. 2015. Desenvolvimento de técnicas de cultivo da macroalga Sargassum filipendula (Ochrophyta, Fucales), no sul do Brasil. MsC. dissertation. Universidade Federal de Santa Catarina, Florianópolis, Brazil, 46 pp.
- Figueiredo, M. A. O. 2006. Diversity of macrophytes on the Abrolhos Bank, Brazil. *In* G. F. Dutra,
 G. F., Allen, G. R., Werner T. & McKenna, S. A. [Eds.] *A Rapid Marine Biodiversity Assessment of the Abrolhos Bank, Bahia, Brazil.* Conservation International, Washington, pp. 67-74
- Filbee-Dexter, K. 2020. Ocean Forests Hold Unique Solutions to Our Current Environmental Crisis. *One Earth*. 2:398-401.
- Filbee-Dexter, K., Feehan, C. J. & Scheibling, R. E. 2016. Large-scale degradation of a kelp ecosystem in an ocean warming hotspot. *Marine Ecology Progress Series* 543:141–152.
- Flores-Moya, A. & Conde, F. 1998. Fragmentos taxonómicos, corológicos, nomenclaturales y fitocenológicos. Nueva citas de macroalgas marinas para las Islas Chafarinas. *Acta Botanica Malacitana* 23:197-199.

- Franco, J., Bertocci, I., Rodriguez, L., Martinez, B., Sousa Pinto, I. & Arenas, F. 2018. The 'golden kelp' *Laminaria ochroleuca* under global change: Integrating multiple eco-physiological responses with species distribution models. *Journal of Ecology* 106:47-58.
- Gabriel, D. & Fredericq, S. 2019. The marine macroalgae of Cabo Verde archipelago: an updated checklist. Arquipelago. *Life and Marine Sciences* 39:39-60.
- Gallardo, T., Bárbara, I., Afonso-Carrillo, J., Bermejo, R., Altamirano, M., Gómez Garreta, A., Barceló Martí, M. C., Rull Lluch, J., Ballesteros, E. & De la Rosa, J. 2016. Nueva lista crítica de las algas bentónicas marinas de España. A new checklist of benthic marine algae of Spain. *Algas. Boletín Informativo de la Sociedad Española de Ficología* 51:7-52.
- GBIF The Global Biodiversity Information Facility. 2020, What is GBIF? Available at: <u>https://www.gbif.org/what-is-gbif</u> (last accessed 1 June 2020).
- Godínez-Ortega, J. L., Cuatlán-Cortés, J. V., López-Bautista, J. M. & Tussenbroek, B. I. v. 2021. A Natural History of Floating Sargassum Species (Sargasso) from Mexico. In Hufnagel, L. [Ed.]. Natural History and Ecology of Mexico and Central America. IntechOpen, London. Available at: <u>https://www.intechopen.com/chapters/76196</u> (last accessed 4 June 2022).
- González-Nieto, D., Oliveira, M., Núñez Resendiz, M., Dreckmann, K., Mateo-Cid, L. & Sentíes, A. 2020. Molecular assessment of the genus *Sargassum* (Fucales, Phaeophyceae) from the Mexican coasts of the Gulf of Mexico and Caribbean, with the description of *S. xochitlae* sp. nov. *Phytotaxa* 461:254-274.
- Gouvêa, L. P., Assis, J., Gurgel, C. F. D., Serrão, E. A., Silveira, T. C. L., Santos, R., Duarte, C. M., Peres, L. M. C., Carvalho, V. F., Batista, M., Bastos, E., Sissini, M. N. & Horta, P. A. 2020.
 Golden carbon of *Sargassum* forests revealed as an opportunity for climate change mitigation. *Science of The Total Environment* 729:138745 pp.1-10.
- Grunow, A. 1868. Algae. In Fenzl, E. [Ed.] Reise der österreichischen Fregatte Novara um die Erde in den Jahren 1857, 1858, 1859 unter den Befehlen des Commodore B. von Wüllerstorf-Urbair. Botanischer Theil. Erster Band. Sporenpflanzen. Aus der Kaiserlich Königlichen Hof- und Staatsdruckeri in Commission bei Karl Gerold's Sohn., Wien [Vienna], pp. 1-104.

- Grunow, A. 1916. Additamenta ad cognitionem Sargassorum. Verhandlungen der Kaiserlich-Koniglichen Zoologisch-Botanischen Gesellschaft in Wien. 66:1-48, 136-185.
- Guimaraens, M. A., Combells, C. & Corbett, C. 1994. Species diversity and richness of reef building corals and macroalgae of reef communities in Discovery Bay, Jamaica. Acta Biologica Leopoldensia 16:41-50.
- Guiry, M.D. & Guiry, G.M. 2020. AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. Available at: <u>https://www.algaebase.org</u> (last accessed 4 June 2022).
- Guitouni, M., Madan, A. A. & Al-Shabeeb, S. S. 2016. Seaweeds as bioindicators of heavy metals pollution in Tarut Bay, Saudi Arabia. *International Journal of Advanced Research*. 4(10):1095-1105.
- Haddad, M. A. & A. P. Chiaverini. 2000. Repartição de espaço entre hidróides (Cnidaria, Hydrozoa) epifiticos em Sargassum stenophylum (Phaeophyta, Fucales) de Guaratuba, Paraná. Anais V Simpósio de Ecossistemas Brasileiros: Conservação 2:101-109.
- Harley, C. D. G., Anderson, K. M., Demes, K. W., Jorve, J. P., Kordas, R. L., Coyle, T. A. & Graham, M. H. 2012. Effects of climate change on global seaweed communities. *Journal of Phycology* 48:1064–1078.
- Haroun, R. J., Gil-Rodríguez, M. C., Díaz de Castro, J. & Prud'homme van Reine, W. F. 2002. A checklist of the marine plants from the Canary Islands (central eastern Atlantic Ocean). *Botanica Marina* 45:139-169.
- Hawkins, S. J., Southward, A., Genner, M. J. 2003. Detection of environmental change in a marine ecosystem – evidence from the western English Channel. *Science of the Total Environment* 310:245–256.

- Helmuth, B., Harley, C. D., Halpin, P. M., O'Donnell, M., Hofmann, G. E. & Blanchette, C. A. 2002. Climate change and latitudinal patterns of intertidal thermal stress. *Science* 298:1015– 1017.
- Ho, M. T. G. & Bantoto-Kinamot, V. 2021. Sargassum, Padina and Turbinaria as bioindicators of cadmium in Bais Bay, Negros Oriental. The Palawan Scientist 13(1):90-98.
- IPCC Intergovernmental Panel on Climate Change. 2014. *Climate Change 2014 Synthesis Report: Summary for Policymakers*. IPCC, Geneva, Switzerland, 31pp.
- John, D. M. & Al-Thani, R. F. 2014. Benthic marine algae of the Arabian Gulf: a critical review and analysis of distribution and diversity patterns. *Nova Hedwigia* 98:341-392.
- John, D. M., Lawson, G. W. & Ameka, G. K. 2003. The marine macroalgae of the Tropical West Africa Subregion. *Beihefte zur Nova Hedwigia* 125:1-217.
- John, D. M., Prud'homme van Reine, W. F., Lawson, G. W., Kostermans, T. B. & Price, J. H. 2004. A taxonomic and geographical catalogue of the seaweeds of the western coast of Africa and adjacent islands. *Beihefte zur Nova Hedwigia* 127:1-339.
- Jonsson, P. R., Kotta, J., Andersson, H. C., Herkül, K., Virtanen, E., Sandman, A. N. & Johannesson, K. 2018. High climate velocity and population fragmentation may constrain climate-driven range shift of the key habitat former *Fucus vesiculosus*. *Diversity and Distributions* 24:892–905.
- Jouanno, J., Benshila, R., Berline, L., Soulié, A., Radenac, M.-E., Morvan, G., Diaz, F., Sheinbaum, J., Chevalier, C., Thibaut, T., Changeux, T., Ménard, F., Berthet, S., Aumont, O., Ethé, C., Nabat, P. & Mallet, M. 2021. A NEMO-based model of *Sargassum* distribution in the tropical Atlantic: Description of the model and sensitivity analysis (NEMO-Sarg1.0). *Geoscientific Model Development* 14:4069-4086.
- Karthick, P., Mohanraju, R., Ramesh, C. & Kada, N. M. 2013. Distribution and diversity of seaweeds in North and South Andaman Island. *Seaweed Res. Utiln.* 35:8-16.

- Kleypas, J. A. 2019. Climate change and tropical marine ecosystems: A review with an emphasis on coral reefs. *UNED Research Journal* 11(1) Núm. especial: S24-S35.
- Kützing, F. T. 1843. Phycologia generalis oder Anatomie, Physiologie und Systemkunde der Tange. Mit 80 farbig gedruckten Tafeln, gezeichnet und gravirt vom Verfasser. F.A. Brockhaus, Leipzig, 458 pp.
- Kützing, F. T. 1849. Species algarum. F.A. Brockhaus, Lipsiae [Leipzig], 922 pp.
- Kützing, F. T. 1861. *Tabulae phycologicae; oder, Abbildungen der Tange. Vol. XI.* Gedruckt auf kosten des Verfassers (in commission bei W. Köhne), Nordhausen, 32pp. 100pls.
- Lapointe, B. E. 1986. Phosphorus-limited photosynthesis and growth of *Sargassum natans* and *Sargassum fluitans* (Phaeophyceae) in the western North Atlantic. *Deep-Sea Res. Pt. A* 33:391–399.
- Lapointe, B. E. 1995. A comparison of nutrient-limited productivity in Sargassum natans from neritic vs. oceanic waters of the western North Atlantic Ocean. Limnol. Oceanogr. 40:625– 633.
- Lawson, G. W. & John, D. M. 1987. The marine algae and coastal environment of Tropical West Africa (second edition). *Beih. Nova Hedwigia* 93:vi-415.
- Leite, F. P. P., Tanaka, M. O. & Gebara, R. S. 2007. Structural variation in the brown alga Sargassum cymosum and its effects on associated amphipod assemblages. Braz. J. Biol. 67(2):215-221.
- Levitus, S., Antonov, J. I., Boyer, T. P. & Stephens, C. 2000. Warming of the World Ocean. *Science* 287:2225–2229.
- Lima, F. P., Ribeiro, P. A., Queiroz, N., Hawkins, S. J., & Santos, A. M. 2007. Do distributional shifts of northern and southern species of algae match the warming pattern? *Global Change Biology* 13:2592–2604.

- Littler, D. S. & Littler, M. M. 1997. An illustrated flora of the Pelican Cays, Belize. *Bulletin of the Biological Society of Washington* 9:1-149.
- Littler, D. S. & Littler, M. M. 2000. Caribbean Reef Plants: An Identification Guide to the Reef Plants of the Caribbean, Bahamas, Florida and Gulf of Mexico. OffShore Graphics, Washington, 542 pp.
- Machado, G. B. O., Neufeld, A. B., Dena, S. A., Siqueira, S. G. L. & Leite, F. P. P. 2015. Variation of amphipod assemblage along the *Sargassum stenophyllum* (Phaeophyta, Fucales) thallus. *Nauplius* 23(1):73-78.
- Mafra Junior, L. L. 2001. Bases para o manejo de *Sargassum cymosum* (Phaeophyta-Fucales) na enseada de Armação do Itapocoroy, Penha, SC. Oceanography monography. Universidade do Vale do Itajaí, Itajaí, Brasil, 51 pp.
- Marcot-Coqueugniot, J. 1991. A preliminary list of marine algae from the Banc d'Arguin (Mauritania). *Botanica Marina* 34:195-199.
- Marins, B. V., Brasileiro, P. S., Barros Barreto, M. B., Nunes, J. M. C., Yoneshigue-Valentin, Y. & Amado Filho, G. M. 2008. Subtidal benthic marine algae of the Todos os Santos bay, Bahia State, Brazil. *Oecol. Bras.* 12(2):229-242.
- Martius, C. F. P. von. [1828?-]1834. Icones plantarum cryptogamicarum quas in itinere annis MDCCCXII-MDCCCXX per Brasiliam jussu et auspiciis Maximiliani Josephi I. Bavariae regis augustissimi instituto collegit et descripsit Carol. Frideric. Philip. de Martius. Impensis auctoris, Monachii [Munich], 138pp.
- Martius, C. F. P. von. 1833. Flora brasiliensis, seu, enumeratio plantarum in Brasilia: tam sua sponte quam accedente cultura provenientium, quas in itinere auspiciis Maximiliani Josephi I. Bavariae Regis annis 1817-1820 peracto collegit, partim descripsit; alias a Maximiliano Seren. Principe Widensi, sellovio aliisque advectas addidit, communibus amicorum propriisque studiis secundum methodum naturalem dispositas et illustratas. Vol. I. Pars prior. Algae, lichenes, hepaticae exposuerunt Martius, Eschweiler, Nees ab Esenbeck. Sumptibus J.G. Cottae, Stuttgartiae/Tubingae [Stuttgart/Tübingen], 390pp.

- Mattio, L. & Payri, C. E. 2009. Taxonomic revision of *Sargassum* species (Fucales, Phaeophyceae) from New Caledonia based on morphological and molecular analyses. *J. Phycol.* 45:1374–88.
- Mattio, L. & Payri, C. E. 2011. 190 years of *Sargassum* taxonomy, facing the advent of DNA phylogenies. *Bot. Rev.* 77:31–70.
- Mattio, L., Payri, C. E. & Stiger-Pouvreau, V. 2008. Taxonomic revision of Sargassum (Fucales, Phaeophyceae) from French Polynesia based on morphological and molecular analyses. J. Phycol. 44:1541–55.
- Mattio, L., Payri, C. E. & Verlaque, M. 2009. Taxonomic revision and geographic distribution of the subgenus *Sargassum* (Fucales, Phaeophyceae) in the Western and Central Pacific islands based on morphological and molecular analyses. *J. Phycol.* 45:1213–27.
- Mattio, L., Zubia, M., Loveday, B., Crochelet, E., Duong, N., Payri, C. E., Bhagooli, R. & Bolton, J. J. 2013. *Sargassum* (Fucales, Phaeophyceae) in Mauritius and Reunion, western Indian Ocean: taxonomic revision and biogeography using hydrodynamic dispersal models. *Phycologia* 52(6):578–94.
- Mattio, L., Anderson, R. J. & Bolton, J. J. 2015. A revision of the genus *Sargassum* (Fucales, Phaeophyceae) in South Africa. S. Afr. J. Bot. 98:95–107.
- Menezes-Széchy, M. T. & Paula, J. C. de. 2010. Phaeophyceae. In Forzza, R. C. [Ed.] Catálogo de plantas e fungos do Brasil. Vol. 1. Andrea Jakobsson Estúdio; Instituto de Pesquisas Jardim Botânico do Rio de Janeiro, Rio de Janeiro, Brazil, pp. 404-408.
- Miki, O., Nagai, T., Marzuki, M., Okumura, C., Kosugi, C. & Kato, T. 2016. Effects of Fe fertilizer eluate on the growth of *Sargassum horneri* at the germling and immature stages. *J. Appl. Phycol.* 28:1775–1782.
- Montagne, C. 1837. Centurie de plantes cellulaires exotiques nouvelles. *Annales des Sciences Naturelles, Botanique, Seconde série* 8: 345-370.

- Montagne, C. 1842. Troisième centurie de plantes cellulaires exotiques nouvelles. Décades V, VI, VII et VIII. *Annales des Sciences Naturelles, Botanique, Seconde Série* 18:241-282.
- Moreira, L. & Cabrera, R. 2007. El género *Sargassum* (Phaeophyceae) en las costas cubanas. Lista y notas taxonómicas. *Revista de Investigaciones Marinas* 28(3):259-267.
- Moreira, L., & Suárez, A. M. 2002. Estudio del género Sargassum C. Agardh, 1820 (Phaeophyta, Fucales, Sargassaceae) en aguas cubanas. 3. Variaciones morfológicas de Sargassum filipendula C. Agardh. Revista de Investigaciones Marinas 23:59 -62
- Morrissey, J. 1980. Community structure and zonation of macroalgae and hermatypic corals on a fringing reef flat of Magnetic Island (Queensland, Australia). *Aquat. Bot.* 8:91-139.
- Muller, R., Laepple, T., Bartsch, I. & Wiencke, C. 2009. Impact of oceanic warming on the distribution of seaweeds in polar and cold-temperate waters. *Bot. Mar.* 52:617–638.
- Nguyen, T. V., Le, N. H., Lin, S.-M., Steen, F. & De Clerck, O. 2013. Checklist of the marine macroalgae of Vietnam. *Botanica Marina* 56(3):207-227.
- Nunes, J. M. C. & Paula, E. J. 2002. Composição e distribuição das Phaeophyta nos arrecifes da Região Metropolitana de Salvador, Bahia, Brasil. *Iheringia Sér. Bot.* 57(1):113-130.
- Oliveira, E. C., Horta, P. A., Amancio, C. E. & Sant'Anna, C. L. 2002. Algas e angiospermas marinhas bênticas do litoral brasileiro: diversidade, explotação e conservação. *In Ministério* do Meio Ambiente [Ed.] *Workshop sobre Avaliação e ações prioritárias para a conservação da Biodiversidade das zonas costeira e marinha. Relatório Técnico.* Brasília, Brazil, 12pp.
- Oliveira, V. P., Fernandes, D. R. P., Figueiredo, N. M., Valenti, Y. Y. & Garla, R. C. 2009. Notes on Geographic Distribution: Four new additions to the marine flora of Fernando de Noronha Archipelago, Tropical western South Atlantic Ocean. *Check List* 5(2):210–215.
- Oliveira, N. M., Meira, C. L. C., Aguiar, R. M., Oliveira, D. M., Moura, C. W. N. & Vieira Filho, S.A. 2015. Biological activities of extracts from *Padina boergesenii* and *Sargassum*

stenophyllum, seaweeds naturally found in Baía de Todos os Santos, Brazil. International Journal of Pharmacy and Pharmaceutical Sciences 7(1):350-353.

- Oliveira Filho, E. C. de. 1977. *Algas marinhas bentônicas do Brasil*. Thesis. Instituto de Biociências, Universidade de São Paulo, São Paulo, Brazil, 406 pp.
- Ornellas, A. B. & Coutinho, R. 1998. Spatial and temporal patterns of distribution and abundance of a tropical fish assemblage in a seasonal *Sargassum* bed, Cabo Frio Island, Brazil. *Journal of Fish Biology* 53(Supplement A):198–208.
- Oviatt, C. A., Huizenga, K., Rogers, C. S. & Miller, W. J. 2019. What nutrient sources support anomalous growth and the recent *Sargassum* mass stranding on Caribbean beaches? A review. *Mar. Pollut. Bull.* 145:517–525.
- Parente, M. I., Gil-Rodríguez, M. C., Haroun, R. J., Neto, A. I., de Smedt, G., Hernández-González, C. L. & Berecibar Zugasti, E. 2000. Flora marina de las Ilhas Selvagens: resultados preliminares de la expedición "Macronesia 2000". *Revista de la Academia Canaria de Ciencias* 12(3-4):9-20
- Parmesan, C. 2006. Ecological and evolutionary responses to recent climate change. *Annual Review* of Ecology, Evolution, and Systematics 37:637–669.
- Paula, E. J. 1988. O Gênero Sargassum C. Ag. (Phaeophyta Fucales) no Litoral do Estado de São
 Paulo, Brasil. Boletim de Botânica da Universidade de São Paulo 10:65-118.
- Paula, E. J. & Oliveira-Filho, E. C.. 1980. Aspectos fenológicos de duas populações de Sargassum cymosum (Phaeophyta – Fucales) do litoral de São Paulo, Brasil. Bolm. Botânica, Univ. S. Paulo. 8:21-39.
- Paula, E. J. & Oliveira-Filho, E. C.. 1982. Wave exposure and ecotypical differentiation in Sargassum cymosum (Phaeophyta - Fucales). Phycologia 21(2):145-153.

- Pereira, R. C. & Yoneshigue-Valentin, Y. 1999. The Role of Polyphenols from the Tropical Brown Alga *Sargassum furcatum* on the Feeding by Amphipod Herbivores. *Botanica Marina* 42:441-448.
- Pereira, S. M. B., Oliveira-Carvalho, M. F., Burgos, D. C. & Araújo, E. L. 2006. Caracterização estrutural das macroalgas do ambiente recifal da praia de Enseada dos Corais - Pernambuco -Brasil. Anais do XI Congresso Brasileiro de Ficologia, Itajaí (SC). Itajaí, Brazil, pp.231-242.
- Phang, S.-M., Yeong, H.-Y., Ganzon-Fortes, E. T., Lewmanomont, K., Prathep, A., Hau, L. N., Gerung, G. S. & Tan, K. S. 2016. Marine algae of the South China Sea bordered by Indonesia, Malaysia, Philippines, Singapore, Thailand and Vietnam. *Raffles Bulletin of Zoology Supplement* 40:13-59
- Phillips, N. 1995. Biogeography of Sargassum (Phaeophyta) in the Pacific basin. In Abbott, I. A. [Eds.] Taxonomy of Economic Seaweeds. Vol. 5. California Sea Grant College System, La Jolla, California, pp. 107-145.
- Phillips, N. & Fredericq, S. 2000. Biogeographic and phylogenetic investigations of the pantropical genus Sargassum (Fucales, Phaeophyceae) with respect to Gulf of Mexico Species. Gulf of Mexico Science 2:77-87.
- Price, J. H., John, D. M. & Lawson, G. W. 1978. Seaweeds of the western coast of tropical Africa and adjacent islands: a critical assessment. II. Phaeophyta. *Bulletin of the British Museum* (*Natural History*) Botany 6:87-182.
- Prince, J. S. & O'Neal, S. W. 1979. The ecology of Sargassum pteropleuron Grunow (Phaeophyceae, Fucales) in the waters off South Florida. I. Growth, reproduction and population structure. *Phycologia* 18(2):109-114.
- Prud'homme Van Reine, W. F. 2011. Report of the Nomenclature Committee for Algae: 11. *Taxon* 60(3):898-899.
- Raut, Y., Morando, M. & Capone, D. G. 2018. Diazotrophic Macroalgal Associations with Living and Decomposing Sargassum. Front. Microbiol. 9:3127.

- Razzera, V. G. 2018. Avaliação da ecofisiologia de Sargassum stenophyllum sob diferentes irradiâncias e concentrações de nutrientes. Oceanography monograph. Universidade Federal de Santa Catarina, Florianópolis, Brazil, 43pp.
- Richardson, W. D. 1975. The marine algae of Trinidad, West Indies. *Bulletin of the British Museum* (*Natural History*) *Botany* 5(3):1-143, 27 pls.
- Robinson, N. M., Galicia-García, C. & Okolodkov, Y. B. 2012. New records of green (Chlorophyta) and brown algae (Phaeophyceae) for Cabezo Reef, National Park Sistema Arrecifal Veracruzano, Gulf of Mexico. *Acta Botanica Mexicana* 101:11-48.
- Rocha, C. M. C 2003. Efeito do substrato fital na comunidade meiofaunística associada, com ênfase aos Nematoda livre. Ph.D. thesis. Universidade Federal de Pernambuco, Recife, Brazil, 117 pp.
- Rocha-Jorge, R. 2015. Composição de Macroalgas em Unidades de Conservação Insulares do Estado de São Paulo, Brasil. Ph.D. thesis. Instituto de Botânica da Secretaria de Estado de Meio Ambiente, São Paulo, Brazil, 238 pp.
- Rosado-Espinosa, L. A. 2012. Caracterización ficológica de comunidades bentónicas y de arribazón en Dzilam de Bravo, Yucatán. MsC. thesis. Universidad Autónoma de Yucatán, Mérida, Yucatán, México, 76 pp.
- Rover, T. 2014. Caracterização estrutural e ultraestrutural do processo de embriogênese de Sargassum cymosum C. Agardh (Phaeophyceae, Fucales) como modelo de estudo de germinação de algas pardas tropicais, e sua alteração pela radiação ultravioleta. Ph.D. thesis. Universidade Federal de Santa Catarina, Florianópolis, Brazil, 153 pp.
- Sahoo, D., Sahoo, N. & Bhattacharya, D. 2001. *Seaweeds of Indian coast.* A.P.H. Publishing, New Delhi, India, 283 pp.

- Salem, W. M., Galal, H. & Nasr El-deen, F. 2011. Screening for antibacterial activities in some marine algae from the Red Sea (Hurghada, Egypt). *African Journal of Microbiology Research*. 5(15):2160–2167.
- Schell, J. M., Goodwin, D. S. & Siuda, A. N. 2015. Recent Sargassum inundation events in the Caribbean: shipboard observations reveal dominance of a previously rare form. *Oceanography* 28:8–11.
- Scherner, F., Bonomi Barufi, J. & Horta, P. A. 2012. Photosynthetic response of two seaweed species along an urban pollution gradient: Evidence of selection of pollution-tolerant species. *Marine Pollution Bulletin* 64:2380–2390.
- Schneider, C. W. 2003. An annotated checklist and bibliography of the marine macroalgae of the Bermuda Islands. *Nova Hedwigia* 76(3-4):275-361.
- Schneider, C. W. & Flook, C. T. 2017. Could marine animal conservation laws be responsible for the decline or extirpation of macroalgal populations in Bermuda over the past century? *Botanica Marina* 60(6):591-602.
- Semidey, A., Busutil, L., Espinosa, J., González-Rodríguez, N., Durán, A., Álvarez, S. & Hernández, J. L. 2013. Inventario de especies marinas de los cayos del norte del Golfo de Batabanó, SO de Cuba. Serie Oceanológica 13:95-109.
- Silva, P. C., Meñez, E. G. & Moe, R. L. 1987. Catalog of the benthic marine algae of the Philippines. *Smithsonian Contributions to Marine Sciences* 27:1-179.
- Silva, P. C., Basson, P. W. & Moe, R. L. 1996. Catalogue of the benthic marine algae of the Indian Ocean. *University of California Publications in Botany* 79:1-1259.
- Siqueira, S. G. L. 2012. Alga parda Sargassum furcatum e anfipodes ampitoídeos associados como potenciais bioindicadores de poluição por hidrocarbonetos de petróleo. Ph.D. thesis. Universidade de Campinas, Campinas, Brazil, 126 pp.
- Solé, M. A. & Pardo, P. 2005. Contribución al conocimiento taxonómico de la ficoflora marina de la Isla de Margarita, Estado Nueva Esparta, Venezuela. *Memoria de la Fundación La Salle de Ciencias Naturales* 165:5-32.
- Southward, A. J., Hawkins, S. J. & Burrows, M. T. 1995 Seventy years' observations of changes in distributions and abundance of zooplankton and intertidal organisms in the Western English Channel in relation to rising sea temperature. *Journal of Thermal Biology* 20:127–155.
- Steen, H. 2004. Effects of reduced salinity on reproduction and germling development in *Sargassum muticum* (Phaeophyceae, Fucales). *Eur. J. Phycol.* 39:293–299.
- Suárez, A. M. 2005. Lista de las macroalgas marinas cubanas. Rev. Invest. Mar. 26(2):93-148.
- Suárez, A. M., Martínez-Daranas, B. & Alfonso, Y. 2015. Macroalgas marinas de Cuba. Editorial UH, Habana, Cuba, 264 pp.
- Széchy, M. T. & Paula, E. J. de. 2000. Padrões estruturais quantitativos de bancos de Sargassum (Phaeophyta, Fucales) do litoral dos estados do Rio de Janeiro e São Paulo, Brasil. Revta. Brasil. Bot., São Paulo 23(2):121-132
- Tanaka, M. O. 2000. Distribuição espacial e distribuição entre manchas da macrofauna associada à alga parda Sargassum stenophyllum (Mertens) Martius. Ph.D. thesis. Universidade Estadual de Campinas, Campinas, Brazil, 88 pp.
- Tanaka, K., Taino, S., Haraguchi, H., Prendergast, G. & Hiraoka, M. 2012. Warming off southwestern Japan linked to distributional shifts of subtidal canopy-forming seaweeds. *Ecology and Evolution* 2:2854-2865.
- Taskin, E., Öztürk, M., Kurt, O. & Öztürk, M. 2008. *The check-list of the marine algae of Turkey*. Ecem Kirtasiye, Manisa, Turkey, 87 pp.
- Taylor, W. R. 1969. Notes on the distribution of West Indian marine algae particularly in the Lesser Antilles with a bibliography of recent works on Eastern American tropical algae. *Contr. Univ. Mich. Herb.* 9:125-203.

- Thompson, R. C., Crowe, T. P. & Hawkins, S. J. 2002. Rocky intertidal communities: past environmental changes, present status and predictions for the next 25 years. *Environmental Conservation* 29:168–191.
- Torrano-Silva, B. N. 2010. Flora de macrófitas marinhas do Arquipélago de Abrolhos e do Recife de Sebastião Gomes (BA). MsC. dissertation. Universidade de São Paulo, São Paulo, Brazil, 422pp.
- Torrano-Silva, B. N. & Oliveira, E. C. 2013. Macrophytobenthic flora of the Abrolhos Archipelago and the Sebastião Gomes Reef, Brazil. *Continental Shelf Research* 70:150–158.
- Tsuda, R., 1972. Morphological, zonational, and seasonal studies on two species of *Sargassum* on the reefs of Guam. *Proc. Int. Seaweed Symp.* 7:40-44.
- Ugadim, Y. 1973. Algas marinhas bentônicas do litoral sul do estado de São Paulo e do litoral do estado do Paraná, I Divisão Chlorophyta. *Boletim de Botânica da Universidade de São Paulo* 1:11-77.
- Venekey, V., Fonsêca-Genevois, V. G., Rocha, C. M. C. & Santos, P. J. P. 2008. Distribuição espaço-temporal da meiofauna em *Sargassum polyceratium* Montagne (Fucales, Sargassaceae) de um costão rochoso do nordeste do Brasil. *Atlântica* 30(1):53-67.
- Vergés, A., Steinberg, P. D., Hay, M. E., Poore, A. G., Campbell, A. H., Ballesteros, E., Heck Jr.,
 K. L., David J. Booth, D. J., Coleman, M. A., Feary, D. A., Figueira, W., Langlois, T.,
 Marzinelli, E. M., Mizerek, T., Mumby, P. J., Nakamura, Y., Roughan, M., van Sebille, E.
 Gupta, A. S., Smale, D. A., Tomas, F., Wernberg, T & Wilson, S. K. 2014. The
 tropicalization of temperate marine ecosystems: climate-mediated changes in herbivory and
 community phase shifts. *Proceedings of the Royal Society B: Biological Sciences* 281(1789):20140846.
- Vergés, A., Doropoulos, C., Malcolm, H. A., Skye, M., Garcia-Pizá, M., Marzinelli, E. M., Campbell, A. H., Ballesteros, E., Hoey, A. S., Vila-Concejo, A., Bozec, Y.-M. & Steinberg, P. D. 2016. Long-term empirical evidence of ocean warming leading to tropicalization of fish

communities, increased herbivory, and loss of kelp. *Proceedings of the National Academy of Sciences* 113(48):13791-13796.

- Vieira, C., N'Yeurt, A. De R., Rasoamanendrika, F. A., D'Hondt, A., Thi Thram L.-A., Van de Spiegel, S., Kawai, H. & De Clerck, O. 2021. Marine macroalgal biodiversity of northern Madagascar: morpho-genetic systematics and implications of anthropic impacts for conservation. *Biodiversity and Conservation* 30:1501-1546.
- Vroman, M. 1968. The Marine Algal Vegetation of St. Martin, St. Eustatius and Saba. Springer Dordrecht, Netherlands Antilles, 130 pp.
- Vroom, P. S. & Braun, C. L. 2010. Benthic Composition of a Healthy Subtropical Reef: Baseline Species-Level Cover, with an Emphasis on Algae, in the Northwestern Hawaiian Islands. *PLoS ONE* 5(3):e9733.
- Wanders, J. B. W., 1976. The role of benthic algae in the shallow reef of Curaçao (Netherlands Antilles). II. Primary productivity of the *Sargassum* beds on the north-east coast submarine plateau. *Aquat. Bot.* 2:327-335.
- Wang, M., Hu, C., Barnes, B. B., Mitchum, G., Lapointe, B. & Montoya, J. P. 2019. The great Atlantic *Sargassum* belt. *Science* 365:83–87.
- Welten, M. C. M., Audiffred, P. A. J. & Prud'homme van Reine, W. F. 2002. Notes on marine algae collected in Guinea-Bissau, Tropical West Africa. *Botanica Marina* 45:380-384.
- Wernberg, T., Thomsen, M. S., Tuya, F., Kendrick, G. A., Staehr, P. A., & Toohey, B. D. 2010. Decreasing resilience of kelp beds along a latitudinal temperature gradient: Potential implications for a warmer future. *Ecology Letters* 13:685–694.
- Wernberg, T., Smale, D. A., & Thomsen, M. S. 2012. A decade of climate change experiments on marine organisms: Procedures, patterns and problems. *Global Change Biology* 18:1491–1498.

- Wernberg, T., Bennett, S., Babcock, R., de Bettignies, T., Cure, K., Depczynski, M., Dufois, F., Fromont, J., Fulton, C., Hovey, R., Harvey, E., Holmes, T., Kendrick, G., Radford, B., Santana-Garcon, J., Saunders, B., Smale, D., Thomsen, M., Tuckett, C. & Wilson, S. 2016. Climate-driven regime shift of a temperate marine ecosystem. *Science* 353:169-172.
- Wiens, J. J. 2016. Climate-related local extinctions are already widespread among plant and animal species. *PLOS Biology* 14:1–18.
- Wynne, M. J. 2011. A checklist of benthic marine algae of the tropical and subtropical western Atlantic: third revision. *Nova Hedwigia Beihefte* 140(1):7-166.
- Wynne, M. J., Bradshaw, T. & Carrington, C. M. S. 2014. A checklist of the benthic marine algae of Barbados, West Indies. *Botanica Marina* 57(3):167-184.
- Wynne, M. J. 2017. A checklist of benthic marine algae of the tropical and subtropical western Atlantic: fourth revision. *Nova Hedwigia Beiheft* 145:1-202.
- Yan, W., Zhong, Y. & Shangguan, Z. 2017. Contrasting responses of leaf stomatal characteristics to climate change: a considerable challenge to predict carbon and water cycles. *Glob. Chang. Biol.* 23, 3781–3793.
- Yip, Z. T., Quek, Z. B. R., Low, J. K. Y., Wilson, B., Bauman, A. G., Chou, L. M., Todd, P. A. & Huang, D. 2018. Diversity and phylogeny of *Sargassum* (Fucales, Phaeophyceae) in Singapore. *Phytotaxa* 369:200–10.
- Yip, Z. T., Quek, Z. B. R. & Huang, D. 2020. Historical biogeography of the widespread macroalga Sargassum (Fucales, Phaeophyceae). Journal of Phycology 56:300-309.
- Yoshida, T., Suzuki, M. & Yoshinaga, K 2015. Checklist of marine algae of Japan. *Japanese Journal of Phycology* 63:129-189.
- Zarco-Perello, S., Wernberg, T., Langlois, T. J. & Vanderklift, M. A. 2017. Tropicalization strengthens consumer pressure on habitat-forming seaweeds. *Scientific Reports* 7(1):1-8.

- Zarco-Perello, S., Bosch, N. E., Bennett, S., Vanderklift, M. A. & Wernberg, T. 2021. Persistence of tropical herbivores in temperate reefs constrains kelp resilience to cryptic habitats. *Journal of Ecology* 109(5):2081–2094.
- Zayas, C. R., Suárez, A. M. &Ocaña, F. A. 2002. Lista de especies y variación estacional del fitobentos marino de Playa Guardalavaca, Cuba. *Rev. Invest. Mar.* 23(2):81-84.