

**UNIVERSIDADE FEDERAL DE SANTA CATARINA
CENTRO DE CIÊNCIAS BIOLÓGICAS
PROGRAMA DE PÓS-GRADUAÇÃO EM BIOQUÍMICA**

Jade de Oliveira

**EFEITO DA HIPERCOLESTEROLEMIA SOBRE A FUNÇÃO
COGNITIVA E A RELAÇÃO COM A FUNÇÃO
MITOCONDRIAL E ESTRESSE OXIDATIVO
EM CÓRTEX CEREBRAL DE CAMUNDONGOS DEFICIENTES
PARA O RECEPTOR DE LDL**

Dissertação apresentada ao Programa de Pós-Graduação em Bioquímica da Universidade Federal de Santa Catarina como requisito parcial para obtenção do título de Mestre em Bioquímica.

Orientadora: Profª. Dra. Andreza Fabro de Bem

Co-orientador: Profº. Dr. Marcelo Farina

Florianópolis
2012

Catalogação na fonte pela Biblioteca Universitária
da
Universidade Federal de Santa Catarina

048e Oliveira, Jade de

Efeito da hipercolesterolemia sobre a função cognitiva e a relação com a função mitocondrial e estresse oxidativo em córtex cerebral de camundongos deficientes para o receptor de LDL [dissertação] / Jade de Oliveira ; orientadora, Andreza Fabro de Bem. - Florianópolis, SC, 2012.
91 p.: il., graf., tabs.

Dissertação (mestrado) - Universidade Federal de Santa Catarina, Centro de Ciências Biológicas. Programa de Pós-Graduação em Bioquímica.

Inclui referências

1. Bioquímica. 2. Demência. 3. Camundongo como animal de laboratório. 4. Hipercolesterolemia. 5. Estresse Oxidativo.
I. Bem, Andreza Fabro de. II. Universidade Federal de Santa Catarina. Programa de Pós-Graduação em Bioquímica. III.
Título.

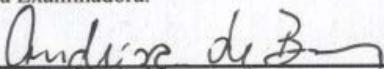
CDU 577

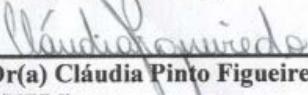
**“Efeito da hipercolesterolemia sobre a função cognitiva e a
relação com a função mitocondrial e estresse oxidativo em
côrtez cerebral de camundongos deficientes para o receptor
de LDL”**
por

Jade de Oliveira

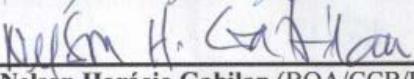
Dissertação julgada e aprovada em sua forma final pelos membros titulares da Banca Examinadora (Port. 05/PPGBQA/2012) do Programa de Pós-Graduação em Bioquímica - UFSC, composta pelos Professores Doutores:

Banca Examinadora:


Prof(a) Dr(a) Andreza Fabro de Bem
(presidente/BQA/CCB/UFSC)


Prof(a) Dr(a) Cláudia Pinto Figueiredo (Faculdade de Farmácia/UFRJ)


Prof(a) Dr(a) Ricardo Castilho Garcez (BEG/CCB/UFSC)


Prof(a) Dr(a) Nelson Horácio Gabilan (BQA/CCB/UFSC)


Prof. Dr. Marcelo Farina
Coordenador do Programa de Pós-Graduação em Bioquímica

Florianópolis, 16 de fevereiro de 2012.

AGRADECIMENTOS

Primeiramente, agradeço minha mãe e meu pai, Jucélia e João, por terem sido o meu exemplo a seguir. Agradeço em especial à minha mãe pelo carinho incondicional em cada momento difícil e o incentivo a nunca desistir dessa jornada.

À meu padrinho Henor, por todo o incentivo, força, compreensão, por acreditar nos meus sonhos, pelo cuidado e amor, principalmente por toda sua dedicação.

À meu irmão Artur, minha cunhada Lucimara e sobrinha Izadora, por todo amor e carinho. À toda a minha família pelo incentivo em todos os momentos.

À meu namorado Thiago, por toda compreensão, paciência e amor incondicional.

Às minhas amigas Francielle e Vanessa, por todos os momentos felizes e apoio.

Um agradecimento especial à minha orientadora Profª. Dra. Andreza Fabro de Bem, por ser um exemplo de profissionalismo e dedicação. Por todo empenho, amizade, auxílio, paciência e incentivo durante todo o mestrado.

Ao meu co-orientador, Prof. Dr. Marcelo Farina, pelo apoio e conhecimentos repassados durante todo o mestrado.

À profª Dra. Alexandra Latini, pela amizade e colaboração na realização deste trabalho.

À profª Dra. Rosa Maria Ribeiro-do-Valle e Prof. Dr. Rui Prediger pela importante colaboração na realização deste trabalho.

Aos colegas que participaram ativamente neste trabalho, Eduardo, Mariana e Vivi, pela amizade, aprendizado e auxílio durante a realização deste trabalho. Em especial ao Eduardo pela parceria, apoio e incentivo durante todo o meu mestrado.

Aos colegas do Laboratório de Bioenergética e Estresse Oxidativo
Aline, Paulo, Fritz, Karina, Roberta, Débora e demais colegas...

Aos colegas de laboratório, Alessandra, Danúbia, Bianca, Gianni,
Dirleise, Renata, Bruna, Luciana e Marcos pela amizade e
companheirismo.

Aos técnicos do LAMEB, Bibiana e Denis, pela amizade,
disponibilidade e auxílio na utilização do espectrofluorímetro.

Ao Prof. Dr. João Batista Teixeira da Rocha da UFSM pelo
fornecimento do Disseleneto de difenila.

Aos professores do curso de Pós-graduação em Bioquímica, pelos
conhecimentos e exemplos passados durante o mestrado.

A todos os amigos, que direta ou indiretamente participaram nesta fase
da minha vida, meu muito obrigada.

À CAPES, CNPq e FAPESC pelo apoio financeiro.

RESUMO

Nos últimos anos, evidências epidemiológicas, clínicas e experimentais indicam a associação entre a hipercolesterolemia e prejuízos cognitivos, como a demência associada à Doença de Alzheimer. Por outro lado, sabe-se que há uma importante participação da disfunção mitocondrial e estresse oxidativo na patogênese das doenças neurodegenerativas e da hipercolesterolemia. Neste contexto, nosso objetivo foi primeiramente avaliar a função cognitiva de camundongos deficientes para o receptor da lipoproteína de baixa densidade ($LDLr^{-/-}$), um modelo de hipercolesterolemia; e sua relação com a função mitocondrial e antioxidante. Para este fim, camundongos controle C57Bl/6 e $LDLr^{-/-}$ foram expostos à uma dieta padrão ou hipercolesterolêmica durante 30 dias e então submetidos ao teste de localização de objeto. Os camundongos $LDLr^{-/-}$ apresentaram prejuízo de aprendizado e memória espacial independentemente da dieta adotada. Além disso, os camundongos $LDLr^{-/-}$ expostos à dieta hipercolesterolêmica apresentaram uma significativa diminuição na atividade dos complexos mitocondriais I e II no córtex cerebral, a qual foi negativamente correlacionada com os respectivos níveis de colesterol plasmático. Este evento foi acompanhado pela diminuição nos níveis de glutationa (GSH), aumento na lipoperoxidação e desequilíbrio na atividade das enzimas integrantes do sistema antioxidante dependente da GSH, glutationa peroxidase (GPx) e glutationa redutase (GR) no córtex cerebral. Estes resultados indicam uma significativa relação entre a hipercolesterolemia, prejuízo cognitivo, e disfunção mitocondrial/estresse oxidativo no córtex cerebral. Considerando que o composto $(PhSe)_2$ vem demonstrando importante papel protetor para doenças cardiovasculares associadas a hipercolesterolemia, analisamos o seu possível efeito neuroprotetor frente ao estresse oxidativo induzido pela hipercolesterolemia no córtex cerebral de camundongos $LDLr^{-/-}$ expostos à dieta hipercolesterolêmica. Nossos resultados demonstraram que o tratamento oral com $(PhSe)_2$ (1 mg/kg) durante 30 dias aumentou significativamente os níveis de GSH e diminuiu a lipoperoxidação no córtex cerebral dos animais hipercolesterolêmicos. Este efeito antioxidante possivelmente está relacionado à sua atividade mimética da GPx. Tomados em conjunto, os resultados apontam este modelo animal de hipercolesterolemia como uma abordagem útil para compreender os eventos moleculares envolvidos na patogênese de doenças neurodegenerativas, bem como o papel neuroprotetor do $(PhSe)_2$.

Palavras-chave: demência, camundongos $LDLr^{-/-}$, hipercolesterolemia, prejuízo cognitivo, disfunção mitocondrial, estresse oxidativo, $(PhSe)_2$.

ABSTRACT

In recent years, epidemiological, clinical and experimental evidence have indicated an association between hypercholesterolemia and cognitive impairment, as well as development of dementia such as Alzheimer's disease. On the other hand, it is known that there is an important involvement of mitochondrial dysfunction and oxidative stress in the pathogenesis of neurodegenerative diseases and hypercholesterolemia. In this context, our objective was to evaluate the cognitive performance in the low density lipoprotein receptor (LDLr)-deficient mice, a model of hypercholesterolemia; and its relationship with mitochondrial and antioxidant functions. Initially, wild type C57Bl/6 or LDLr^{-/-} mice were fed with either standard or cholesterol-enriched diet for a 4-week period and tested for spatial learning and memory in the object location task. LDLr^{-/-} mice displayed spatial learning and memory impairments regardless of diet. Moreover, LDLr^{-/-} mice fed cholesterol-enriched diet presented a significant decrease in the mitochondrial complexes I and II activities in the cerebral cortex, which were negatively correlated with respective blood cholesterol levels. This event was accompanied by decreased in the glutathione (GSH) levels, lipoperoxidation and an imbalance between the peroxide-removing-related enzymes glutathione peroxidase (GPx)/glutathione reductase (GR) activities in the cerebral cortex. These findings indicate a significant relationship between hypercholesterolemia, cognitive impairment, and cortico-cerebral mitochondrial dysfunctional/ oxidative stress. Moreover, considering that the organoselenium compound (PhSe)₂ has shown important protective role in cardiovascular disease associated with hypercholesterolemia, we evaluated the potential neuroprotective effect against hypercholesterolemia-induces oxidative stress in the cerebral cortex of LDLr^{-/-} mice fed with cholesterol-enriched diet. Our results demonstrated that the oral treatment with (PhSe)₂ 1 mg/kg during 30 days significantly increased the GSH levels and decreased the lipoperoxidation in the cerebral cortex of hypercholesterolemic mice. Taken together, the results suggest this mouse model of hypercholesterolemia a useful approach to comprehend the molecular events mediating neurodegenerative diseases pathogenesis. In addition, our results suggested the neuroprotective role of (PhSe)₂.

Key words: dementia, LDLr^{-/-} mice, hypercholesterolemia, cognitive impairment, mitochondrial dysfunction, oxidative stress, (PhSe)₂.

LISTA DE FIGURAS

LISTA DE FIGURAS DA DISSERTAÇÃO

Figura 1	Modelo esquemático do mecanismo pelo qual os receptores de lipoproteína de baixa densidade (LDL) no fígado controlam a produção e o catabolismo de LDL plasmática.....	21
Figura 2	Cadeia respiratória (CR) mitocondrial.....	24
Figura 3	Principais fontes celulares de geração de espécies reativas de oxigênio (ERO).....	25
Figura 4	Principais mecanismos de defesa da glutathiona (GSH) contra o estresse oxidativo.....	28
Figura 5	Sistema antioxidant dependente da GSH.....	29
Figura 6	Liberação de ERO mitocondrial é aumentada em camundongos deficientes de receptor de LDL.....	31
Figura 7	Estrutura química e mecanismo catalítico de organoselêniros.....	33

LISTA DE FIGURAS DO ARTIGO 1

Figura 1	Effects of hypercholesterolemia on object location memory in wild-type C57Bl/6 and LDL ^{r/-} mice treated with standard or cholesterol-enriched diet	40
Figura 2	Plasma cholesterol levels of wild-type C57Bl/6 and LDL ^{r/-} mice treated with standard or cholesterol-enriched diet	40
Figura 3	: Effect of hypercholesterolemia on respiratory chain complexes activities in cerebral cortex homogenates from wild-type C57Bl/6 and LDL ^{r/-} mice treated with standard or cholesterol-enriched diet	41
Figura 4	Effect of hypercholesterolemia on oxidative stress parameters in cerebral cortex homogenates from	

wild-type C57Bl/6 and $\text{LDLr}^{-/-}$ mice treated with standard or cholesterol-enriched diet	42
---	----

LISTA DE FIGURAS DO MANUSCRITO 1

Figura 1 Effects of $(\text{PhSe})_2$ on cerebral cortex oxidative stress parameters in the $\text{LDLr}^{-/-}$ mice fed standard or cholesterol-enriched diet	66
---	----

LISTA DE TABELAS

LISTA DE TABELAS DO MANUSCRITO 1

Tabela 1	Effect of (PhSe) ₂ on plasma total cholesterol levels and non-HDL-cholesterol levels in the LDLr ^{-/-} mice fed standard or cholesterol-enriched diet.....	66
-----------------	--	----

LISTA DE ABREVIATURAS

•NO	Óxido nítrico
•OH	Radical hidroxil
(PhSe) ₂	Disseleeno de difenila
ApoE	Apolipoproteína E
ApoE ^{-/-}	Camundongo deficiente para apolipoproteína E
ATP	Adenosina trifosfato
DA	Doença de Alzheimer
BHE	Barreira hematoencefálica
CR	Cadeia respiratória
eNOS	Óxido nítrico sintase endotelial
ERO	Espécies reativas de oxigênio
FADH ₂	Dinucleotídeo de flavina adenina reduzida
GPx	Glutationa Peroxidase
GR	Glutationa Redutase
GSH	Glutationa reduzida
H ₂ O ₂	Peróxido de hidrogênio
IDL	Lipoproteína de densidade intermediária
LDL	Lipoproteína de baixa densidade
LDL ^{r^{-/-}}	Camundongo deficiente para o receptor de LDL
MDA	Malondialdeído
NADH	Dinucleotídeo de nicotinamida adenina reduzida
NADPH	Dinucleotídeo fosfato de nicotinamida adenina reduzida
Nrf-2	Fator nuclear eritróide-2
O ₂ ^{•-}	Ânion superóxido
ONOO ⁻	Peroxinitrito

Se-OH	Selenol
SNC	Sistema nervoso central
SOD	Superóxido dismutase
TBARS	Substâncias reativas ao ácido tiobarbitúrico
Vad	Demência Vascular
VLDL	Lipoproteína de densidade muito baixa

SUMÁRIO

1	INTRODUÇÃO.....	19
1.1	HIPERCOLESTEROLEMIA.....	19
1.1.1	Hipercolesterolemia e Doenças Neurodegenerativas.....	21
1.2	MITOCÔNDRIA.....	23
1.2.1	Mitocôndria e Doenças Neurodegenerativas.....	25
1.3	ESTRESSE OXIDATIVO E DEFESAS ANTIOXIDANTES.....	26
1.4	DISFUNÇÃO MITOCONDRIAL E HIPERCOLESTEROLEMIA.....	30
1.5	COMPOSTOS ORGÂNICOS DE SELÊNIO.....	32
1.5.1	Disseleneto de Difenila.....	33
2	OBJETIVOS.....	35
2.1	OBJETIVO GERAL.....	35
2.2	OBJETIVOS ESPECÍFICOS.....	35
3	JUSTIFICATIVA.....	36
4	RESULTADOS.....	37
4.1	ARTIGO 1.....	38
4.2	MANUSCRITO 1.....	47
5	DISCUSSÃO.....	67
6	CONCLUSÕES.....	73
7	PERSPECTIVAS.....	74
8	REFERÊNCIAS BIBLIOGRÁFICAS.....	75

1 INTRODUÇÃO

1.1 HIPERCOLESTEROLEMIA

O colesterol é um constituinte lipídico importante de todas as membranas celulares e bainhas de mielina, desempenhando papel crucial para integridade sináptica e função neuronal (Simons e Ikonen, 2000; Pfrieger, 2003). O colesterol também funciona como o substrato para síntese de ácidos biliares no fígado, e como o precursor de hormônios esteróides em tecidos endócrinos (Liscum e Underwood, 1995). Entretanto, apesar do seu papel em processos bioquímicos essenciais e no suporte estrutural de membranas, fortes evidências clínicas e experimentais suportam a estreita ligação entre níveis plasmáticos elevados de colesterol e o desenvolvimento e progressão de lesões ateroscleróticas (Brown e Goldstein, 1986; Stokes et al., 1987).

A hipercolesterolemia, particularmente os níveis elevados de colesterol presente na lipoproteína de baixa densidade (LDL), é um fator de risco bem estabelecido para incidência de atherosclerose e suas complicações patológicas (Ross e Harker, 1976; Libby, 2002). A associação entre a hipercolesterolemia e a atherosclerose foi estabelecida, em parte, com base em estudos sobre a hipercolesterolemia familiar, uma doença autossômica dominante, que têm como característica principal mutações no gene do receptor de LDL. No ano de 1985, os pesquisadores americanos Michael Brown e Joseph Goldstein conquistaram o prêmio Nobel de Medicina por caracterizarem o receptor de LDL e atribuírem à deficiência no gene deste receptor a causa da hipercolesterolemia familiar. Heterozigotos para esta doença manifestam uma elevação de duas a cinco vezes nos níveis plasmáticos de LDL-colesterol, em função de um prejuízo funcional do receptor de LDL, resultando em uma deficiência na depuração sanguínea desta lipoproteína. Indivíduos homozigotos apresentam um aumento de quatro a seis vezes no colesterol plasmático e desenvolvem atherosclerose precocemente (Gotto e Grundy, 1999; Steinberg, 2002) (Figura 1).

Diferentes espécies animais têm sido utilizadas como modelos de hipercolesterolemia e atherosclerose, entre elas destacam-se coelhos e camundongos. A primeira evidência de atherosclerose experimental foi descrita em 1908 por Ignatowski, através da observação de lesões ateroscleróticas em coelhos alimentados com uma dieta rica em proteína animal. Apesar de não desenvolverem atherosclerose espontaneamente, os coelhos são ainda bastante utilizados como modelos de

hipercolesterolemia, por serem altamente responsivos à ingestão de colesterol e desenvolverem lesões em curto prazo. Porém, nesta espécie, os níveis plasmáticos de colesterol são muito elevados e as lesões formadas apresentam conteúdo lipídico e inflamatório muito superior às lesões observadas em humanos (Jawien et al., 2004).

Por sua vez, os camundongos tornaram-se uma espécie extensamente empregada nas investigações biomédicas, e sua utilização para o estudo do processo aterosclerótico apresenta inúmeras vantagens em relação aos demais modelos animais (Daugherty, 2002). No entanto, não há conhecimento de que camundongos do tipo selvagem desenvolvam aterosclerose de maneira espontânea. Além disso, muitas linhagens destes animais respondem fracamente à dieta rica em colesterol e desenvolvem apenas estrias gordurosas no arco aórtico (Paigen et al., 1990). Neste contexto, destaca-se o surgimento de camundongos geneticamente modificados, os quais solucionaram muitos problemas relacionados ao estudo experimental da aterosclerose (Ohashi et al., 2004). Os camundongos com deleção gênica de apolipoproteína E ($\text{apoE}^{-/-}$) ou do receptor de LDL ($\text{LDLr}^{-/-}$) são amplamente empregados na atualidade, e ambos fornecem uma ferramenta prática para o estudo da hipercolesterolemia e suas consequências.

Os camundongos $\text{LDLr}^{-/-}$, desenvolvidos em 1993 por Ishibashi e colaboradores, são um modelo de hipercolesterolemia familiar humana. Estes animais apresentam hipercolesterolemia, caracterizada por níveis moderados de LDL-colesterol, mesmo quando submetidos a uma dieta padrão, podendo desenvolver lesões ateroscleróticas a longo prazo. Ademais, são muito susceptíveis a modificações dietéticas quando alimentados com dieta rica em colesterol, tornando-se severamente hipercolesterolêmicos, com o desenvolvimento de intensa aterosclerose aórtica e xantomas subcutâneos (Ishibashi et al., 1993; Kowala et al., 2000; Daugherty, 2002). Algumas características deste modelo animal podem trazer vantagens para sua utilização, tais como: (1) a semelhança à condição humana de hipercolesterolemia familiar, causada por mutações no gene para o receptor de LDL; (2) o perfil de lipoproteínas plasmáticas, que se assemelha ao de humanos, estando a maior parte do colesterol confinado na fração LDL; e (3) o grau de dislipidemia intermediário, desenvolvendo lesões menos avançadas do que os camundongos $\text{apoE}^{-/-}$ (Zadelaar et al., 2007).

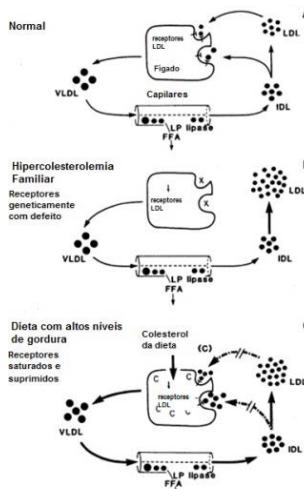


Figura 1. Modelo esquemático do mecanismo pelo qual os receptores de lipoproteína de baixa densidade (LDL) no fígado controlam a produção e o catabolismo de LDL plasmática. (A) em indivíduos normais, (B) em indivíduos com hipercolesterolemia familiar, e (C) em indivíduos consumindo uma dieta rica em gorduras saturadas e colesterol. VLDL, lipoproteína de muito baixa densidade; IDL, lipoproteína de densidade intermediária; LP lipase, lipase lipoprotéica; FFA; ácidos graxos livres (Adaptado de Brown e Goldstein, 1986)

1.1.1 Hipercolesterolemia e Doenças Neurodegenerativas

Evidências epidemiológicas e neuroquímicas suportam a associação entre alterações no metabolismo do colesterol e o aparecimento de prejuízos cognitivos, bem como de demência (Panza et al., 2006). Todavia, os mecanismos moleculares pelos quais os níveis de colesterol contribuem para patofisiologia de doenças neurodegenerativas ainda não estão totalmente elucidados (Wolozin, 2004; Panza et al., 2006; Duron e Hanon, 2008).

A demência é um dos distúrbios neurológicos com maior relevância em idosos e um dos principais problemas de saúde pública. Nas últimas décadas a expectativa de vida está aumentando. Em decorrência deste envelhecimento populacional estima-se que a prevalência global de demência quadruplique de 24,3 milhões em 2001 para 81,1 milhões de indivíduos afetados em 2040 (Ferri et al., 2005). Nos países ocidentais, as formas mais comuns de demência são a

Doença de Alzheimer (DA) e a Demência Vascular (Vad), com respectivas frequências de 70% e 15% entre todas as demências (Whitehouse et al., 1997).

Estudos epidemiológicos longitudinais com pacientes idosos indicam que indivíduos hipercolesterolêmicos durante a idade adulta são mais susceptíveis a desenvolver DA e Vad em idades avançadas (Kivipelto et al., 2001, 2002, 2005), e que quando recebem tratamento para doenças cardiovasculares como terapias que reduzem os níveis de colesterol (por exemplo as estatinas), apresentam uma menor prevalência de demência e diminuição da deterioração cognitiva (Sparks et al., 2005). Todavia, a relação entre hipercolesterolemia e funções cognitivas é mais complexa que uma simples relação linear.

Muitos dos fatores de risco vasculares clássicos, incluindo hipertensão, diabetes mellitus, e em particular a hipercolesterolemia, também são considerados fatores de risco para doenças neurodegenerativas, principalmente a DA (Casserly eTopol, 2004; Shobab et al., 2005; Beach et al., 2007). O grau de comprometimento destes fatores para o desenvolvimento das doenças neurodegenerativas ainda pode ser influenciado por fatores genéticos, como a presença do alelo ε4 da ApoE, que tem um papel bem estabelecido na doença arterial coronariana e no desenvolvimento de aterosclerose, mas também é fortemente associado com a DA (Davignon et al., 1988; Eichner et al., 2002).

A descoberta da disfunção neurovascular como parte integrante da DA, levou a um maior interesse ao que se tornou conhecido como “hipótese vascular” das doenças neurodegenerativas (Kolovou et al., 2002; Luthra et al., 2002). A existência de um componente vascular que reduz a perfusão cerebral tem sido proposta como um possível mecanismo envolvido na patofisiologia da DA (Humpel, 2011). Estudos epidemiológicos evidenciam que há um aumento no risco de desenvolvimento de DA em indivíduos com aterosclerose grave (Hofman et al., 1997). Além disso, estudos demonstraram a relação entre doença arterial coronariana e a severidade de neuropatologias (Beeri et al., 2006). Esse envolvimento da aterosclerose poderia ser explicado pela ocorrência de doenças cerebrovasculares, como o acidente vascular cerebral, doença de pequenos vasos cerebrais, ou ainda ser resultante da hipoperfusão cerebral, situações estas relacionadas com o comprometimento da barreira hematoencefálica (BHE) (de la Torre, 2002, 2004). Sabe-se que a disfunção de células endoteliais que compõem a BHE também tem sido correlacionada com a severidade da DA (Dede et al., 2007).

Corroborando com estas evidências clínicas e epidemiológicas, estudos experimentais em animais utilizando primeiramente coelhos e em seguida camundongos transgênicos modelos de DA alimentados com uma dieta rica em colesterol, demonstraram que a hipercolesterolemia intensifica a patogênese da DA (Sparks et al., 1994; Refolo et al., 2000). De fato, Ullrich e colaboradores (2010) demonstraram que ratos hipercolesterolêmicos apresentam prejuízo de memória e desenvolvem uma patologia com características semelhantes às encontradas na DA. Recentemente, Ramirez e colaboradores (2011) utilizando camundongos ApoB100/LDL $r^{-/-}$ sugeriram a utilização da hipercolesterolemia como um biomarcador chave para o monitoramento de prejuízo cognitivo leve, e propuseram o uso destes camundongos transgênicos como um modelo de declínio cognitivo.

1.2 MITOCÔNDRIA

A mitocôndria é a mais complexa e dinâmica organela celular, indispensável para muitos processos biossintéticos e responsável pela maior produção líquida de energia (DiMauro e Schon, 2003; Spees et al., 2006). Esta organela tem uma estrutura basicamente membranosa, sendo seu envoltório formado por duas membranas, a membrana externa e a membrana interna. A membrana externa é mais permeável que a membrana interna, e entre ambas é determinado um espaço denominado intermembranoso onde ocorrem reações essenciais ao metabolismo celular. A membrana interna é formada por pregas que se expandem no espaço intramitocondrial (matriz mitocondrial) denominadas cristas mitocondriais (Lehninger et al., 2004; Rousset et al., 2004).

A produção energética mitocondrial é resultante de dois processos metabólicos estreitamente coordenados, o ciclo de Krebs e a cadeia transportadora de elétrons ou cadeia respiratória (CR). O ciclo de Krebs tem como função principal produzir as coenzimas NADH e FADH $_2$ para que sejam inseridas na CR; todas as enzimas envolvidas neste ciclo oxidativo se encontram localizadas na matriz mitocondrial (Di Donato, 2000). A mitocôndria produz mais de 90% da energia celular via fosforilação oxidativa (Chance et al., 1979); e essa produção bioenergética assume importância máxima no sistema nervoso central (SNC), devido à limitada capacidade glicolítica das células neuronais e sua alta dependência da fosforilação oxidativa (Moreira et al., 2010).

A fosforilação oxidativa é um processo que requer a ação orquestrada de cinco complexos enzimáticos distribuídos de forma

especial na membrana mitocondrial interna, os quais constituem a CR (Figura 2) (Alberts et al., 2002). Cada um destes complexos é constituído de várias subunidades protéicas que se encontram associados com uma variedade de grupamentos prostéticos com potencial de oxiredução sucessivamente maiores (Lehninger et al., 2004). Durante este processo, NADH e/ou FADH₂ são oxidados provocando a transferência de seus elétrons para o complexo I (NADH desidrogenase) ou complexo II (Succinato desidrogenase), e então para o complexo III (Citocromo c redutase) via coenzima Q (CoQ). O complexo III transfere os elétrons da CoQ para o carreador móvel de elétrons, o citocromo c. O complexo IV (Citocromo c oxidase) é o complexo terminal da cadeia transportadora de elétrons, transferindo os elétrons a partir do ferrocitocromo c para o oxigênio molecular, o acceptor final de elétrons que através da adição de quatro elétrons é reduzido a H₂O (Barrientos et al., 2002; de Moura et al., 2010).

Concomitante com a transferência de elétrons entre os complexos I, III e IV ocorre à translocação de prótons através da membrana mitocondrial interna e a síntese endergônica de ATP, empregando como força motriz a energia armazenada como gradiente eletroquímico de prótons (Babcock e Wikstrom, 1992). Este gradiente determina uma polarização da membrana mitocondrial interna (potencial de membrana mitocondrial); que pode ser revertida pelo fluxo desses prótons através do complexo V (F₁-F₀ ATP sintase) (Figura 2). O fluxo de prótons leva à condensação do ADP e de fosfato inorgânico em ATP, que; por sua vez, é a moeda molecular de transferência de energia em uma célula (Saraste, 1990; Wallace, 1999).

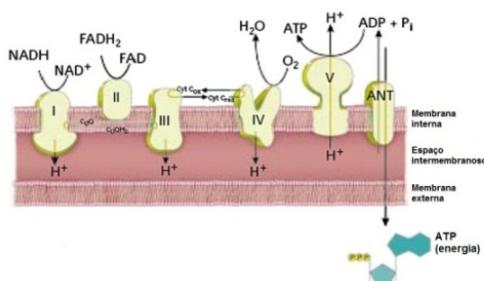


Figura 2. Cadeia respiratória (CR) mitocondrial. Esquema da CR incorporada na membrana interna da mitocôndria. É composto por cinco complexos: complexo I, NADH desidrogenase; complexo II, Succinato desidrogenase; complexo III, Citocromo c redutase, complexo IV, Citocromo c

oxidase; e complexo V, F₁-F₀ ATP sintase. ANT é o transportador de nucleotídeos adenina (Adaptado de Pieczenik e Neustadt, 2007)

Na maioria dos tipos celulares, as mitocôndrias representam as principais fontes de espécies reativas de oxigênio (ERO), mesmo na presença de mecanismos de defesa antioxidantes; a Figura 3 ilustra as principais fontes celulares de ERO. Os ERO são continuamente gerados pela CR, isto porque uma consequência da fosforilação oxidativa é a geração de elétrons desemparelhados, principalmente no complexo I e em menor grau no complexo III. A interação destes elétrons com oxigênio molecular resulta na formação do ânion superóxido (O_2^-), ERO altamente reativa que é rapidamente interconvertida em outras espécies radicais e mediador de reações oxidativas em cadeia. Dismutação de O_2^- produz peróxido de hidrogênio (H_2O_2), o qual pode ser completamente reduzido a H_2O ou parcialmente reduzido ao radical hidroxil (OH^-), um dos mais fortes oxidantes da natureza (Turrens, 2003; Duchen, 2004).

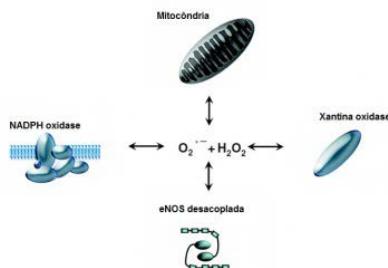


Figura 3. Principais fontes celulares de geração de espécies reativas de oxigênio (ERO). eNOS, Óxido nítrico sintase endotelial; O_2^- , ânion superóxido; H_2O_2 , peróxido de hidrogênio (Adaptado de Ray e Shah, 2005)

1.2.1. Mitocôndria e Doenças Neurodegenerativas

A cognição humana depende, entre outras coisas, da habilidade do SNC em sustentar altas taxas de produção energética continuamente ao longo da vida mantendo um ambiente interno eletroquímico saudável (Dikalov, 2011). Por outro lado, a disfunção mitocondrial e altas concentrações de ERO têm papel importante na patogênese das mais

comuns doenças neurodegenerativas, incluindo a DA e as doenças cerebrovasculares (de Moura et al., 2010). Durante o desenvolvimento e progressão destas doenças neurodegenerativas, devido à hipoperfusão vascular cerebral, as mitocôndrias são danificadas tornando-se incapazes de manter a demanda energética da célula (Hirai et al., 2001), o que resulta em aumento na produção de ERO, interrupção da fosforilação oxidativa; e por fim diminuição dos níveis de ATP, necessários para homeostase energética normal (Schulz et al., 1997). Estes processos estão intimamente associados com morte e degeneração neuronal; danos, os quais podem levar a graves distúrbios neurológicos tais como prejuízo cognitivo e desenvolvimento de demência (Aliiev et al., 2004).

A correlação positiva entre as doenças neurodegenerativas e as doenças cardiovasculares identifica a hipoperfusão vascular cerebral como fator desencadeador das doenças neurodegenerativas (de la Torre, 2008). Estudos experimentais em animais e culturas de células demonstram que as lesões vasculares induzidas pelo baixo fluxo sanguíneo cerebral causam maior geração de ERO mitocondrial, resultando em dano oxidativo e morte às células neuronais. Além disso, há crescentes evidências que a disfunção mitocondrial e a redução da atividade de algumas enzimas chaves mitocondriais, como os complexos respiratórios estão correlacionados com perda neuronal em indivíduos com doenças neurodegenerativas (Aliyev et al., 2005; Aliyev 2009).

1.3 ESTRESSE OXIDATIVO E DEFESAS ANTIOXIDANTES

ERO são geradas tanto em resposta a condições fisiológicas como patológicas (Morgan et al., 2007). O estado redox das células é uma consequência de um crítico balanço entre a produção de ERO e as defesas antioxidantes (Emerit et al., 2004; Halliwell, 2006). A elevação na formação de ERO e/ou o prejuízo dos sistemas de defesas antioxidantes, resulta em estresse oxidativo potencialmente citotóxico (Sies, 1997; Droege, 2002; Turrens, 2003). Sob esta condição pró-oxidante, radicais altamente reativos danificam indiscriminadamente proteínas (Stadtman e Levine, 2000), lipídios (Rubbo et al., 1994), polissacáideos (Kaur e Halliwell, 1994) e DNA (Richter et al., 1988; LeDoux et al., 1999), levando a morte celular.

A reação da ERO com os ácidos graxos poliinsaturados, presentes nas membranas celulares e nas lipoproteínas, inicia um processo em cadeia conhecido como peroxidação lipídica ou lipoperoxidação. As alterações nas membranas celulares devido à lipoperoxidação levam a

transtornos da permeabilidade, alterando o fluxo iônico e o fluxo de outras substâncias, o que resulta na perda da seletividade para entrada e/ou saída de nutrientes e substâncias tóxicas à célula, alterações do DNA, oxidação da LDL e comprometimento dos componentes da matriz extracelular (Barber e Harris, 1994). Os produtos gerados durante as fases do processo de lipoperoxidação também chamados de substâncias reativas ao ácido tiobarbitúrico (TBARS) podem ser avaliados e utilizados como um indicador do estresse oxidativo celular (Niki, 2009).

A produção excessiva de ERO superando os mecanismos de defesas antioxidantes tem sido implicada no desenvolvimento de aterosclerose e doenças cardiovasculares (Harrison et al., 2003). Um excesso de ERO em muitos sistemas celulares, incluindo as células das paredes vasculares e células da circulação sanguínea, é descrito em indivíduos com aterosclerose avançada (Forstermann, 2010). Neste sentido, o aumento do estresse oxidativo está associado com a presença dos fatores de risco cardiovasculares que estão envolvidos na formação das placas ateroscleróticas (Mansego et al., 2011). Portanto, o estresse oxidativo é um elo entre os fatores de risco cardiovasculares e as doenças vasculares. Nesse sentido, um grande número de evidências clínicas e experimentais propõe a associação entre o principal fator de risco cardiovascular: a hipercolesterolemia, e o estresse oxidativo (Csont et al., 2007; Hulsmans e Holvoet, 2010; Drummond et al., 2011).

O SNC é especialmente suscetível ao estresse oxidativo, esta susceptibilidade é devido ao grande conteúdo lipídico altamente peroxidável das bainhas de mielina e à alta taxa de metabolismo oxidativo cerebral (Halliwell, 1992, 2001, 2006). Neste contexto, o estresse oxidativo tem sido implicado em muitos mecanismos de neurotoxicidade, desempenhando papel importante em diferentes patologias neurodegenerativas. Elevados níveis de dano oxidativo ao DNA são encontrados em tecidos de pacientes com prejuízos cognitivos (Mao e Reddy, 2011), bem como tecidos post-mortem apresentam níveis aumentados de estresse oxidativo celular e perda sináptica em determinadas regiões cerebrais relacionadas com doenças neurodegenerativas (Hensley et al., 1995; Halliwell, 2006).

Em condições fisiológicas os níveis basais de ERO gerados são rapidamente eliminados por sistemas antioxidantes: superóxido dismutase (SODs), catalase, glutationa peroxidases (GPxs), glutationa redutase (GR), glutationa (GSH), vitamina E e C, tiol peroxidases entre outros (Dirnagl et al., 2003; Rhee et al., 2005). A GSH desempenha papel fundamental na proteção celular contra o estresse oxidativo, principalmente no SNC, exercendo suas funções através de mecanismos

diversos (Figura 4 apresenta as principais funções da GSH). Esta molécula efetivamente sequestra ERO, antes que estes iniciem as reações. Além disso, o sistema antioxidante dependente da GSH é o principal sistema antioxidante endógeno, desempenhando assim, papel central na proteção celular contra oxidantes. Neste ciclo (Figura 5), a GSH serve como um cofator essencial, trabalhando como doador de equivalentes redutores para a detoxificação de H_2O_2 ou outros peróxidos e produtos de peroxidação catalisada pela GPx. Neste processo, H_2O_2 é reduzido a H_2O pela reação da GPx com a GSH, a qual é oxidada a GSH dissulfeto (GSSG). GSSG é então reduzida novamente a GSH, pela ação da flavoenzima GR à custa da oxidação do NADPH (Dringen et al., 1999; Maher, 2005; Aoyama et al., 2008).

A mitocôndria possui 10% da massa celular, entretanto seu conteúdo de GSH é comparável ao restante do conteúdo presente na célula. O transporte do GSH citosólico para a matriz mitocondrial é o único determinante do conteúdo de GSH nesta organela (Lash, 2006). A depleção e/ou oxidação de GSH mitocondrial está associada a diversos estados patológicos, tais como, doenças cardiovasculares e distúrbios neurodegenerativos; sendo o estado redox da GSH crítico para que haja uma adequada função mitocondrial, uma vez que a GSH preserva a integridade de proteínas e lipídeos mitocondriais e controla a propagação de ERO mitocondrial (Circu e Aw, 2008).



Figura 4. Principais mecanismos de defesa da glutationa (GSH) contra o estresse oxidativo. GSH é o principal antioxidante no SNC, o qual não enzimaticamente reage com ânion superóxido (O_2^-), óxido nítrico ($^{\bullet}NO$), radical hidroxil (OH^{\bullet}), e peroxinitrito ($ONOO^-$). GSH também reage com peróxido de hidrogênio (H_2O_2) ou outros peróxidos através da reação catalisada pela glutationa peroxidase (GPx)/ catalase (Adaptado de Aoyama et al., 2008)

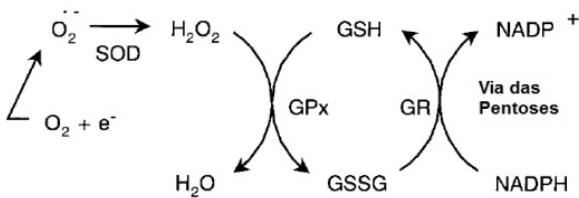


Figura 5. Sistema antioxidante dependente da glutatona (GSH). O ânion superóxido (O_2^-) sob a ação da superóxido dismutase (SOD) é convertido em peróxido de hidrogênio (H_2O_2), o qual é reduzido à água pela ação do sistema antioxidante dependente da GSH. GPx, Glutatona peroxidase; GR, Glutatona redutase; GSSG, Glutatona oxidada (Adaptado de O'Donovan e Fernandes, 2000)

As GPxs compreendem uma família de enzimas bem conhecidas por serem importantes componentes do sistema de defesa antioxidante humano. Nos seres humanos, foram descritas até o presente momento cinco tipos de GPxs: citóslica (GPx-1), gastrointestinal (GPx-2), plasmática (GPx-3), fosfolipídica (GPx-4), e a GPx do epitélio olfativo e tecido embriônico específica (GPx-6) (Kryukov et al., 2003). As GPx1-3 catalisam a redução de H_2O_2 e hidroperóxidos orgânicos, enquanto que a GPx-4 pode reduzir diretamente os hidroperóxidos de fosfolipídios e colesterol (Ursini et al., 1999).

Estudos clínicos recentes têm demonstrado que a diminuição da atividade da enzima GPx-1 eritrocitária está associada ao risco aumentado de eventos cardiovasculares (Blankenberg et al., 2003; Espinola-Klein et al., 2007). Além disso, a atividade da GPx-1 está diminuída ou ausente em placas ateroscleróticas humanas e está associada ao desenvolvimento de lesões mais severas (Lapenna et al., 1998). Em camundongos, a deficiência de GPx-1 causa disfunção endotelial, acompanhada de aumento do estresse oxidativo, anormalidades funcionais e estruturais do sistema cardiovascular (Forgione et al., 2002), além de acelerar a progressão da lesão aterosclerótica em camundongos apoE^{-/-} (Torzewski et al., 2007). De particular interesse, um recente estudo demonstrou que a atividade da GPx, que é crucial para a detoxificação do H_2O_2 no SNC, está diminuída em pacientes com DA quando comparados com indivíduos normais (Dringen et al., 2005; Kharrazi et al., 2008).

1.4 DISFUNÇÃO MITOCONDRIAL E HIPERCOLESTEROLEMIA

Alterações na função mitocondrial estão intimamente ligadas com doenças metabólicas. Recentes estudos propõem a relação entre hipercolesterolemia e alterações nas funções mitocondriais, tais como metabolismo energético, transporte de íons e estado redox (Vercesi et al., 2007). Logo, um dos possíveis mecanismos pelo qual a hipercolesterolemia induz estresse oxidativo pode estar relacionado à perturbação da função mitocondrial (McCommis et al., 2011).

Oliveira e colaboradores (2005) demonstraram que as mitocôndrias de tecidos isolados (cérebro, fígado e rim) de camundongos $LDLr^{-/-}$ hipercolesterolêmicos produzem taxas mais elevadas de ERO que mitocôndrias de controles. Em contraste as mitocôndrias dos controles, as mitocôndrias dos camundongos hipercolesterolêmicos não foram capazes de sustentar NADPH no estado reduzido. Este baixo conteúdo de nucleotídeos reduzidos nos hepatócitos dos camundongos $LDLr^{-/-}$ pode ser resultado da alta taxa de lipogênese, uma vez que estas células são deficientes em captar o colesterol exógeno. Os processos de lipogênese/ esteroidogênese consomem grande quantidade de equivalentes redutores do NADPH, uma vez que a biossíntese de um mol de colesterol oxida 24 mols de NADPH. De fato, a taxa de secreção hepática de triglicerídeos e colesterol *in vivo* foram duas vezes maior nos camundongos $LDLr^{-/-}$ que em camundongos controles. Além disso, a síntese *de novo* de colesterol e de outros lipídeos foi maior no fígado de camundongos $LDLr^{-/-}$ (Oliveira et al., 2005).

Uma das principais fontes de NADPH mitocondrial em animais é a isocitrato desidrogenase. Interessantemente, níveis endógenos mitocondriais de isocitrato e outros intermediários do ciclo de Krebs estão diminuídos no fígado de camundongos $LDLr^{-/-}$ e a suplementação das mitocôndrias destes camundongos com isocitrato efetivamente reverteu a falta de NADPH e o aumento de liberação de ERO observados; bem como aumentou o consumo de oxigênio nestas mitocôndrias. Ademais, a suplementação dos camundongos $LDLr^{-/-}$ *in vivo* com citrato, mostrou uma melhora na capacidade destes animais sustentarem o estado reduzido do NADPH, parcialmente revertendo a disfunção mitocondrial do fígado destes camundongos. Portanto, o estresse oxidativo em camundongos $LDLr^{-/-}$ hipercolesterolêmicos é, em parte, resultado da depleção dos substratos relacionados ao NADPH e de quantidades insuficientes de equivalentes redutores para

reconstituir os sistemas enzimáticos antioxidantes mitocondriais (Paim et al., 2008).

Este desequilíbrio redox mediado pela mitocôndria pode ser uma importante etapa na patogênese de muitas doenças que cursam com aumento de lipogênese e hipercolesterolemia, como a aterosclerose. O defeito no receptor de LDL leva a dois efeitos pró-aterogênicos: níveis aumentados de substratos oxidáveis extracelulares (LDL) e um desequilíbrio nos processos celulares redox. O último fenômeno é responsável por estresse oxidativo local que desencadeia a oxidação de lipoproteínas, o que por sua vez induz dano mitocondrial. O ciclo vicioso resultante leva à morte celular e progresso da aterogênese (Figura 6) (Vindis et al., 2005; Zmijewski et al., 2005; Vercesi et al., 2007).

Neste sentido, acreditamos que a disfunção mitocondrial e o consequente estresse oxidativo desencadeados pela hipercolesterolemia possam desempenhar importante papel nos prejuízos cognitivos e processos neurodegenerativos.

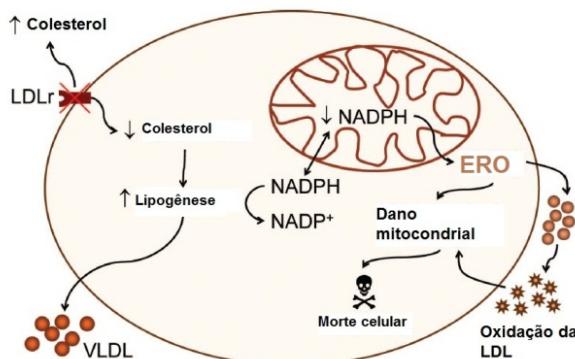


Figura 6: Liberação de espécies reativas de oxigênio (ERO) mitocondrial é aumentada em camundongos deficientes para o receptor de lipoproteína de baixa densidade ($LDLr^{-/-}$). A deficiência dos receptores de LDL resulta em deficiência de transporte de colesterol para dentro da célula, estimulando a lipogênese intracelular e secreção de lipoproteína de muito baixa densidade (VLDL), aumentando os níveis plasmáticos de lipídios. NADPH é utilizado para a lipogênese, resultando em diminuição da razão de $NADPH/NADP^+$ citoplasmática e mitocondrial. Como muitos dos sistemas de remoção de ERO dependem de NADPH como uma fonte redox, ERO mitocondrial ficam

acumuladas e são liberados em níveis elevados. A produção aumentada de ERO nestas condições pode contribuir para dano oxidativo tecidual, oxidação da LDL e aterosclerose (Adaptado de Vercesi et al., 2007)

1.4 COMPOSTOS ORGÂNICOS DE SELÊNIO

O selênio é um elemento traço essencial, componente estrutural de enzimas com atividades antioxidantes, particularmente das isoformas da GPx (Flohe et al., 1973). Estas enzimas têm importantes papéis na defesa celular, protegendo contra processos oxidativos pela detoxificação de hidroperóxidos de hidrogênio ou lipídicos (Sies e Arteel, 2000; Klotz e Sies, 2003). Diferentes classes de compostos orgânicos de selênio exibem atividade mimética da GPx e decompõem H₂O₂ e hidroperóxidos orgânicos utilizando GSH ou outros tióis como doadores de hidrogênio (Wilson et al., 1989; Nogueira et al., 2004). A partir dos anos 80, o interesse em bioquímica, farmacologia e toxicologia de organoselêniros aumentou de maneira significativa devido à variedade de atividades biológicas desempenhadas por esta classe de compostos (Nogueira e Rocha, 2011).

O primeiro exemplo desta classe de compostos foi o ebselen (Parnham e Kindt, 1984). O ebselen (Figura 7A) (2-fenil-1,2-benzilsosenazol-3(2H)-ona) é um composto orgânico de selênio cujas propriedades antioxidantes e anti-inflamatórias têm merecido destaque no campo da farmacologia. Este composto foi descrito e caracterizado como um mimético da enzima GPx na década de 80 (Muller et al., 1984), entretanto, apenas a partir da década de 90, cresceu enormemente o número de trabalhos demonstrando seus efeitos protetores em diferentes tipos celulares e para os mais diversos tipos de injúria.

Recentemente, um estudo utilizando camundongos apoE^{-/-} mostrou o efeito antiaterogênico do ebselen na aterosclerose associada à hiperglicemia (Chew et al., 2009). Outros trabalhos verificaram que o ebselen restaura a função endotelial em ratos diabéticos (Brodsky et al., 2004) e diminui as lesões ateroscleróticas em camundongos transgênicos com expressão aumentada da NADPH oxidase (Khatri et al., 2004). De particular importância, ebselen têm demonstrado ser neuroprotetor em estudos pré-clínicos e clínicos e em modelos animais de uma variedade de condições neuropatológicas (Saito et al., 1998; Davalos, 1999; Porciuncula et al., 2001; Centuriao et al., 2005; Yamagata et al., 2008; Yin et al., 2011).

1.4.1 Disseleneto de Difenila

O disseleneto de difenila ($(\text{PhSe})_2$) (Figura 7B), assim como o ebselen, é um composto orgânico de selênio que reage eficientemente com hidroperóxidos e peróxidos orgânicos, através de reação similar a catalisada pela GPx. Todavia, o $(\text{PhSe})_2$ demonstrou ser mais ativo como mimético da GPx (Wilson et al., 1989) e menos tóxico em roedores que o ebselen (Meotti et al., 2003; Nogueira et al., 2003a).

O mecanismo catalítico para a detoxificação de peróxidos pelo $(\text{PhSe})_2$ foi proposto. A atividade tiol-peroxidase do $(\text{PhSe})_2$ parece ser cineticamente semelhante a reação catalisada pela enzima. Inicialmente o $(\text{PhSe})_2$ reage com um grupamento tiol (RSH) (por exemplo GSH) originando um selenilsulfeto, o qual reage com um segundo equivalente de GSH formando um selenol (Se-H), este selenol reduz o H_2O_2 ou peróxidos lipídicos em H_2O , fechando assim o ciclo catalítico (Figura 7C) (Nogueira e Rocha, 2010).

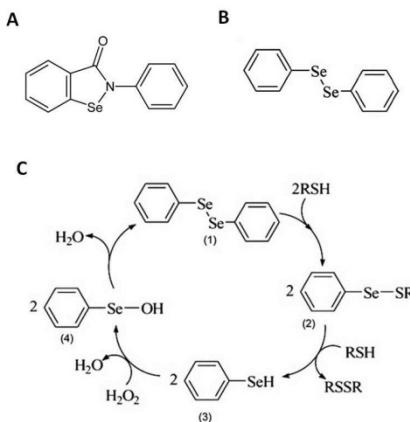


Figura 7. Estrutura química e mecanismo catalítico de organoselênios. Estrutura química do (A) ebselen e do (B) disseleneto de difenila ($(\text{PhSe})_2$). (C) Mecanismo catalítico do $(\text{PhSe})_2$ para a detoxificação de peróxido de hidrogênio (H_2O_2). O $(\text{PhSe})_2$ reage com um grupamento tiol (ex. glutationa (GSH)) originando um selenopersulfato, o qual reage com um segundo equivalente de GSH formando um selenol (Se-H). Finalmente, este selenol reduz o H_2O_2 ou peróxidos lipídicos liberando uma molécula de H_2O , originando um ácido selenínico (Se-OH). Este Se-OH, por sua vez, libera uma molécula de H_2O , fechando assim o ciclo catalítico (Adaptado de Nogueira et al., 2004)

Uma das primeiras evidências das propriedades farmacológicas do $(\text{PhSe})_2$ ocorreu em 2003, quando Nogueira e colaboradores demonstraram as atividades anti-inflamatória e antinociceptiva deste composto em roedores (Nogueira et al., 2003b). Desde então, estudos têm relatado o importante papel protetor deste composto em uma variedade de modelos experimentais associados à produção exacerbada de ERO e estresse oxidativo tais como, inflamação, diabetes, neurotoxicidade e hepatotoxicidade (Ghisleni et al., 2003; Burger et al., 2004; Meotti et al., 2004; Borges et al., 2005), além de diversas outras propriedades farmacológicas.

Nosso grupo de pesquisa tem demonstrado que o $(\text{PhSe})_2$ possui importantes propriedades farmacológicas, que o tornam uma molécula interessante no manejo de doenças cardiovasculares. De Bem e colaboradores (2009) demonstraram que o tratamento por via oral com $(\text{PhSe})_2$ reduz a hipercolesterolemia e o estresse oxidativo em coelhos alimentados com uma dieta rica em colesterol. O $(\text{PhSe})_2$ também foi capaz de inibir a oxidação da LDL humana isolada *in vitro* e este efeito foi relacionado com a sua atividade tiol-peroxidase (de Bem et al., 2008). Além disso, verificamos que este composto foi capaz de reduzir a formação de lesões ateroscleróticas em camundongos $\text{LDLr}^{-/-}$ e diminuir a formação de células espumosas, produção de mediadores inflamatórios e ERO em macrófagos expostos a LDL oxidada (Hort et al., 2011).

A atividade deste composto no SNC tem sido estudada, uma vez que este composto tem característica lipofílica e pode atravessar a BHE, levando ao aumento dos níveis de selênio no cérebro após tratamentos agudos ou crônicos (Jacques-Silva et al., 2001; Maciel et al., 2003). De fato, o $(\text{PhSe})_2$ demonstrou ter atividade neuroprotetora em inúmeros estudos, tais como o de Posser e colaboradores (2008) que demonstraram efeitos protetores do $(\text{PhSe})_2$ *in vitro* contra dano oxidativo induzido por H_2O_2 em fatias hipocampais, e o de da Silva e colaboradores (2011), onde o $(\text{PhSe})_2$ reverteu dano oxidativo e disfunção mitocondrial em cérebros de camundongos expostos ao acetaminofeno. Além disso, administração sistêmica de $(\text{PhSe})_2$ em camundongos melhorou propriedades cognitivas, facilitando a memória de longa duração no teste de reconhecimento de objeto (Rosa et al., 2003).

Diante das importantes propriedades farmacológicas descritas para o $(\text{PhSe})_2$, bem como da participação do estresse oxidativo nos processos neurodegenerativos, hipotetizamos que este composto possa ser uma molécula neuroprotetora efetiva contra danos oxidativos cerebrais induzidos pela hipercolesterolemia em camundongos $\text{LDLr}^{-/-}$.

2 OBJETIVOS

2.1 OBJETIVO GERAL

O objetivo geral deste trabalho foi investigar os efeitos da hipercolesterolemia sobre parâmetros comportamentais e bioquímicos em córtex cerebral de camundongos $LDLr^{-/-}$. Ademais, avaliamos o potencial efeito neuroprotetor de um composto mimético da GPx, $(PhSe)_2$, neste modelo experimental.

2.2 OBJETIVOS ESPECÍFICOS

- Investigar os efeitos da hipercolesterolemia sobre a função cognitiva avaliada através do teste comportamental localização do objeto em camundongos $LDLr^{-/-}$ alimentados com dieta padrão ou hipercolesterolêmica;
- Investigar os efeitos da hipercolesterolemia sobre a função mitocondrial em córtex cerebral de camundongos $LDLr^{-/-}$ alimentados com dieta padrão ou hipercolesterolêmica;
- Investigar os efeitos da hipercolesterolemia sobre parâmetros de estresse oxidativo em córtex cerebral de camundongos $LDLr^{-/-}$ alimentados com dieta padrão ou hipercolesterolêmica;
- Investigar os efeitos neuroprotetores do composto orgânico de selênio $(PhSe)_2$, contra estresse oxidativo induzido pela hipercolesterolemia em córtex cerebral de camundongos $LDLr^{-/-}$ alimentados com dieta hipercolesterolêmica.

3 JUSTIFICATIVA

Nos próximos anos, devido ao aumento da expectativa de vida e consequente “envelhecimento populacional”, enfrentaremos um grande incremento na prevalência de prejuízos cognitivos bem como de demência associadas à idade tal como a DA. Estudos epidemiológicos demonstram um aumento exponencial na prevalência de demência em indivíduos acima de 70 anos, chegando a 45% em indivíduos com 95 anos ou mais. Desse modo uma melhor compreensão acerca dos mecanismos envolvidos nos processos de neurodegeneração e busca por novos alvos terapêuticos torna-se relevante.

Um elevado número de estudos epidemiológicos e bioquímicos suporta a associação entre os fatores de risco cardiovasculares, particularmente a hipercolesterolemia com a predisposição a estes distúrbios neurológicos. Sabe-se que a hipercolesterolemia está relacionada com aumento do estresse oxidativo, e o principal mecanismo proposto para esse aumento de produção de ERO é a disfunção mitocondrial. Ademais, a disfunção mitocondrial e altas concentrações de ERO têm papel importante na patogênese das mais comuns doenças neurodegenerativas, incluindo a DA e as doenças cerebrovasculares. A relação entre tais eventos e como os mesmos contribuem na patogênese das doenças neurodegenerativas ainda são desconhecidos.

O estudo do eventual potencial neuroprotetor do composto orgânico de selênio (PhSe_2) é de grande importância, tendo em vista que várias propriedades farmacológicas já foram descritas para este composto. Nossos trabalhos anteriores comprovam sua propriedade anti-inflamatória e antioxidante, ação hipocolesterolêmica e sua capacidade de inibir a oxidação de LDL humana isolada *in vitro* e o estresse oxidativo em coelhos hipercolesterolêmicos, bem como sua capacidade de reduzir as lesões ateroscleróticas em camundongos $\text{LDLr}^{-/-}$. Além disso, outros estudos apontarem esta molécula como um efetivo agente neuroprotetor. Baseado nestas evidências, acreditamos na possibilidade futura deste composto orgânico de selênio como eficiente agente neuroprotetor em patologias relacionadas ao estresse oxidativo, em especial as decorrentes da hipercolesterolemia.

4 RESULTADOS

Os resultados que fazem parte desta dissertação estão apresentados sob a forma de artigo e manuscrito em fase de redação final, os quais encontram-se aqui organizados.

Os itens Materiais e Métodos, Resultados, Discussão dos Resultados e Referências Bibliográficas, encontram-se nos próprios artigo e manuscrito.

4.1 ARTIGO 1

Positive correlation between elevated plasma cholesterol levels and cognitive impairments in LDL receptor knockout mice: relevance of cortico-cerebral mitochondrial dysfunction and oxidative stress
Artigo publicado no periódico: Neuroscience, 2011.

POSITIVE CORRELATION BETWEEN ELEVATED PLASMA CHOLESTEROL LEVELS AND COGNITIVE IMPAIRMENTS IN LDL RECEPTOR KNOCKOUT MICE: RELEVANCE OF CORTICO-CEREBRAL MITOCHONDRIAL DYSFUNCTION AND OXIDATIVE STRESS

J. DE OLIVEIRA,^a M. A. HORT,^a E. L. G. MOREIRA,^b
V. GLASER,^a R. M. RIBEIRO-DO-VALLE,^b
R. D. PREDIGER,^b M. FARINA,^a A. LATINI^a AND
A. F. DE BEM^{a*}

^aDepartamento de Bioquímica, Universidade Federal de Santa Catarina, 88040–900, Florianópolis, SC, Brazil

^bDepartamento de Farmacologia, Universidade Federal de Santa Catarina, 88049–900, Florianópolis, SC, Brazil

Abstract—Convergent epidemiological, clinical, and experimental findings indicate that hypercholesterolemia contributes to the onset of Alzheimer's disease (AD)-like dementia, but the exact underlying mechanisms remains unknown. In this study, we evaluated the cognitive performance of mice submitted to a model of hypercholesterolemia, as well as its relationship with mitochondrial dysfunction and oxidative stress, two key events involved in AD pathogenesis. Wild-type C57BL/6 or low density lipoprotein receptor (LDLr)-deficient mice were fed with either standard or cholesterol-enriched diet for a 4-week period and tested for spatial learning and memory in the object location task. LDLr^{-/-} mice displayed spatial learning and memory impairments regardless of diet. Moreover, LDLr^{-/-} mice fed cholesterol-enriched diet presented a significant decrease in the mitochondrial complexes I and II activities in the cerebral cortex, which were negatively correlated with respective blood cholesterol levels. Additionally, hypercholesterolemic LDLr^{-/-} mice presented a significant decrease in glutathione levels, about 40% increase in the thiobarbituric acid-reactive substances levels, as well as an imbalance between the peroxide-removing-related enzymes glutathione peroxidase/glutathione reductase activities in the cerebral cortex. These findings indicate a significant relationship between hypercholesterolemia, cognitive impairment, and cortico-cerebral mitochondrial dysfunction/oxidative stress. Because of the involvement of such alterations in AD patients, our data render this mouse model of hypercholesterolemia a useful approach to comprehend the molecular events mediating AD pathogenesis. © 2011 IBRO. Published by Elsevier Ltd. All rights reserved.

Key words: Alzheimer's disease, low density lipoprotein receptor (LDLr)-deficient mice, hypercholesterolemia, cognition

* These authors contribute equally to this work.

^aCorresponding author. Tel.: +55-48-3721-6656; fax: +55-48-33379672. E-mail address: andrezadebem@ccb.ufsc.br (A. Fabro de Bem PhD). Abbreviations: AD, Alzheimer's disease; ANOVA, analysis of variance; EDTA, ethylenediaminetetraacetic acid; ECTA, ethylene glycol tetraacetic acid; GPx, glutathione peroxidase; GR, glutathione reductase; GSH, glutathione; GSSG, oxidized glutathione; LDLr, low density lipoprotein receptor; NADH, beta-nicotinamide adenine dinucleotide reduced dipotassium salt; NADPH, beta-nicotinamide adenine dinucleotide phosphate sodium salt; ROS, reactive oxygen species; TBARS, thiobarbituric acid-reactive substance; TC, total cholesterol.

0306-4522/\$ - see front matter © 2011 IBRO. Published by Elsevier Ltd. All rights reserved.
doi:10.1016/j.neuroscience.2011.09.009

cognitive impairment, mitochondrial dysfunctional, oxidative stress.

There is increasing evidence linking abnormal cholesterol metabolism with cognitive impairments, especially in Alzheimer's disease (AD) (Herrmann and Knapp, 2002; Rojo et al., 2006). Cholesterol is essential for building and maintaining cell membranes; nevertheless, hypercholesterolemia is associated with negative health outcomes, especially those related to vascular disease (Stary, 1989; Sharrett et al., 2006).

Clinical studies have indicated that individuals with hypercholesterolemia are more prone to develop AD (Evans et al., 2000; Yaffe et al., 2002), and a previous case-control study showed a protective effect of lipid-lowering agents on the incidence of dementia (Cramer et al., 2008). Furthermore, experimental studies using New Zealand white rabbits (Sparks et al., 2000) and transgenic mouse models of AD (Rehfeld et al., 2000; Levin-Allerhand et al., 2002) have demonstrated that diet-induced hypercholesterolemia could enhance brain amyloid-beta (A β) protein accumulation. However, the exact mechanisms underlying the accumulation of cholesterol increasing the susceptibility to dementia are still not fully understood.

The mitochondria represent a biologically important source and target for reactive oxygen species (ROS), mainly in the CNS (Kowaltowski and Vercesi, 1999; Brookes et al., 2004). The rate of ROS production is modulated by mitochondrial energetic state and is favored by high membrane potential values (Cadenas and Davies, 2000). ROS production is also largely increased in cases of respiratory chain inhibition, as observed in mitochondrial disease, or in experimental and animal models of oxidative phosphorylation (OXPHOS) deficiencies (Leonard and Schapira, 2000; Fontanesi et al., 2009). The CNS is especially susceptible to ROS-induced damage because of its greater oxygen availability and consumption, high levels of membrane polyunsaturated fatty acids susceptible to oxidative damage, its relatively low levels of antioxidant defenses, and high content of redox metals (Valko et al., 2007). In this way, the concept that mitochondrial damage and dysfunction are relevant in chronic, age-related diseases is not new (Ballinger, 2005). Mitochondrial dysfunction is one of the earliest and most prominent features in hypercholesterolemia (Madamanchi and Runge, 2007; Vercesi et al., 2007) and AD (Hauptmann et al., 2009). Moreover, recent findings in the field support an involve-

ment of mitochondrial-dependent mechanisms in the pathogenesis of both the hypercholesterolemia and AD. One useful tool for the study of the impact of high circulating cholesterol levels on metabolic and functional parameters in different organs is the use of the hypercholesterolemic LDL receptor knockout mice ($LDLr^{-/-}$), which represent a model of familial hypercholesterolemia (FH), a major autosomal dominant disorder associated with increased risk of premature coronary heart disease (Zadelka et al., 2007). Recent studies provided evidence that mitochondria from various tissues from the hypercholesterolemic $LDLr^{-/-}$ mice generate higher amounts of ROS when compared to those from wild-type mates, thus suggesting that mitochondrial ROS may be involved in the early steps of atherogenesis in this model (Oliveira et al., 2005; Paim et al., 2008).

In the present study, we aimed to investigate the effects of hypercholesterolemia on cognitive function as well as its relationship with mitochondrial function and oxidative stress condition in cerebral cortex of $LDLr^{-/-}$ mice fed with a standard or cholesterol-enriched diet.

EXPERIMENTAL PROCEDURES

Animals

Wild-type C57bl/6 and low density lipoprotein receptor knockout ($LDLr^{-/-}$) mice were obtained from Universidade Estadual de Campinas (UNICAMP, São Paulo, Brazil), by homologous recombination, as previously described by Ishibashi et al. (1993). The progenitors were purchased from Jackson Laboratory (Bar Harbor, ME, USA). The animals were kept at $21 \pm 2^\circ\text{C}$ under a 12-h light/12-h dark cycle with free access to food and water. Efforts were made to minimize the number of animals used and their suffering. All procedures used in the present study complied with the guidelines on animal care of the local Ethics Committee on the Use of Animals (CEUA/UFGSC), which follows the NIH publication "Principles of Laboratory Animal Care." Process number: 23080.04093/2010-28.

Experimental protocol

Male wild-type C57bl/6 mice (3-month old; 24–26 g) fed with a standard commercial diet (Nuvilab CR-1, Nuvital Nutrientes S/A, Paraná, Brazil) were used as control. Male $LDLr^{-/-}$ mice (3-month old; 24–26 g) were randomly divided into two experimental groups ($n=6$ –8 animals per group for the behavioral task and $n=5$ animals per group for the biochemical analyses), which were fed either a standard or a cholesterol-enriched chow (20% fat, 1.25% cholesterol, 0.5% cholic acid) as described by Hort et al. (2011). After 30 days, mice were tested on a cognitive paradigm (object location task), and after an overnight food deprivation, the blood was collected from the ocular plexus for posterior determination of total cholesterol (TC) levels. The animals were then euthanized by decapitation, their brains were immediately removed from the skull, and the cerebral cortices were dissected for the posterior biochemical analyses.

Object location task. The experimental apparatus used in this study was an open-field box (50 cm wide × 50 cm deep × 40 cm high) made of transparent polyvinyl chloride (PVC), placed in a dimly lit (7 lx) and sound-isolated room. Identical plastic rectangles (4 cm high × 4.5 cm wide) were used as objects.

The protocol was based on the previous studies described by Assini et al. (2009). The mice were placed in the center of the apparatus with two identical objects for 5 min. The objects were

placed 7 cm away from the walls of the open field. Exploration of the objects was timed by a stopwatch when mice sniffed, whisked, or looked at the objects from no more than 1 cm away. After the training phase, one object was removed from the apparatus for 180 min. After the delay, one object was moved to a new location. The time spent exploring the objects in new (novel) and old (familiar) locations was recorded during 5 min. Sessions were recorded and later analyzed. All locations for the objects were counterbalanced among the groups. After each trial, the experimental apparatus was cleaned only with dry paper, thus ensuring that it was saturated with the smell of the animals. In order to analyze the cognitive performance, a location index was calculated as previously described by Murali et al. (2007): $(\text{Novel} \times 100)/(\text{Novel} + \text{Familiar})$, where Novel is the time spent exploring the displaced object and Familiar is the time spent exploring the non-displaced object (Murali et al., 2007).

Determination of TC levels. TC was measured in plasma using enzymatic kit according to the manufacturer's instructions (Gold Analytics Diagnóstica Ltda., Minas Gerais, Brazil).

Cerebral cortex supernatant preparation. For the analyses of oxidative stress parameters, cerebral cortex was homogenized in 20 mM phosphate buffer (pH 7.4). Homogenates were centrifuged at $1000 \times g$ for 10 min at 4°C to discard nuclei and cell debris. The pellet was discarded and the supernatant, a suspension of mixed and preserved organelles, including mitochondria, was separated and immediately used for the analyses.

For the measuring of respiratory chain complex activities, mitochondrial suspensions from cerebral cortex were prepared. Briefly cerebral cortex samples were homogenized in 10 volumes (1:10, w/v) of phosphate buffer (pH 7.4), containing 0.3 M sucrose, 5 mM MOPS, 1 mM EGTA, and 0.1% bovine serum albumin. The homogenates were centrifuged at $15000 \times g$ for 10 min at 4°C , and the pellet was discarded. The supernatant was centrifuged at $15000 \times g$ in order to concentrate mitochondria in the pellet, which was finally dissolved in the same buffer (Latini et al., 2005).

Determination of the respiratory chain enzyme activities. Complex I activity was measured by the rate of NADH-dependent ferricyanide reduction as described in Cassina and Radl (1996). The activity of succinate-2,6-dichlorindophenol (DCIP)-oxidoreductase (complex II) was determined according to the method of Fischer et al. (1985) and cytochrome c oxidase (complex IV) activity according to Rustin et al. (1994). The methods described were slightly modified, as detailed in a previous report (Latini et al., 2005). The activities of the respiratory chain complexes were calculated as $\text{nmoL min}^{-1} \cdot \text{mg protein}^{-1}$.

Glutathione determination. Glutathione (GSH) levels were determined as described by Ellman (1959) with slight modifications. GSH were measured in tissue homogenates after protein precipitation with 1 volume of 10% trichloroacetic acid ($1000 \times g$ for 10 min). An aliquot of the protein-free supernatant was added to 800 mmol L^{-1} phosphate buffer, pH 7.4, and 500 mmol L^{-1} DTNB (5,5'-dithio-bis-2-nitrobenzoic acid). Color development resulting from the reaction between DTNB and thiols was read at 412 nm after 10 min. A standard curve of reduced GSH was used in order to calculate the GSH levels in the samples, and the results were expressed as $\text{nmoL GSH mg protein}^{-1}$.

Determination of thiobarbituric acid-reactive substances. Thiobarbituric acid-reactive substances (TBARS) were determined in tissue homogenates as described by Ohkawa et al. (1979), in which malondialdehyde (MDA), an end product of lipid peroxidation, reacts with thiobarbituric acid (TBA) to form a colored complex. In brief, samples were incubated at 100°C for 60 min in acid medium containing 0.45% sodium dodecyl sulfate and 0.6% TBA. After centrifugation, the reaction product was determined at 532 nm using 1,1,3,3-tetramethoxypropane as the standard, and the results were expressed as $\text{nmoL MDA mg protein}^{-1}$.

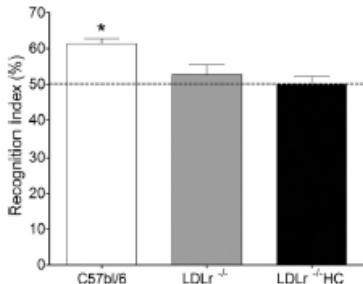


Fig 1. Effects of hypercholesterolemia on object location memory in wild-type C57bl/6 and $\text{LDLr}^{-/-}$ mice treated with standard or cholesterol-enriched diet. Each value represents the mean \pm SEM of six to eight animals in each group. * $P<0.05$ versus chance level (50% of displaced object investigation in test trial).

Glutathione reductase assay. Glutathione reductase (GR) activity was determined by the method described by Carlberg and Mannervik (1975), using oxidized glutathione (GSSG) as substrate. The enzyme activity was assessed in a solution containing 50 mM potassium phosphate buffer, pH 7.0, containing 1 mM EDTA, 0.2 mM NADPH and the supernatant containing 0.2–0.3 mg protein.mL⁻¹. The reaction was initiated by the addition of 1 mM GSSG, and the rate of GSSG reduction was indirectly determined through monitoring the NADPH disappearance at 340 nm. GR activity was expressed as nmol NADPH oxidized. min⁻¹.mg protein⁻¹, using an extinction coefficient $6.22 \times 10^3 \text{ M}^{-1} \cdot \text{cm}^{-1}$ for NADPH.

Glutathione peroxidase assay. Glutathione peroxidase (GPx) activity was measured according to the protocol developed by Wendel (1981) using tert-butyl hydroperoxide as substrate. The enzyme activity was determined by monitoring the NADPH disappearance at 340 nm in 50 mM potassium phosphate buffer, pH 7.0, containing 1 mM EDTA, 1 mM GSH, 0.2 U mL⁻¹ GR, 1 mM azide, 0.2 mM tert-butyl hydroperoxide, 0.2 mM NADPH and the supernatant containing 0.2–0.3 mg protein.mL⁻¹. GPx activity was expressed as nmol NADPH oxidized. min⁻¹.mg protein⁻¹, using an extinction coefficient of $6.22 \times 10^3 \text{ M}^{-1} \cdot \text{cm}^{-1}$ for NADPH.

Protein determination. The protein content of homogenate and mitochondrial preparation was determined by the method of Bradford (Bradford, 1976), using bovine serum albumin as the standard.

Statistical analysis

Data are expressed as means \pm SEM. Statistical analyses were carried out using one-way analysis of variance (ANOVA). Following significant ANOVAs, multiple post hoc comparisons were performed using the Duncan's test. Pearson's correlations (r , P) were calculated for associations between cholesterol levels and respiratory chain enzyme activities (the plot includes values from all the animals groups). The object location task was analyzed by one-sample t -tests to determine whether the location index was different from chance performance (50%). The accepted level of significance for the tests was $P\leq 0.05$. All tests were performed using the Statistica® software package (StatSoft Inc., Tulsa, OK, USA).

RESULTS

Hypercholesterolemia disrupts cognitive performance of mice in the object location task

Hypercholesterolemic $\text{LDLr}^{-/-}$ mice displayed spatial learning and memory impairments regardless of diet. As depicted in Fig. 1, the wild-type group showed a location index significantly higher than chance performance ($t=8.124$, $P=0.0001$), while $\text{LDLr}^{-/-}$ mice treated with standard diet ($t=-0.9498$, $P=0.3739$) or cholesterol-enriched diet ($t=-0.0312$, $P=0.9761$) were not able to identify the spatial alteration in the open field. The cholesterol-enriched diet did not trigger cognitive deficits in wild-type mice (data not show).

TC levels

Plasma cholesterol levels of wild-type and $\text{LDLr}^{-/-}$ mice are shown in Fig. 2. As expected, the cholesterol level of $\text{LDLr}^{-/-}$ mice treated with a standard diet were significantly higher when compared to wild-type group ($P\leq 0.05$). Furthermore, when the $\text{LDLr}^{-/-}$ mice were fed with a cholesterol-enriched diet, the cholesterol levels were around three-fold higher than $\text{LDLr}^{-/-}$ mice fed with standard diet ($P\leq 0.05$). By the other hand, the cholesterol levels were not modified in wild-type mice treated with a cholesterol-enriched diet (data not show).

Hypercholesterolemia inhibits the respiratory chain complexes I and II activities in cerebral cortex

The effect of hypercholesterolemia on cerebral cortex respiratory chain complexes is shown in Fig. 3. $\text{LDLr}^{-/-}$ mice treated with a cholesterol-enriched chow presented a significant inhibition of the activities of complexes I and II of the respiratory chain in cerebral cortex when compared to wild-type group ($P\leq 0.05$) and to $\text{LDLr}^{-/-}$ mice treated with standard chow ($P\leq 0.05$) (Fig. 3A). Additionally, the activities of complexes I and II in cerebral cortex were negatively

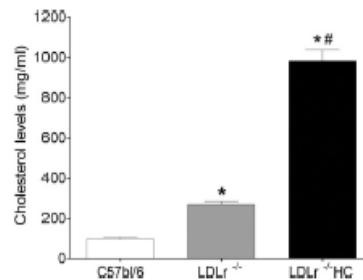


Fig 2. Plasma cholesterol levels of wild-type C57bl/6 and $\text{LDLr}^{-/-}$ mice treated with standard or cholesterol-enriched diet. Each value represents the mean \pm SEM of five to six animals in each group. * $P<0.05$ compared to wild-type C57bl/6 mice; # $P<0.05$ compared to $\text{LDLr}^{-/-}$ mice treated with standard diet (One-way ANOVA followed by the Duncan multiple range test).

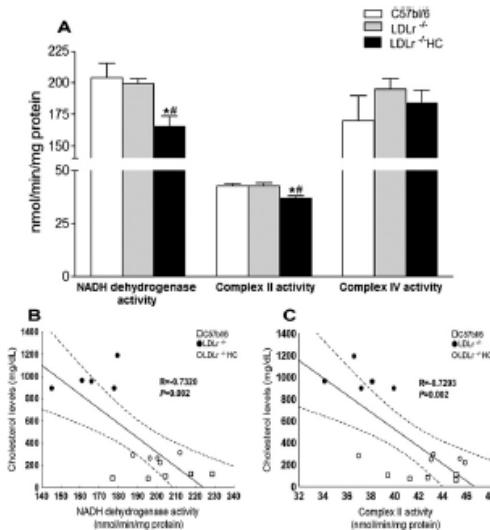


Fig 3. Effect of hypercholesterolemia on (A) respiratory chain complexes activities in cerebral cortex homogenates from wild-type C57bl/6 and LDL^{-/-} mice treated with standard or cholesterol-enriched diet. Each value represents the mean \pm SEM of five to six animals in each group. * $P<0.05$ compared to wild-type C57bl/6 mice; ** $P<0.05$ compared to LDL^{-/-} mice treated with standard diet (one-way ANOVA followed by the Duncan multiple range test). Significant correlation between (B) cholesterol levels and cerebral NADH dehydrogenase activity and (C) cholesterol levels and cerebral complex II activity.

correlated with blood cholesterol levels ($R=-0.7320$, $P=0.002$ and $R=-0.7293$, $P=0.002$, respectively; Fig. 3B and C, respectively), suggesting a potential link between hypercholesterolemia and impaired respiratory chain function in cerebral cortex of LDL^{-/-} mice.

Hypercholesterolemia modified oxidative stress-related parameters in cerebral cortex

The effect of hypercholesterolemia on cerebral oxidative stress parameters is shown in Fig. 4. Hypercholesterolemic mice presented a significantly impairment of the GSH-dependent antioxidant system in the cerebral cortex. GSH levels were significantly decreased when LDL^{-/-} mice were treated with a cholesterol-enriched chow when compared to wild-type group ($P=0.05$) and to LDL^{-/-} mice treated with standard chow ($P\leq0.05$) (Fig. 4A). The activities of the peroxide-removing-related enzymes, GPx and GR, are depicted in Fig. 4C, D, respectively. GPx activity was significantly reduced in hypercholesterolemic mice when compared to wild-type group ($P\leq0.05$) and to LDL^{-/-} mice treated with standard chow ($P\leq0.05$), while GR activity was significantly increased in both LDL^{-/-} mice treated with a standard or a cholesterol-enriched diet when compared to wild-type group ($P\leq0.05$). Moreover,

regardless of chow, LDL^{-/-} mice presented an increase in approximately 40% in cerebral lipid peroxidation ($P=0.26$), assessed through the TBARS measurement, when compared to wild-type group (Fig. 4B); nevertheless, this difference did not reach statistical significance.

DISCUSSION

Confirming previous studies (Mulder et al., 2004), the present findings demonstrate spatial learning and memory deficits of hypercholesterolemic LDL^{-/-} mice when compared to their C57bl/6 wild-type controls. More importantly, our results demonstrate for the first time that hypercholesterolemia impaired mitochondrial electron transfer chain activity in the cerebral cortex, which was negatively correlated with blood cholesterol levels. We also present evidence that cholesterol-enriched diet induces impairment in the GSH-dependent antioxidant defenses in the cerebral cortex.

Early literature data has demonstrated that a cholesterol-enriched diet impairs spatial learning and the ability to store long-term memory in rodents (Ulrich et al., 2010; Granholm et al., 2008). In the present study, LDL^{-/-} mice treated with either standard or cholesterol-enriched diet were not able to identify the spatial alteration in the open

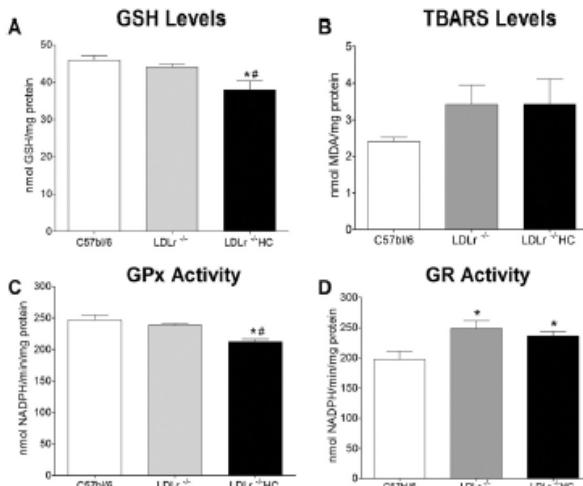


Fig. 4. Effect of hypercholesterolemia on oxidative stress parameters in cerebral cortex homogenates from wild-type C57BL/6 and LDLr^{-/-} mice treated with standard or cholesterol-enriched diet. Each value represents the mean ± SEM of five to six animals in each group. *P<0.05 compared to wild-type C57BL/6 mice; #P<0.05 compared to LDLr^{-/-} mice treated with standard diet (one-way ANOVA followed by the Duncan multiple range test). (A) GSH levels, (B) TBARS levels, (C) GPx activity, and (D) GR activity.

field. However, unlike LDLr^{-/-} mice, a 1-month treatment with cholesterol-enriched diet to wild-type mice is not able to modify cholesterol levels and to trigger cognitive deficits (data not show). In accordance with these observations, previous studies have demonstrated the absence of plasma cholesterol modification in wild-type mice receiving a high cholesterol diet (Ishibashi et al., 1994; Elder et al., 2008).

The object location/placement memory is proposed as a one-trial spatial memory model where the animals could learn without reinforcers (such as food) shaping the training process (Murai et al., 2007). The task has been widely used to evaluate the cognitive function of a variety of transgenic mice (Favre et al., 2011). This task, which took advantage of the preference normal mice show for an object that has moved from its previous position, revealed an impairment following radiofrequency lesions of the fornix and cytotoxic lesions of the cingulate cortex (Ennaceur et al., 1997). Also, it has long been known that the medial prefrontal cortex is important for spatial working memory (Brozoski et al., 1979), and, in this line of evidence, Nelson et al. (2011) demonstrated that lesions to the prelimbic cortex impaired performance in the object-location task. Nevertheless, this memory test, as others cognitive tasks, provides just a qualitative index (i.e. the animals either learned or did not learn), and, thus, probably for this reason we could not determine a further decrease in cognitive

function when LDLr^{-/-} mice were exposed to a high cholesterol diet.

The exact mechanisms responsible for the cognitive deficit induced by hypercholesterolemia in mice are still unknown. One possible explanation was elicited by Evola et al. (2010) using apoE (apolipoprotein E) knockout mice (a widely used model of hypercholesterolemia) and by Thirumangalakudi et al. (2008) using LDLr^{-/-} mice fed with high-fat/cholesterol diet. They argued that a noxious oxidative-inflammatory cycle in the vasculature could have deleterious consequences for brain function and cognition. In this line of evidence, Hafezi-Moghadam et al. (2007) showed that the genetic deletion of apoE in aged hypercholesterolemic mice induces a destabilizing effect on the cerebral microcirculation leading to blood-brain barrier leakage. Failure of the blood-brain barrier, with leakage of serum components into and through the walls of small cerebral vessels, can lead to neuronal and glial damage with persistent activation of microglia and astrocytes, which may be responsible for cognitive impairment (Rapp et al., 2008).

Neuroinflammatory processes also lead to increased production of ROS, which are detrimental to neurons (Witte et al., 2010). The mitochondria are victims of extensive oxidative damage and inflammation is likely to further contribute to mitochondrial dysfunction (Witte et al., 2010). Neurons are vulnerable to mitochondrial defects because

they require high levels of energy for the survival and specialized function (Chen and Chan, 2006). Of major interest, there is ample literature supporting a crucial role of mitochondrial dysfunction in AD, with altered energy metabolism and ROS production being the major correlates (Moreira et al., 2010; Lin and Beal, 2006).

On the other hand, changes in brain cholesterol levels seem not to be responsible for the observed cognitive impairments of $LDLr^{-/-}$ mice. There is evidence that the homeostasis of brain cholesterol is independent from the circulating cholesterol (Di Paolo and Kim, 2011). Moreover, previous studies demonstrated that even when submitted to high cholesterol, high-fat or high-fat/high-cholesterol diets, the $LDLr^{-/-}$ mice do not present any significant rise on brain cholesterol levels (Elder et al., 2007).

An interesting observation from our study was that a three-fold increase in serum cholesterol levels was enough to elicit a decrease in the behavioral performance without any significant impairment in mitochondrial function. However, a 10-fold increase (observed in $LDLr^{-/-}$ plus high-cholesterol diet) decreased mitochondrial complex function and induced GSH depletion, but did not further decrease performance in the recognition test. Based on these findings, one could suppose the occurrence of a "threshold phenomenon" for memory performance and mitochondrial function. However, based on the well-known relationship of cognitive impairment with either mitochondrial dysfunction or cholesterol dyshomeostasis, and considering the consistent literature, we realize that the occurrence of a threshold phenomenon for the memory performance and mitochondrial function can be ruled out. For example, in the study by Mulder et al. (2004), $LDLr^{-/-}$ mice displayed impaired hippocampal-dependent memory functions (demonstrated in several behavioral tests) when compared to $LDLr^{+/+}$ littermates. Mice lacking the LDL receptor displayed a decrease in the number of synaptophysin-immunoreactivity presynaptic boutons in the hippocampus CA1 region. These authors provided evidence for a role of the LDL receptor in the maintenance of normal synaptic plasticity in the hippocampus during processes of learning and memory. Since cholesterol is required for cell homeostasis, the reduced cellular uptake of cholesterol observed in $LDLr^{-/-}$ mice might underlie deleterious consequences in neuronal membrane function. Therefore, the lack of brain LDL receptor rather than the effect of the elevated serum cholesterol levels (two to three times higher than in the wild-type animals) on brain mitochondrial functions could be responsible for the cognitive deficits in $LDLr^{-/-}$ mice fed with normal diet. A similar event was already observed when evaluating the influence of hypercholesterolemia on the cognitive performance of $LDLr^{-/-}$ mice and its relationship with neuroinflammatory changes (Thirumangalakudi et al., 2008). The authors observed that memory deficits occurred regardless of diet, but pro-inflammatory markers were exacerbated in $LDLr^{-/-}$ treated with a hypercholesterolemic diet.

Furthermore, the relatively small period of exposure to this two-fold increase in cholesterol levels by $LDLr^{-/-}$ mice treated with standard chow (only 3 months) has to be

considered. This short-term exposure may explain the absence of brain mitochondrial dysfunction and oxidative stress. Thus, raising the serum cholesterol with the aid of a cholesterol-enriched diet would be expected to further accelerate the emergence of a noxious oxidative-inflammatory cycle in the vasculature. In our study, a cholesterol-enriched diet increased the total plasma cholesterol levels (around 10 times) in $LDLr^{-/-}$ mice, leading to a significant decrease in the mitochondrial complexes I and II activities in the cerebral cortex. Of high interest, we presented evidence that this inhibition of complexes I and II activities was negatively correlated with respective blood cholesterol levels. A previous study conducted in liver tissue of $LDLr^{-/-}$ mice by Oliveira et al. (2005) demonstrated that mitochondrial dysfunction and oxidative stress triggered by hypercholesterolemia are due to low content of pyridine nucleotides in the reduced state, which were presumably consumed by augmented lipogenesis. It is noteworthy that cortico-cerebral GPx activity was decreased in $LDLr^{-/-}$ mice fed a high cholesterol diet in our methodological approach, which uses saturated amounts of the pyridine nucleotide substrate (NADPH), thus suggesting that not only the levels of this coenzyme but also the enzyme catalytic activity was decreased as a result of hypercholesterolemia.

It is well documented that mitochondrial electron transport chain is the major source of cellular ROS and oxidative stress, with ROS generation occurring mainly at complex I (Sanz et al., 2006; Pamplona and Barja, 2007). Cellular oxidative stress and its downstream consequences are also a well-documented feature of AD pathophysiology (Sultana and Butterfield, 2011), and the CNS is especially prone to ROS-induced damage (Gemma et al., 2007). Here we demonstrated an imbalance in GPx/GR activity that might be related to hypercholesterolemia-induced oxidative stress, mainly by causing lipid peroxidation and GSH-dependent antioxidant system impairment. Several reports have shown a link between high dietary exposures to fat and/or cholesterol and oxidative stress in the brain of mice and rats (Crisby et al., 2004; Montilla et al., 2006) and our present data may support a role of mitochondrial dysfunction in the consequent oxidative imbalance induced by hypercholesterolemia.

GSH is an abundant intracellular antioxidant and scavenger of ROS (Dringen and Hirrlinger, 2003). Previous studies have shown that the GSH system may be activated as a response to oxidative stress in the brains of AD patients (Lovell et al., 1995; Aksenov et al., 2001). Additionally, hypercholesterolemia has been demonstrated to trigger oxidative stress by increasing the generation of ROS leading to a depletion of GSH levels, besides impairing the mitochondrial transport of GSH, resulting in mitochondrial GSH depletion and mitochondrial oxidative stress (Colell et al., 1997; Mari et al., 2008; Fernández et al., 2009). Corroborating these findings, Esposito et al. (2000) demonstrated that isolated liver mitochondria from GPx1-deficient mice have increased rates of H_2O_2 production, reduced mitochondrial respiratory control ratio, and decreased mitochondrial power output index.

CONCLUSIONS

To our knowledge, this is the first study to report a strong correlation between high levels of blood cholesterol and mitochondrial dysfunction in the cerebral cortex. This event was related with cognitive impairments, as well as lipid peroxidation, GSH depletion, and changes on peroxide-removing-related enzymes GPx and GR in the cerebral cortex of hypercholesterolemic $LDL^{-/-}$ mice. In conclusion, our findings provide new evidence about the positive correlation between elevated plasma cholesterol levels and cognitive impairments in hypercholesterolemic $LDL^{-/-}$ mice, which are mediated, at least in part, by mitochondrial dysfunctional and oxidative stress in cerebral cortex. In addition, they render this mouse model of hypercholesterolemia a useful approach to comprehend the molecular events mediating AD pathogenesis.

Acknowledgments—This work was supported by grants from the Brazilian Institutions: Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), and the Fundação de Apoio à Pesquisa do Estado de Santa Catarina (FAPESC). R.P., M.F., R.M.R.V., A.L., and A.B. are productivity fellows from the CNPq. Scholarships to J.O. (CAPES) and V.G. (FAPESC) are also acknowledged. M.A.H. is a PNPD/CAPES postdoctoral grantee. The authors are grateful to Ms. Renate Schlinke, an English professor, for checking the language of this manuscript.

REFERENCES

- Aksenov MY, Aksenova MV, Butterfield DA, Geddes JW, Markesberry WR (2001) Protein oxidation in the brain in Alzheimer's disease. *Neuroscience* 103:373–383.
- Asai FL, Duzzioni M, Takahashi RN (2009) Object location memory in mice: pharmacological validation and further evidence of hippocampal CA1 participation. *Behav Brain Res* 204:206–211.
- Ballinger SW (2005) Mitochondrial dysfunction in cardiovascular disease. *Free Radic Biol Med* 38:1278–1295.
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72:248–254.
- Brookes PS, Yoon Y, Robotham JL, Anders MW, Sheu SS (2004) Calcium, ATP, and ROS: a mitochondrial love-hate triangle. *Am J Physiol Cell Physiol* 287:C817–C833.
- Brozowski TJ, Brown RM, Roswold HE, Goldman PS (1979) Cognitive deficit caused by regional depletion of dopamine in prefrontal cortex of rhesus monkey. *Science* 205:929–932.
- Cadenas E, Davies KJ (2000) Mitochondrial free radical generation, oxidative stress, and aging. *Free Radic Biol Med* 29:222–230.
- Carlberg I, Mannervik B (1975) Purification and characterization of the flavoenzyme glutathione reductase from rat liver. *J Biol Chem* 250:5475–5480.
- Cassina A, Radi R (1996) Differential inhibitory action of nitric oxide and peroxynitrite on mitochondrial electron transport. *Arch Biochem Biophys* 328:309–316.
- Chen H, Chan DC (2006) Critical dependence of neurons on mitochondrial dynamics. *Curr Opin Cell Biol* 18:453–459.
- Collell A, García-Ruiz C, Morales A, Ballesta A, Oohkhtens M, Rodés J, Kaplowitz N, Fernández-Checa JC (1997) Transport of reduced glutathione in hepatic mitochondria and mitoplasts from ethanol-treated rats: effect of membrane physical properties and S-adenosyl-methionine. *Hepatology* 26:699–708.
- Cramer C, Haan MN, Galea S, Langa KM, Kalbfleisch JD (2008) Use of statins and incidence of dementia and cognitive impairment without dementia in a cohort study. *Neurology* 71:344–350.
- Crisby M, Rahman SM, Sylvén C, Winblad B, Schultzberg M (2004) Effects of high cholesterol diet on glialosis in apolipoprotein E knockout mice. Implications for Alzheimer's disease and stroke. *Neurosci Lett* 369:87–92.
- Di Paolo G, Kim TW (2011) Linking lipids to Alzheimer's disease: cholesterol and beyond. *Nat Rev Neurosci* 12:284–296.
- Dringen R, Hirfringer J (2003) Glutathione pathways in the brain. *Biochem* 384:505–516.
- Elder GA, Cho JY, English DF, Franciosi S, Schmeidler J, Sosa MA, Gasperini RD, Fisher EA, Mathews PM, Haroutunian V, Buxbaum JD (2007) Elevated plasma cholesterol does not affect brain Abeta in mice lacking the low-density lipoprotein receptor. *J Neurochem* 102:1220–1231.
- Elder GA, Raghauth A, Don N, Franciosi S, Schmeidler J, Haroutunian V, Buxbaum JD (2008) Increased locomotor activity in mice lacking the low-density lipoprotein receptor. *Behav Brain Res* 191:256–265.
- Ellman GL (1959) Tissue sulfhydryl groups. *Arch Biochem Biophys* 82:70–77.
- Ennaceur A, Neave N, Aggleton JP (1997) Spontaneous object recognition and object location memory in rats: the effects of lesions in the cingulate cortices, the medial prefrontal cortex, the cingulum bundle and the fornix. *Exp Brain Res* 113:509–519.
- Esposito LA, Kokoszka JE, Waymire KG, Cotrell B, MacGregor GR, Wallace DC (2000) Mitochondrial oxidative stress in mice lacking the glutathione peroxidase-1 gene. *Free Radic Biol Med* 28:754–766.
- Evans RM, Emaley CL, Gao S, Sahota A, Hall KS, Farlow MR, Hendrie H (2000) Serum cholesterol, APOE genotype, and the risk of Alzheimer's disease: a population-based study of African Americans. *Neurology* 54:240–242.
- Evola M, Hall A, Wall T, Young A, Grammas P (2010) Oxidative stress impairs learning and memory in apoE knockout mice. *Pharmacol Biochem Behav* 96:181–186.
- Fairen E, Gault VA, Thorens B, Hölscher C (2011) Glucose-dependent insulinotropic polypeptide receptor knockout mice are impaired in learning, synaptic plasticity, and neurogenesis. *J Neurophysiol* 105:1574–1580.
- Fernández A, Llucena L, Fernández-Checa JC, Collell A (2009) Mitochondrial cholesterol loading exacerbates amyloid beta peptide-induced inflammation and neurotoxicity. *J Neurosci* 29:6394–6405.
- Fischer JC, Ruitenberg W, Berden JA, Trijbels JM, Veerkamp JH, Stadhouders AM, Senger RC, Janssen AJ (1985) Differential investigation of the capacity of succinate oxidation in human skeletal muscle. *Clin Chim Acta* 153:23–36.
- Fonantesi F, Diaz B, Barrientos A (2009) Evaluation of the mitochondrial respiratory chain and oxidative phosphorylation system using yeast models of OXPHOS deficiencies. In: *Curr Protoc Hum Genet*, vol. 63 (Boyle A, ed), Unit 19.5, pp 1–20.
- Granholm AC, Bimonte-Nelson HA, Moore AB, Nelson ME, Freeman LR, Sambamurti K (2008) Effect of a saturated fat and high cholesterol diet on memory and hippocampal morphology in the middle-aged rat. *J Alzheimers Dis* 14:133–145.
- Hafezi-Moghadam A, Thomas KL, Wagner DD (2007) ApoE deficiency leads to a progressive age-dependent blood-brain barrier leakage. *Am J Physiol Cell Physiol* 292:C1256–C1262.
- Hauptmann S, Scherping I, Dröse S, Brandl U, Schulz KL, Jendrach M, Leuner K, Eckert A, Müller WE (2009) Mitochondrial dysfunction: an early event in Alzheimer pathology accumulates with age in AD transgenic mice. *Neurobiol Aging* 30:1574–1586.
- Hermann W, Knapp JP (2002) Hyperhomocysteinemia: a new risk factor for degenerative diseases. *Clin Lab* 48:471–481.
- Hort MA, Stratioto MR, Netto PM, da Rocha JB, de Bem AF, Ribeiro-Do-Valle RM (2011) Diphenyl diselenide effectively reduces atherosclerotic lesion in $LDL^{-/-}$ mice by attenuation of oxidative stress and inflammation. *J Cardiovasc Pharmacol* 58:91–101.

- Ishibashi S, Brown MS, Goldstein JL, Gerard RD, Hammer RE, Herz J (1993) Hypercholesterolemia in low density lipoprotein receptor knockout mice and its reversal by adenovirus-mediated gene delivery. *J Clin Invest* 92:883–893.
- Ishibashi S, Goldstein JL, Brown MS, Herz J, Burns DK (1994) Massive xanthomatosis and atherosclerosis in cholesterol-fed low density lipoprotein receptor-negative mice. *J Clin Invest* 93:1885–1893.
- Kowalewski AJ, Vercesi AE (1999) Mitochondrial damage induced by conditions of oxidative stress. *Free Radic Biol Med* 26:463–471.
- Latin A, da Silva CG, Ferreira GC, Schuck PF, Scussiato K, Sarkis JJ, Dutra Filho CS, Wyse AT, Wannmacher CM, Wajner M (2005) Mitochondrial energy metabolism is markedly impaired by D-2-hydroxyglutaric acid in rat tissues. *Mol Genet Metab* 86:188–199.
- Leonard JV, Schapira AH (2000) Mitochondrial respiratory chain disorders I: mitochondrial DNA defects. *Lancet* 355:299–304.
- Levin-Alerhand JA, Lominack CE, Smith JD (2002) Increased amyloid-levels in APPSWE transgenic mice treated chronically with a physiological high-fat/high-cholesterol diet. *J Nutr Health Aging* 6:315–319.
- Lin MT, Beal MF (2006) Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 443:787–795.
- Lovell MA, Ehrmann WD, Butler SM, Marksberry WR (1995) Elevated thiobarbituric acid-reactive substances and antioxidant enzyme activity in the brain in Alzheimer's disease. *Neurology* 45:1594–1601.
- Medamanchi NR, Runge MS (2007) Mitochondrial dysfunction in atherosclerosis. *Circ Res* 100:460–473.
- Mari M, Colell A, Morales A, Caballero F, Molera A, Fernández A, Terresos O, Basañez G, Antonsson B, García-Ruiz C, Fernández-Checa JC (2008) Mechanism of mitochondrial glutathione-dependent hepatoocular susceptibility to TNF despite NF-κB activation. *Gastroenterology* 134:1507–1520.
- Montilla P, Espejo I, Muñoz MC, Bujalance I, Muñoz-Castañeda JR, Tuniez J (2006) Protective effect of red wine on oxidative stress and antioxidant enzyme activities in the brain and kidney induced by feeding high cholesterol in rats. *Clin Nutr* 25:146–153.
- Moreira PI, Carvalho C, Zhu X, Smith MA, Perry G (2010) Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. *Biochim Biophys Acta* 1802:2–10.
- Mulder M, Jansen PJ, Janssen BJ, van de Berg WD, van der Boom H, Hawekes LM, de Koei RE, Ramaekers FC, Blokland A (2004) Low-density lipoprotein receptor-knockout mice display impaired spatial memory associated with a decreased synaptic density in the hippocampus. *Neurobiol Dis* 16:212–219.
- Murai T, Okuda S, Tanaka T, Ohta H (2007) Characteristics of object location memory in mice: behavioral and pharmacological studies. *Physiol Behav* 90:116–124.
- Nelson AJD, Cooper MT, Thur KE, Mansden CA, Cassaday HJ (2011) The effect of catecholaminergic depletion within the prelimbic and infralimbic medial prefrontal cortex on recognition memory for recency, location, and objects. *Behav Neurosci* 125:396–403.
- Okawa H, Ohishi N, Yagi K (1979) Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 95:351–358.
- Oliveira HC, Cossio RG, Alberici LC, Maciel EN, Salerno AG, Donghelli GG, Velho JA, de Faria EC, Vercesi AE (2005) Oxidative stress in atherosclerosis-prone mouse is due to low antioxidant capacity of mitochondria. *FASEB J* 19:278–280.
- Palm BA, Velho JA, Castilho RF, Oliveira HC, Vercesi AE (2008) Oxidative stress in hypercholesterolemic LDL (low-density lipoprotein) receptor knockout mice is associated with low content of mitochondrial NADP-linked substrates and is partially reversed by citrate replacement. *Free Radic Biol Med* 44:444–451.
- Pampushny R, Barja G (2007) Highly resistant macromolecular components and low rate of generation of endogenous damage: two key traits of longevity. *Ageing Res Rev* 6:189–210.
- Rapp JH, Pan XM, Neumann M, Hong M, Hollenbeck K, Liu J (2008) Microemboli composed of cholesterol crystals disrupt the blood-brain barrier and reduce cognition. *Stroke* 39:2354–2361.
- Reñido LM, Malester B, LaFrancesco J, Bryant-Thomas T, Wang R, Tint GS, Samambunti K, Duff K, Pappolla MA (2000) Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. *Neurobiol Dis* 7:321–331.
- Rojo L, Sjöberg MK, Hernández P, Zambrano C, Maccioni RB (2006) Roles of cholesterol and lipids in the etiopathogenesis of Alzheimer's disease. *J Biomed Biotechnol* 73976:2006.
- Rustin P, Chreden D, Bourgeron T, Gérard B, Röög A, Saudubray JM, Munich A (1994) Biochemical and molecular investigations in respiratory chain deficiencies. *Clin Chim Acta* 228:35–51.
- Sanz A, Pamplona R, Barja G (2006) Is the mitochondrial free radical theory of aging intact? *Antioxid Redox Signal* 8:582–599.
- Shanbhag AR, Ding J, Criqui MH, Saad MF, Liu K, Polak JF, Folsom AR, Tsai MY, Burke GL, Szkoła M (2006) Smoking, diabetes, and blood cholesterol differ in their association with subclinical atherosclerosis: the Multiethnic Study of Atherosclerosis (MESA). *Atherosclerosis* 186:441–447.
- Sparks DL, Kuo YM, Roher A, Martin T, Lukas RJ (2000) Alterations of Alzheimer's disease in the cholesterol-fed rabbit, including vascular inflammation. Preliminary observations. *Ann N Y Acad Sci* 903:335–344.
- Stary HC (1989) Evolution and progression of atherosclerotic lesions in coronary arteries of children and young adults. *Arteriosclerosis* 8:119–132.
- Sultana R, Butterfield DA (2011) Identification of the oxidative stress proteome in the brain. *Free Radic Biol Med* 50:487–494.
- Thirumangalakudi L, Prakasam A, Zhang R, Bimonte Nelson H, Samambunti K, Kindy MS, Bhat NR (2008) High cholesterol-induced neuroinflammation and amyloid precursor protein processing correlate with loss of working memory in mice. *J Neurochem* 100: 475–485.
- Ulrich C, Pirchi M, Humpel C (2010) Hypercholesterolemia in rats impairs the cholinergic system and leads to memory deficits. *Mol Cell Neurosci* 45:408–417.
- Valko M, Leiblitz D, Moncol J, Cronin MT, Mazur M, Telszer J (2007) Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* 39:44–84.
- Vercesi AE, Castilho RF, Kowalewski AJ, Oliveira HC (2007) Mitochondrial energy metabolism and redox state in dyslipidemias. *IUBMB Life* 59:263–268.
- Wendel A (1981) Glutathione peroxidase. *Methods Enzymol* 77:325–333.
- Witte ME, Geurts JJ, de Vries HE, van der Valk P, van Horssen J (2010) Mitochondrial dysfunction: a potential link between neuroinflammation and neurodegeneration? *Mitochondrion* 10:411–418.
- Yaffe K, Barrett-Connor E, Lin F, Grady D (2002) Serum lipoprotein levels, statin use, and cognitive function in older women. *Arch Neurol* 59:378–384.
- Zadekaar S, Kleemann R, Verschuren L, de Vries-Van der Weij J, van der Hoorn J, Princen HM, Kooistra T (2007) Mouse models for atherosclerosis and pharmaceutical modifiers. *Arterioscler Thromb Vasc Biol* 27:1706–1721.

(Accepted 5 September 2011)
(Available online 16 September 2011)

4.2 MANUSCRITO 1

Diphenyl diselenide, a glutathione peroxidase mimetic, prevents cortico-cerebral oxidative stress induced by hypercholesterolemia in the LDL receptor knockout mice

Manuscrito em fase final de preparação.

Diphenyl diselenide, a glutathione peroxidase mimetic, prevents cortico-cerebral oxidative stress induced by hypercholesterolemia in the LDL receptor knockout mice.

Jade de Oliveira^a, Eduardo Luiz Gasnhar Moreirab, Mariana Appel Horta, João Batista Teixeira da Rochac, Rosa Maria Ribeiro-do-Valle^b, Marcelo Farina^a, Andreza Fabro de Bema,*.

^aDepartamento de Bioquímica, Universidade Federal de Santa Catarina, 88040-900, Florianópolis, SC, Brazil.

^bDepartamento de Farmacologia, Universidade Federal de Santa Catarina, 88049-900, Florianópolis, SC, Brazil.

^cDepartamento de Química, Universidade Federal de Santa Maria, 97105900, Santa Maria, RS, Brazil.

*Corresponding author:

Andreza Fabro de Bem, PhD

Departamento de Bioquímica, Universidade Federal de Santa Catarina, 88040-900, Florianópolis, SC, Brazil.

Phone: +55 48 3721 5565; Fax: +55 48 3337 9672

E-mail: andrezadebem@ccb.ufsc.br

Abstract

Considerable evidence supports an association between hypercholesterolemia and brain oxidative stress. The interaction between antioxidant function and oxidative stress has emerged as a common causative factor in the pathology of several brain disorders. In the low density lipoprotein receptor knockout ($\text{LDLr}^{-/-}$) mice, cholesterol-enriched diet leads to memory impairment, as well, brain mitochondrial dysfunction and oxidative stress. In the present study, we evaluated the potential neuroprotective effect of diphenyl diselenide ($(\text{PhSe})_2$), a glutathione peroxidase (GPx) mimetic, against hypercholesterolemia-induced cortico-cerebral oxidative stress in the $\text{LDLr}^{-/-}$ mice submitted to a model of hypercholesterolemia. Our results showed that oral treatment with $(\text{PhSe})_2$ prevented the cortico-cerebral oxidative stress in the 3 months-old $\text{LDLr}^{-/-}$ mice fed with cholesterol-enriched diet, significantly increased the GSH content, and decreased the lipid peroxidation. These antioxidant actions seem to be in part related to its thiol peroxidase-like activity. Based on our findings, in addition to its protective effects in cardiovascular diseases associated with hypercholesterolemia, $(\text{PhSe})_2$ can be pointed as a promising neuroprotective molecule against hypercholesterolemia-induced effects deleterious on brain.

Keywords:

Hypercholesterolemia, cerebral oxidative stress, diphenyl diselenide, low density lipoprotein receptor knockout mice.

Introduction

Cholesterol is an important lipid constituent of all cellular membranes and myelin (Simons and Ikonen, 2000). Cholesterol also functions as the substrate for the synthesis of bile acids in the liver, and as the precursor of steroid hormones in endocrine tissue (Jackson et al., 1997). Meanwhile, despite its role in essential biochemical processes and support of membrane structure, elevated plasma cholesterol levels are associated with negative health outcomes especially its association with vascular disease (Lusis, 2000, Grundy et al., 2004, Lewington et al., 2007). Furthermore, alterations in cholesterol metabolism have been implicated with deleterious consequences of brain function (Sparks et al., 2000, Wolozin, 2004b, a, Ullrich et al., 2010). For instance, epidemiological and neurochemical investigations are providing increasing evidences that altered cholesterol metabolism contributes to the development of Alzheimer's disease (Pappolla et al., 2002, Di Paolo and Kim, 2011).

Indeed, there is compelling findings indicating a causal link between high dietary cholesterol intake and brain oxidative stress (Crisby et al., 2004, Montilla et al., 2006, Amin et al., 2011). Previous reports demonstrated that hypercholesterolemia triggers a neuroinflammatory response and brain oxidative stress, ultimately resulting in cognitive impairments and neurodegeneration (Sparks and Schreurs, 2003, Thirumangalakudi et al., 2008, Lu et al., 2009, Lu et al., 2010) For instance, Lu et al. (2010) observed that hypercholesterolemia increases reactive oxygen species (ROS) and reduces superoxide dismutase (SOD) activity in the brain of aged C57Bl/6 mice, and these alterations were attenuated by antioxidant compounds. In this context, we recently observed an imbalance in the glutathione (GSH)-dependent antioxidant defenses and an enhanced lipid peroxidation in the cerebral cortex of hypercholesterolemic low density lipoprotein receptor knockout ($LDLR^{-/-}$) mice (de Oliveira et al., 2011).

However, even though several epidemiological studies demonstrated a connection between cholesterol-lowering agents (e.g. statins) and a significant reduction in the prevalence of neurologic disorders in general population (Jick et al., 2000, Hajjar et al., 2002, Rodriguez et al., 2002, Dufouil et al., 2005, Wolozin et al., 2007), preliminary clinical studies reported modest success (Simons et al., 2002, Sparks et al., 2005, Sparks et al., 2006). In this scenario, there is compelling evidences that organoselenium compounds are promising pharmacological agents (Nogueira et al., 2004, Rosa et al., 2007). Of

particular significance, organoselenium compounds can mimicry endogenous antioxidant enzymes, such as glutathione peroxidase (GPx).

Diphenyl diselenide ($(\text{PhSe})_2$) is a simple diorganoil selenium compound that displays GPx-like activity, and presented neuroprotective and antioxidant activities *in vitro* (Rossato et al., 2002, Meotti et al., 2004, Puntel et al., 2007, Posser et al., 2008) and *in vivo* (Barbosa et al., 2006, Barbosa et al., 2008). Of particular relevance, our research group reported that oral treatment with lower doses of $(\text{PhSe})_2$ potently reduced the formation of atherosclerotic lesion in the $\text{LDLr}^{-/-}$ mice (Hort et al., 2011), as well as, led to beneficial effects on serum total cholesterol and on several parameters related to oxidative stress in hypercholesterolemic rabbits (de Bem et al., 2009). Moreover, we demonstrated that $(\text{PhSe})_2$ inhibited human LDL oxidation and that this phenomenon was related to its GPx-like activity (de Bem et al., 2008). Based on the above mentioned effects of $(\text{PhSe})_2$, we hypothesized that this compound may present neuroprotective effects against hypercholesterolemia- induces cortico-cerebral oxidative stress. To test this hypothesis, in the present study we examined the effect of $(\text{PhSe})_2$ on cortico-cerebral oxidative stress in the 3-months-old $\text{LDLr}^{-/-}$ mice fed daily with cholesterol-enriched diet during 30 days.

Material and methods

Chemicals

Diphenyl diselenide ($(\text{PhSe})_2$) was synthesized according to the literature methods (Paulmier, 1986). Analysis of the ^1H NMR and ^{13}C NMR spectra showed that the compound obtained presented analytical and spectroscopic data in full agreement with its assigned structure. The chemical purity of $(\text{PhSe})_2$ (99.9%) was determined by GC/HPLC. All other chemicals were of analytical grade and obtained from standard commercial suppliers.

Animals

C57Bl/6 low density lipoprotein receptor knockout ($\text{LDLr}^{-/-}$) mice were obtained by homologous recombination, as previously described by Ishibashi et al. (1993). The progenitors were purchased from Jackson Laboratory (Bar Harbor, Maine, USA). Animals were maintained at controlled room temperature ($21 \pm 2^\circ\text{C}$) under a 12 h light/12 h dark cycle with free access to food and water. All experiments were

undertaken in accordance with the guidelines on animal care of the local Ethics Committee on the Use of Animals (CEUA/UFSC), which follows the NIH publication “Principles of Laboratory Animal Care”.

Experimental protocols

In order to evaluate the neuroprotective effects of (PhSe)₂ against hypercholesterolemia-induces cortico-cerebral oxidative stress, male 3-months-old LDLr^{-/-} mice weighing 24–26 g were randomly divided into two experimental groups, that were fed with a standard or a cholesterol-enriched diet (20% fat, 1.25% cholesterol, 0.5% cholic acid) (de Oliveira et al., 2011). Half of the animals from each group were concomitantly treated daily with (PhSe)₂ (1 mg/kg; o.g.), and the other half with vehicle (canola oil), totalizing four experimental groups. Following 30 days of treatments, the animals were food-deprived overnight and the blood was collected by cardiac puncture in heparinized tubes for posterior determination of plasma cholesterol levels. After that, the mice were sacrificed by decapitation, and the cerebral cortex was removed for the biochemical analyses.

Plasma cholesterol levels

Whole blood was centrifuged at 3,000 x g, at room temperature for 10 min and the obtained plasma was used to measure total cholesterol and non-HDLcholesterol levels. Total and HDL plasma cholesterol were measured in plasma by an enzymatic method (Labtest Diagnostica®, Lagoa Santa-MG, Brazil). The concentration of non-HDL-cholesterol was calculated using the equation: (LDL + VLDL + IDL) = TC – HDL.

Cerebral cortex homogenates preparation

Cerebral cortex was removed and homogenized (1:10 w/v) in HEPES buffer (20 mM, pH 7.0). The tissue homogenates were centrifuged at 16,000 x g, at 4°C for 20 min and the supernatants obtained were used for the determination of glutathione peroxidase (GPx) and glutathione reductase (GR) activities and for the quantification of the levels of reduced glutathione (GSH) and thiobarbituric acid reactive substances (TBARS).

Reduced glutathione levels

Glutathione (GSH) levels were determined as described by Ellman (1959) with slight modifications. GSH was measured in cerebral cortex homogenates after precipitation with 1 vol. of 10% trichloroacetic acid and centrifuged at 1,000 x g at 4 °C for 10 min. An aliquot of the protein-free supernatant was added to 800 mmolM phosphate buffer, pH 7.4, and 500 mmolM1 DTNB (5,5'-dithio-bis-2-nitrobenzoic acid). Color development resulting from the reaction between DTNB and thiols was read at 412 nm after 10 min. A standard curve of reduced GSH was used in order to calculate the GSH levels in the samples, and the results were expressed as nmol GSH.mg protein⁻¹.

Thiobarbituric acid-reactive substances levels

Thiobarbituric acid-reactive substances (TBARS) were determined in tissue homogenates as described by Ohkawa et al. (1979), in which malondialdehyde (MDA), an end product of lipid peroxidation, reacts with thiobarbituric acid (TBA) to form a colored complex. In brief, samples were incubated at 100 °C for 60 min in acid medium containing 0.45% sodium dodecyl sulfate and 0.6% TBA. After centrifugation, the reaction product was determined at 532 nm using 1,1,3,3-tetramethoxypropane as the standard, and the results were expressed as nmol MDA .mg protein⁻¹.

Glutathione reductase activity

Glutathione reductase (GR) activity was determined according to previously described by Carlberg and Mannervik (1985). The enzyme activity was assessed in a solution containing 50mM potassium phosphate buffer, pH 7.0, containing 1 mM EDTA, 0.2 mM NADPH and the supernatant containing 0.2–0.3 mg protein.ml⁻¹. The reaction was initiated by the addition of 1 mM oxidized glutathione and a change in absorbance was measured at 340 nm. GR activity was expressed as nmol NADPH oxidized.min⁻¹.mg protein⁻¹, using an extinction coefficient 6.22 x 103 M-1.cm⁻¹ for NADPH.

Glutathione peroxidase activity

Glutathione peroxidase (GPx) activity was measured according to the protocols developed by Wendel (1981) using tert-

butylhydroperoxide as substrate. The enzyme activity was determined by monitoring the NADPH disappearance at 340 nm in 50 mM potassium phosphate buffer, pH 7.0, containing 1mM EDTA, 1 mM glutathione, 0.2 U ml⁻¹ glutathione reductase, 1 mM azide, 0.2 mM tert-butyl-hydroperoxide, 0.2 mM NADPH and the supernatant containing 0.2–0.3 mg protein.ml⁻¹. GPx activity was expressed as nmol NADPH oxidized.min⁻¹.mg protein⁻¹, using an extinction coefficient of 6.22 x 103 M⁻¹ cm⁻¹ for NADPH.

Protein determination

Sample protein content was determined by the method of Lowry et al. (1951) using bovine serum albumin as the standard.

Statistical analysis

All data are expressed as mean ± S.E.M. The statistical evaluation was performed using two-way analysis of variance (ANOVA). Following significant ANOVAs, multiple post-hoc comparisons were performed using the Duncan's test. The accepted level of significance for the tests was $P \leq 0.05$. All tests were performed using the Statistica software package (StatSoft Inc., Tulsa, OK, USA).

Results

Plasma cholesterol concentration

In previous studies we demonstrated that when the 3 months-old LDLr^{-/-} mice are submitted to a standard diet they presents a two- to four-fold increase in plasma cholesterol concentration when compared to the wild type mice. Moreover, LDLr^{-/-} mice submitted to a cholesterol-enriched diet during 30 days presents around three-fold increase in total cholesterol levels compared to LDLr^{-/-} mice fed with a standard diet (de Oliveira et al., 2011; Hort et al., 2011). Therefore, we investigated the effects of (PhSe)₂ treatment in the plasma cholesterol concentration of the hypercholesterolemic LDLr^{-/-} mice.

Two-way ANOVA indicated only a significant effect for the diet factor on total plasma cholesterol levels [$F(1, 16) = 212.68, P \leq 0.05$],

and on non-HDL-cholesterol levels [$F(1, 16) = 140.32, P \leq 0.05$]. Subsequent post-hoc comparisons indicated that $\text{LDLr}^{-/-}$ mice submitted to the cholesterol-enriched diet presented a significantly increase in the total cholesterol and non-HDL cholesterol levels when compared to the $\text{LDLr}^{-/-}$ mice submitted to the standard diet. Meanwhile, as observed in Table 1, $(\text{PhSe})_2$ treatment did not alter total plasma cholesterol levels and non-HDL cholesterol levels in the $\text{LDLr}^{-/-}$ mice regardless of diet.

Effects of $(\text{PhSe})_2$ against hypercholesterolemia-induced cortico-cerebral oxidative stress

In a previous report, we demonstrated that 3 months-old $\text{LDLr}^{-/-}$ mice submitted to a cholesterol-enriched diet during 30 days presented a significant inhibition of the mitochondrial complexes I and II activities, as well as, an increased lipid peroxidation and an impairment in the GSH-dependent antioxidant system in the cerebral cortex, characterized by a diminishing in the reduced glutathione levels and a decrease in the GPx activity (de Oliveira et al., 2011). Herein, we demonstrated that the treatment with a GPx mimic compound, $(\text{PhSe})_2$, protects against hypercholesterolemia- induced cortico-cerebral oxidative stress.

Two-way ANOVA indicated a significant effect for the treatment factor [$F(1, 27) = 73.31, P \leq 0.05$], and for the interaction between diet and treatment factors [$F(1, 27) = 34.77, P \leq 0.05$] on GSH levels. Subsequent post-hoc comparisons revealed that the cholesterol-enriched diet significantly reduced the GSH levels in the cerebral cortex of $\text{LDLr}^{-/-}$ mice ($P \leq 0.05$, Figure 1A), and $(\text{PhSe})_2$ treatment prevent this reduction in the $\text{LDLr}^{-/-}$ mice fed with cholesterol enriched-diet ($P \leq 0.05$, Figure 1A). Furthermore, indicated a significant effect for the treatment factor [$F(1, 15) = 122.68, P \leq 0.05$], and for the interaction between diet and treatment factors [$F(1, 15) = 4.84, P \leq 0.05$] on GPx activity. Subsequent post-hoc comparisons revealed that the cholesterol-enriched diet significantly reduced the GPx activity in the cerebral cortex of $\text{LDLr}^{-/-}$ mice ($P \leq 0.05$, Figure 1C), while the $(\text{PhSe})_2$ treatment increased the GPx activity in the $\text{LDLr}^{-/-}$ mice fed with a standard diet but not in the $\text{LDLr}^{-/-}$ mice fed cholesterol-enriched diet. Finally, Two-way ANOVA indicated a significant effect for the diet factor [$F(1, 16) = 10.50, P \leq 0.05$] and for the interaction between diet and treatment factors [$F(1, 16) = 7.42, P \leq 0.05$] on GR activity in cerebral cortex of $\text{LDLr}^{-/-}$ mice. Subsequent post-hoc comparisons revealed that treatment with $(\text{PhSe})_2$ decreased the GR activity in the $\text{LDLr}^{-/-}$ mice fed cholesterol-enriched diet ($P \leq 0.05$, Figure 1C). In addition, two-way ANOVA indicated a significant effect for the

treatment factor [$F(1, 25) = 12.496, P \leq 0.05$] on lipid peroxidation in cerebral cortex of $\text{LDLr}^{-/-}$ mice. Subsequent post-hoc comparisons revealed a decreased lipid peroxidation in the cerebral cortex of $\text{LDLr}^{-/-}$ mice fed cholesterol-enriched diet following treatment with $(\text{PhSe})_2$ ($P \leq 0.05$, Figure 1B).

Discussion

There are considerable evidences that hypercholesterolemia is associated with brain oxidative stress in humans and in rodents models (Stokes et al., 2002, Lu et al., 2009, Lu et al., 2010, Ramirez et al., 2011). Furthermore, an imbalance between antioxidant functions and oxidative stress is implicated in diverse brain pathologies (Harish et al., 2011). In this context, several studies have reported the beneficial effects of organoselenium compounds against pathological conditions associated with oxidative stress (e.g., inflammation, diabetes, neurotoxicity and hepatotoxicity) (Wilson et al., 1989, Nogueira et al., 2004, Nogueira and Rocha, 2011). Our present results demonstrated that $(\text{PhSe})_2$ effectively prevented the cortico-cerebral oxidative stress induced by cholesterol-enriched diet in the 3-months-old $\text{LDLr}^{-/-}$ mice. Here, the antioxidant action of $(\text{PhSe})_2$ was linked to a reduction in lipid peroxidation and increased in GSH content in cortico-cerebral homogenates of $\text{LDLr}^{-/-}$ mice submitted to a hipercholesterolemic diet.

Hypercholesterolemia has been demonstrated to trigger oxidative stress by increasing reactive oxygen species (ROS) generation and decreasing oxidative defenses (Colell et al., 1997, Mari et al., 2008, Fernandez et al., 2009). One potential mechanism by which hypercholesterolemia may induce oxidative stress is related to a disruption of mitochondrial function (McCommis et al., 2011). Previous studies showed that mitochondria obtained from various tissues of the hypercholesterolemic $\text{LDLr}^{-/-}$ mice generate higher amounts of ROS when compared to those of wild-type mates (Oliveira et al., 2005, Paim et al., 2008, de Oliveira et al., 2011). The authors suggested that higher rates of lipogenesis in the $\text{LDLr}^{-/-}$ mice deplete the mitochondrial reducing equivalents from NADPH leading to a state of oxidative stress.

However, since the homeostasis of brain cholesterol is independent from the circulating cholesterol (Di Paolo and Kim, 2011), the exact mechanisms responsible for the cortico-cerebral oxidative stress induced by hypercholesterolemia in the $\text{LDLr}^{-/-}$ mice remains unclear. One possible explanation is that the increased metabolic flux to the brain during hypercholesterolemia can orchestrate the blood-brain

barrier (BBB) disruption, recruitment of inflammatory immune cells from peripheral blood and microglial cells activation leading to neuroinflammation (Kalayci et al., 2009). In fact, inflammation of brain capillary endothelial cells may play a potent role in the neurodegenerative diseases, and it is well known that endothelial cells strongly respond to inflammatory stimuli (Moser et al., 2004) especially involving production of ROS (Iadecola, 2004). In this line of evidence, (Thirumangalakudi et al., 2008) using $\text{LDLr}^{-/-}$ mice fed with high-fat/cholesterol diet demonstrated that a noxious oxidative-inflammatory cycle in the vasculature could have deleterious consequences for brain function. Indeed, recently we proposed an oxidative stress condition in the cortex cerebral of hypercholesterolemic $\text{LDLr}^{-/-}$ mice after 30 days feeding with high cholesterol diet, which was characterized by impairment in the GSH metabolism and lipid peroxidation (de Oliveira et al., 2011).

In mammalian cells, glutathione (GSH) and the GPxs constitute one of the most important antioxidant defense systems (Raes et al., 1987). GSH is an abundant intracellular antioxidant and scavenger of ROS, providing the neuronal cell with important protection against oxidative damage (Brigelius-Flohe, 1999, Lima et al., 2006). Herein, we showed that $(\text{PhSe})_2$ significantly increased the GSH content in the cerebral cortex of 3-months-old $\text{LDLr}^{-/-}$ mice fed with cholesterol-enriched diet, which possibly contributed to the enhanced detoxification of lipid peroxidation products as demonstrated by decreased in the MDA levels. In this way, there are two major pathways involved in regulating of cellular GSH contents: one is GR, which catalyzes the reduction of GSSG to GSH via consumption of NADPH (Carlberg and Mannervik, 1985), and the other is γ -glutamyl cysteine synthase (γ GCS), which is the rate-limiting enzyme in GSH synthesis (Griffith and Mulcahy, 1999). In this study, the treatment with $(\text{PhSe})_2$ paradoxically decreased the GR activity, which may occurred due to increase in the GSH levels by increased biosynthesis, not requiring regeneration. Furthermore, the $(\text{PhSe})_2$ treatment increased the GPx activity in the $\text{LDLr}^{-/-}$ mice fed standard diet. In this scenario, we can speculate that $(\text{PhSe})_2$ might modulate intracellular signaling pathways related to antioxidant response. In line with this, recent studies demonstrated that the organoselenium compound ebselen activates the Nrf-2, a transcription factor orchestrating antioxidant and cytoprotective responses on oxidative stress that ultimately increases the expression of genes encoding enzymes of the glutathione redox system, such as γ GCS and GPx (Tamasi et al., 2004, Kim et al., 2009).

GPx is an antioxidant defense enzyme that can directly detoxify H₂O₂ and lipid hydroperoxides at the expense of GSH (Brigelius-Flohe, 1999), and accumulating evidence suggested that GPx functions as the primary protection against acute oxidative stress, particularly in neuropathological situations (De Haan et al., 2003, Ran et al., 2006). In line of this, another possible mechanism associated with the observed (PhSe)₂ neuroprotection in the hypercholesterolemic LDLr^{-/-} mice, is the its capacity of mimic endogenous antioxidant enzymes, such as GPx (Nogueira et al., 2004, Borges et al., 2006, de Freitas and Rocha, 2011). In this regard, Posser et al. (2008) demonstrated that (PhSe)₂ exerts protective effects against H₂O₂-induced oxidative damage in hippocampal slices, and this neuroprotective effect seems to be related to its thiol-peroxidase-like activity. Indeed, we previously demonstrated that (PhSe)₂ inhibited human LDL oxidation and that this phenomenon was related to its GPx-like activity (de Bem et al., 2008). The GPx-like activity of (PhSe)₂ seems to be kinetically identical to that of the enzyme reaction (Nogueira et al., 2004, Barbosa et al., 2006, Nogueira and Rocha, 2011). In brief, (PhSe)₂ reacts with 2 thiol equivalents, such as GSH, to generate a product characterized as selenol. The selenol reacts with H₂O₂ or organic peroxides to form water and a seleninic acid, which spontaneously produces another molecule of water and turns to (PhSe)₂ (Nogueira et al., 2004, Posser et al., 2008, Nogueira and Rocha, 2011). Then, (PhSe)₂ might be conferring its protective effect by decomposing lipid hydroperoxides.

Of particular pharmacological importance, previous studies suggest that (PhSe)₂, a very lipophilic compound that can cross the BBB, leads to increasing selenium levels into the brain after acute and chronic treatments (Jacques-Silva et al., 2001, Maciel et al., 2003). Furthermore, toxicological studies have been conducted to establish the safety of this organoselenium compound. The level of toxicity induced by (PhSe)₂ depends on the route of administration and the species (rat, mice, or rabbits) (Nogueira et al., 2003, de Bem et al., 2007, Straliotto et al., 2010). In fact, the antioxidant effect of (PhSe)₂ showed in this study was obtained within a very safe dose range. Recently, Savegnago et al. (2007) characterized that the LD₅₀ for (PhSe)₂ to mice by the oral route was >312 mg/kg and that this oral treatment did not provide evidence for renal or hepatic toxicity. The quantification of plasma levels of (PhSe)₂ after oral administration of 500 mg/kg in mice, showed a (PhSe)₂ peak at 30 minutes post dosing of approximately 10 µg/mL (Prigol et al., 2009).

Conclusions

In conclusion, the present findings constitute an *in vivo* evidence for neuroprotective role of low dose of (PhSe)₂ in cortico-cerebral oxidative stress induced by hypercholesterolemic diet in the 3-months-old LDL^{r/-} mice. The antioxidant effect of (PhSe)₂ is, at least in part, associated to the involvement of glutathione system. Further studies are warranted to elucidate the precise mechanism of (PhSe)₂ antioxidant effect in cerebral cortex of LDL^{r/-} mice. In addition, to its protective effects in cardiovascular diseases associated with hypercholesterolemia showed in ours previous studies, based on our findings (PhSe)₂ can be pointed as a promising neuroprotective molecule against hypercholesterolemia-induced effects deleterious on brain.

Acknowledgments

This work was supported by grants from the Brazilian institutions Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), and the Fundação de Apoio à Pesquisa do Estado de Santa Catarina (FAPESC).

References

- Amin KA, Kamel HH, Abd Eltawab MA (2011) The relation of high fat diet, metabolic disturbances and brain oxidative dysfunction: modulation by hydroxy citric acid. *Lipids Health Dis* 10:74.
- Barbosa NB, Rocha JB, Soares JC, Wondracek DC, Goncalves JF, Schetinger MR, Nogueira CW (2008) Dietary diphenyl diselenide reduces the STZ-induced toxicity. *Food Chem Toxicol* 46:186-194.
- Barbosa NB, Rocha JB, Wondracek DC, Perottoni J, Zeni G, Nogueira CW (2006) Diphenyl diselenide reduces temporarily hyperglycemia: possible relationship with oxidative stress. *Chem Biol Interact* 163:230-238.
- Borges LP, Nogueira CW, Panatieri RB, Rocha JB, Zeni G (2006) Acute liver damage induced by 2-nitropropane in rats: effect of diphenyl diselenide on antioxidant defenses. *Chem Biol Interact* 160:99-107.
- Brigelius-Flohe R (1999) Tissue-specific functions of individual glutathione peroxidases. *Free Radic Biol Med* 27:951-965.
- Carlberg I, Mannervik B (1985) Glutathione reductase. *Methods Enzymol* 113:484-490.
- Colell A, Garcia-Ruiz C, Morales A, Ballesta A, Ookhtens M, Rodes J, Kaplowitz N, Fernandez-Checa JC (1997) Transport of reduced glutathione in hepatic mitochondria and mitoplasts from ethanol-treated rats: effect of membrane physical properties and S-adenosyl-L-methionine. *Hepatology* 26:699-708.
- Crisby M, Rahman SM, Sylven C, Winblad B, Schultzberg M (2004) Effects of high cholesterol diet on gliosis in apolipoprotein E knockout mice. Implications for Alzheimer's disease and stroke. *Neurosci Lett* 369:87-92.
- de Bem AF, de Lima Portella R, Farina M, Perottoni J, Paixao MW, Nogueira CW, Teixeira Rocha JB (2007) Low toxicity of diphenyl diselenide in rabbits: a long-term study. *Basic Clin Pharmacol Toxicol* 101:47-55.
- de Bem AF, Farina M, Portella Rde L, Nogueira CW, Dinis TC, Laranjinha JA, Almeida LM, Rocha JB (2008) Diphenyl diselenide, a simple glutathione peroxidase mimetic, inhibits human LDL oxidation in vitro. *Atherosclerosis* 201:92-100.
- de Bem AF, Portella Rde L, Colpo E, Duarte MM, Frediane A, Taube PS, Nogueira CW, Farina M, da Silva EL, Teixeira Rocha JB (2009) Diphenyl diselenide decreases serum levels of total

- cholesterol and tissue oxidative stress in cholesterol-fed rabbits. *Basic Clin Pharmacol Toxicol* 105:17-23.
- de Freitas AS, Rocha JB (2011) Diphenyl diselenide and analogs are substrates of cerebral rat thioredoxin reductase: a pathway for their neuroprotective effects. *Neurosci Lett* 503:1-5.
- De Haan JB, Crack PJ, Flentjar N, Iannello RC, Hertzog PJ, Kola I (2003) An imbalance in antioxidant defense affects cellular function: the pathophysiological consequences of a reduction in antioxidant defense in the glutathione peroxidase-1 (Gpx1) knockout mouse. *Redox Rep* 8:69-79.
- de Oliveira J, Hort MA, Moreira EL, Glaser V, Ribeiro-do-Valle RM, Prediger RD, Farina M, Latini A, de Bem AF (2011) Positive correlation between elevated plasma cholesterol levels and cognitive impairments in LDL receptor knockout mice: relevance of cortico-cerebral mitochondrial dysfunction and oxidative stress. *Neuroscience* 197:99-106.
- Di Paolo G, Kim TW (2011) Linking lipids to Alzheimer's disease: cholesterol and beyond. *Nat Rev Neurosci* 12:284-296.
- Dufouil C, Richard F, Fievet N, Dartigues JF, Ritchie K, Tzourio C, Amouyel P, Alperovitch A (2005) APOE genotype, cholesterol level, lipid-lowering treatment, and dementia: the Three-City Study. *Neurology* 64:1531-1538.
- Ellman GL (1959) Tissue sulphhydryl groups. *Arch Biochem Biophys* 82:70-77.
- Fernandez A, Llacuna L, Fernandez-Checa JC, Colell A (2009) Mitochondrial cholesterol loading exacerbates amyloid beta peptide-induced inflammation and neurotoxicity. *J Neurosci* 29:6394-6405.
- Griffith OW, Mulcahy RT (1999) The enzymes of glutathione synthesis: gamma-glutamylcysteine synthetase. *Adv Enzymol Relat Areas Mol Biol* 73:209-267, xii.
- Grundy SM, Cleeman JI, Merz CN, Brewer HB, Jr., Clark LT, Hunnighake DB, Pasternak RC, Smith SC, Jr., Stone NJ (2004) Implications of recent clinical trials for the National Cholesterol Education Program Adult Treatment Panel III guidelines. *Circulation* 110:227-239.
- Hajjar I, Schumpert J, Hirth V, Wieland D, Eleazer GP (2002) The impact of the use of statins on the prevalence of dementia and the progression of cognitive impairment. *J Gerontol A Biol Sci Med Sci* 57:M414-418.

- Harish G, Venkateshappa C, Mahadevan A, Pruthi N, Srinivas Bharath MM, Shankar SK (2011) Glutathione metabolism is modulated by postmortem interval, gender difference and agonal state in postmortem human brains. *Neurochem Int* 59:1029-1042.
- Hort MA, Straliotto MR, Netto PM, da Rocha JB, de Bem AF, Ribeiro-do-Valle RM (2011) Diphenyl diselenide effectively reduces atherosclerotic lesions in LDL^r -/- mice by attenuation of oxidative stress and inflammation. *J Cardiovasc Pharmacol* 58:91-101.
- Iadecola C (2004) Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat Rev Neurosci* 5:347-360.
- Ishibashi S, Brown MS, Goldstein JL, Gerard RD, Hammer RE, Herz J (1993) Hypercholesterolemia in low density lipoprotein receptor knockout mice and its reversal by adenovirus-mediated gene delivery. *J Clin Invest* 92:883-893.
- Jackson SM, Ericsson J, Edwards PA (1997) Signaling molecules derived from the cholesterol biosynthetic pathway. *Subcell Biochem* 28:1-21.
- Jacques-Silva MC, Nogueira CW, Broch LC, Flores EM, Rocha JB (2001) Diphenyl diselenide and ascorbic acid changes deposition of selenium and ascorbic acid in liver and brain of mice. *Pharmacol Toxicol* 88:119-125.
- Jick H, Zornberg GL, Jick SS, Seshadri S, Drachman DA (2000) Statins and the risk of dementia. *Lancet* 356:1627-1631.
- Kalayci R, Kaya M, Uzun H, Bilgic B, Ahishali B, Arican N, Elmas I, Kucuk M (2009) Influence of hypercholesterolemia and hypertension on the integrity of the blood-brain barrier in rats. *Int J Neurosci* 119:1881-1904.
- Kim SJ, Park C, Han AL, Youn MJ, Lee JH, Kim Y, Kim ES, Kim HJ, Kim JK, Lee HK, Chung SY, So H, Park R (2009) Ebselen attenuates cisplatin-induced ROS generation through Nrf2 activation in auditory cells. *Hear Res* 251:70-82.
- Lewington S, Whitlock G, Clarke R, Sherliker P, Emberson J, Halsey J, Qizilbash N, Peto R, Collins R (2007) Blood cholesterol and vascular mortality by age, sex, and blood pressure: a meta-analysis of individual data from 61 prospective studies with 55,000 vascular deaths. *Lancet* 370:1829-1839.
- Lima CF, Fernandes-Ferreira M, Pereira-Wilson C (2006) Phenolic compounds protect HepG2 cells from oxidative damage: relevance of glutathione levels. *Life Sci* 79:2056-2068.

- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951) Protein measurement with the Folin phenol reagent. *J Biol Chem* 193:265-275.
- Lu J, Wu DM, Zheng YL, Hu B, Zhang ZF, Shan Q, Zheng ZH, Liu CM, Wang YJ (2010) Quercetin activates AMP-activated protein kinase by reducing PP2C expression protecting old mouse brain against high cholesterol-induced neurotoxicity. *J Pathol* 222:199-212.
- Lu J, Wu DM, Zheng YL, Sun DX, Hu B, Shan Q, Zhang ZF, Fan SH (2009) Trace amounts of copper exacerbate beta amyloid-induced neurotoxicity in the cholesterol-fed mice through TNF-mediated inflammatory pathway. *Brain Behav Immun* 23:193-203.
- Lusis AJ (2000) Atherosclerosis. *Nature* 407:233-241.
- Maciel EN, Flores EM, Rocha JB, Folmer V (2003) Comparative deposition of diphenyl diselenide in liver, kidney, and brain of mice. *Bull Environ Contam Toxicol* 70:470-476.
- Mari M, Colell A, Morales A, Caballero F, Moles A, Fernandez A, Terrones O, Basanez G, Antonsson B, Garcia-Ruiz C, Fernandez-Checa JC (2008) Mechanism of mitochondrial glutathione-dependent hepatocellular susceptibility to TNF despite NF-kappaB activation. *Gastroenterology* 134:1507-1520.
- McCommis KS, McGee AM, Laughlin MH, Bowles DK, Baines CP (2011) Hypercholesterolemia increases mitochondrial oxidative stress and enhances the MPT response in the porcine myocardium: beneficial effects of chronic exercise. *Am J Physiol Regul Integr Comp Physiol* 301:R1250-1258.
- Meotti FC, Stangerlin EC, Zeni G, Nogueira CW, Rocha JB (2004) Protective role of aryl and alkyl diselenides on lipid peroxidation. *Environ Res* 94:276-282.
- Montilla P, Espejo I, Munoz MC, Bujalance I, Munoz-Castaneda JR, Tunez I (2006) Protective effect of red wine on oxidative stress and antioxidant enzyme activities in the brain and kidney induced by feeding high cholesterol in rats. *Clin Nutr* 25:146-153.
- Moser KV, Reindl M, Blasig I, Humpel C (2004) Brain capillary endothelial cells proliferate in response to NGF, express NGF receptors and secrete NGF after inflammation. *Brain Res* 1017:53-60.
- Nogueira CW, Meotti FC, Curte E, Pilissao C, Zeni G, Rocha JB (2003) Investigations into the potential neurotoxicity induced by diselenides in mice and rats. *Toxicology* 183:29-37.

- Nogueira CW, Zeni G, Rocha JB (2004) Organoselenium and organotellurium compounds: toxicology and pharmacology. *Chem Rev* 104:6255-6285.
- Nogueira CW, Rocha JB (2011) Toxicology and pharmacology of selenium: emphasis on synthetic organoselenium compounds. *Arch Toxicol* 85:1313-1359.
- Ohkawa H, Ohishi N, Yagi K (1979) Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 95:351-358.
- Oliveira HC, Cocco RG, Alberici LC, Maciel EN, Salerno AG, Dorighello GG, Velho JA, de Faria EC, Vercesi AE (2005) Oxidative stress in atherosclerosis-prone mouse is due to low antioxidant capacity of mitochondria. *FASEB J* 19:278-280.
- Paim BA, Velho JA, Castilho RF, Oliveira HC, Vercesi AE (2008) Oxidative stress in hypercholesterolemic LDL (low-density lipoprotein) receptor knockout mice is associated with low content of mitochondrial NADP-linked substrates and is partially reversed by citrate replacement. *Free Radic Biol Med* 44:444-451.
- Paulmier, C. (1986) Selenium reagents and intermediates. In: *Organic Synthesis*. Pergamon, Oxford.
- Pappolla MA, Smith MA, Bryant-Thomas T, Bazan N, Petanceska S, Perry G, Thal LJ, Sano M, Refolo LM (2002) Cholesterol, oxidative stress, and Alzheimer's disease: expanding the horizons of pathogenesis. *Free Radic Biol Med* 33:173-181.
- Posser T, Franco JL, dos Santos DA, Rigon AP, Farina M, Dafre AL, Teixeira Rocha JB, Leal RB (2008) Diphenyl diselenide confers neuroprotection against hydrogen peroxide toxicity in hippocampal slices. *Brain Res* 1199:138-147.
- Prigol M, Schumacher RF, WayneNogueira C, Zeni G (2009) Convulsant effect of diphenyl diselenide in rats and mice and its relationship to plasma levels. *Toxicol Lett* 189:35-39.
- Puntel RL, Roos DH, Paixao MW, Braga AL, Zeni G, Nogueira CW, Rocha JB (2007) Oxalate modulates thiobarbituric acid reactive species (TBARS) production in supernatants of homogenates from rat brain, liver and kidney: effect of diphenyl diselenide and diphenyl ditelluride. *Chem Biol Interact* 165:87-98.
- Raes M, Michiels C, Remacle J (1987) Comparative study of the enzymatic defense systems against oxygen-derived free radicals: the key role of glutathione peroxidase. *Free Radic Biol Med* 3:3-7.

- Ramirez C, Sierra S, Tercero I, Vazquez JA, Pineda A, Manrique T, Burgos JS (2011) ApoB100/LDLR-/ hypercholesterolaemic mice as a model for mild cognitive impairment and neuronal damage. *PLoS One* 6:e22712.
- Ran Q, Gu M, Van Remmen H, Strong R, Roberts JL, Richardson A (2006) Glutathione peroxidase 4 protects cortical neurons from oxidative injury and amyloid toxicity. *J Neurosci Res* 84:202-208.
- Rodriguez EG, Dodge HH, Birzescu MA, Stoehr GP, Ganguli M (2002) Use of lipid-lowering drugs in older adults with and without dementia: a community-based epidemiological study. *J Am Geriatr Soc* 50:1852-1856.
- Rosa RM, Roesler R, Braga AL, Saffi J, Henriques JA (2007) Pharmacology and toxicology of diphenyl diselenide in several biological models. *Braz J Med Biol Res* 40:1287-1304.
- Rossato JI, Ketzer LA, Centuriao FB, Silva SJ, Ludtke DS, Zeni G, Braga AL, Rubin MA, Rocha JB (2002) Antioxidant properties of new chalcogenides against lipid peroxidation in rat brain. *Neurochem Res* 27:297-303.
- Savegnago L, Jesse CR, Pinto LG, Rocha JB, Nogueira CW (2007) Diphenyl diselenide attenuates acute thermal hyperalgesia and persistent inflammatory and neuropathic pain behavior in mice. *Brain Res* 1175:54-59.
- Simons K, Ikonen E (2000) How cells handle cholesterol. *Science* 290:1721-1726.
- Simons M, Schwarzler F, Lutjohann D, von Bergmann K, Beyreuther K, Dichgans J, Wormstall H, Hartmann T, Schulz JB (2002) Treatment with simvastatin in normocholesterolemic patients with Alzheimer's disease: A 26-week randomized, placebo-controlled, double-blind trial. *Ann Neurol* 52:346-350.
- Sparks DL, Connor DJ, Sabbagh MN, Petersen RB, Lopez J, Browne P (2006) Circulating cholesterol levels, apolipoprotein E genotype and dementia severity influence the benefit of atorvastatin treatment in Alzheimer's disease: results of the Alzheimer's Disease Cholesterol-Lowering Treatment (ADCLT) trial. *Acta Neurol Scand Suppl* 185:3-7.
- Sparks DL, Kuo YM, Roher A, Martin T, Lukas RJ (2000) Alterations of Alzheimer's disease in the cholesterol-fed rabbit, including vascular inflammation. Preliminary observations. *Ann N Y Acad Sci* 903:335-344.

- Sparks DL, Sabbagh MN, Connor DJ, Lopez J, Launer LJ, Petanceska S, Browne P, Wassar D, Johnson-Traver S, Lochhead J, Ziolkowski C (2005) Atorvastatin therapy lowers circulating cholesterol but not free radical activity in advance of identifiable clinical benefit in the treatment of mild-to-moderate AD. *Curr Alzheimer Res* 2:343-353.
- Sparks DL, Schreurs BG (2003) Trace amounts of copper in water induce beta-amyloid plaques and learning deficits in a rabbit model of Alzheimer's disease. *Proc Natl Acad Sci U S A* 100:11065-11069.
- Stokes KY, Cooper D, Tailor A, Granger DN (2002) Hypercholesterolemia promotes inflammation and microvascular dysfunction: role of nitric oxide and superoxide. *Free Radic Biol Med* 33:1026-1036.
- Straliotto MR, Mancini G, de Oliveira J, Nazari EM, Muller YM, Dafre A, Ortiz S, Silva EL, Farina M, Latini A, Rocha JB, de Bem AF (2010) Acute exposure of rabbits to diphenyl diselenide: a toxicological evaluation. *J Appl Toxicol* 30:761-768.
- Tamasi V, Jeffries JM, Arteel GE, Falkner KC (2004) Ebselen augments its peroxidase activity by inducing nrf-2-dependent transcription. *Arch Biochem Biophys* 431:161-168.
- Thirumangalakudi L, Prakasam A, Zhang R, Bimonte-Nelson H, Sambamurti K, Kindy MS, Bhat NR (2008) High cholesterol-induced neuroinflammation and amyloid precursor protein processing correlate with loss of working memory in mice. *J Neurochem* 106:475-485.
- Ullrich C, Pirchl M, Humpel C (2010) Hypercholesterolemia in rats impairs the cholinergic system and leads to memory deficits. *Mol Cell Neurosci* 45:408-417.
- Wendel A (1981) Glutathione peroxidase. *Methods Enzymol* 77:325-333.
- Wilson SR, Zucker PA, Huang RRC, Spector A (1989) Development of synthetic compounds with glutathione peroxidase activity. *J Am Chem Soc* 111: 5936-5939.
- Wolozin B (2004a) Cholesterol and the biology of Alzheimer's disease. *Neuron* 41:7-10.
- Wolozin B (2004b) Cholesterol, statins and dementia. *Curr Opin Lipidol* 15:667-672.
- Wolozin B, Wang SW, Li NC, Lee A, Lee TA, Kazis LE (2007) Simvastatin is associated with a reduced incidence of dementia and Parkinson's disease. *BMC Med* 5:20

Figures and Legends

Groups	Total Cholesterol (mg/dL)	non-HDL cholesterol (mg/dL)
control + vehicle	211.6 ± 22.25	183.0 ± 22.70
control + (PhSe) ₂	197.8 ± 13.45	183.9 ± 12.35
cholesterol + vehicle	833.9 ± 56.69*	764.7 ± 51.72*
cholesterol + (PhSe) ₂	864.4 ± 42.51	834.5 ± 58.55

Table 1: Effect of (PhSe)₂ on plasma total cholesterol levels and non-HDL-cholesterol levels in the LDLr^{-/-} mice fed standard or cholesterol-enriched diet . Each value represents the mean ± S.E.M. of four to five animals in each group. *P≤0.05 compared to 3-month-old LDLr^{-/-} mice fed standard diet. (Two-way ANOVA followed by Duncan's post-hoc test).

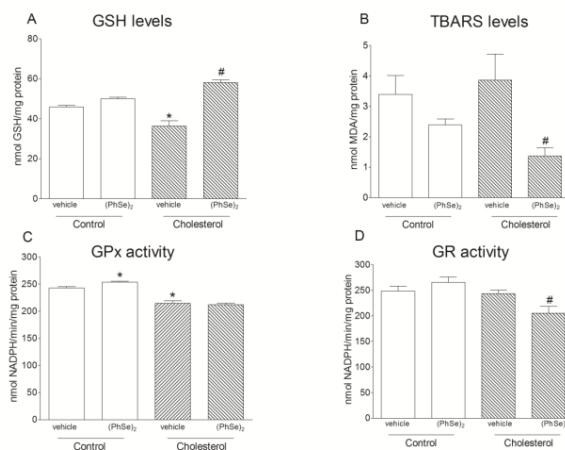


Figure 1: Effects of (PhSe)₂ on cerebral cortex oxidative stress parameters in the LDLr^{-/-} mice fed standard or cholesterol-enriched diet: A) GSH levels, B) TBARS levels, C) GPx activity and D) GR activity. Each value represents the mean ± S.E.M. of four to six animals in each group. *P≤0.05 compared to 3-month-old LDLr^{-/-} mice fed standard diet, #P≤0.05 compared to 3-month-old LDLr^{-/-} mice fed cholesterol-enriched diet (Two-way ANOVA followed by Duncan's post-hoc test).

5 DISCUSSÃO

A condição de hipercolesterolemia (monogênica e multifatorial) afeta 1 em cada 20 indivíduos na população em geral, por outro lado, a frequência da hipercolesterolemia familiar (HF) é de 1 em 500 para heterozigotos. Homozigotos são raros, com uma frequência de 1 em um milhão. A HF é uma patologia autossômica codominante caracterizada pela elevação isolada do colesterol presente na LDL plasmática e fortemente associada com alto risco de doença cardiovascular prematura (Brown e Goldstein, 1986; Civeira, 2004; Setia et al., 2012). Nos últimos anos, crescentes evidências clínicas e patológicas indicam que elevados níveis de LDL-colesterol podem contribuir para o desenvolvimento de prejuízos cognitivos e demência (Pappolla et al., 2002; Mielke et al., 2005, Whitmer et al., 2005, Zambon et al., 2010). Nesse contexto, uma ferramenta útil para o estudo do impacto dos altos níveis plasmáticos de colesterol sobre parâmetros metabólicos e funcionais em diferentes órgãos é o uso de camundongos $LDLr^{-/-}$, os quais representam muito bem um modelo de HF (Ishibashi et al., 1993; Zadelaar et al., 2007).

De fato, Mulder e colaboradores (2004) demonstraram que camundongos $LDLr^{-/-}$ apresentam prejuízo de aprendizado e memória quando comparados com camundongos controles em diversos testes comportamentais que avaliam memória espacial e de trabalho. Neste sentido, os resultados apresentados na primeira parte desta dissertação, referem-se à avaliação da função cognitiva, bem como da função mitocondrial e antioxidante em córtex cerebral de camundongos $LDLr^{-/-}$ expostos à dieta padrão ou hipercolesterolêmica durante 30 dias. O conjunto de resultados apresentados no Artigo 1 demonstram que independentemente da dieta adotada os camundongos $LDLr^{-/-}$ apresentam prejuízo de aprendizado e memória espacial no teste de localização do objeto. Além disso, a dieta hipercolesterolêmica causou inibição na atividade dos complexos I e II da CR no córtex cerebral de camundongos $LDLr^{-/-}$, evento que foi associado com lipoperoxidação, diminuição dos níveis de GSH e desequilíbrio na atividade das enzimas integrantes do sistema antioxidante dependente da GSH. Estes últimos resultados caracterizaram uma situação de estresse oxidativo no córtex cerebral destes camundongos hipercolesterolêmicos. De particular importância, a disfunção mitocondrial observada no córtex cerebral dos camundongos foi negativamente correlacionada com os níveis plasmáticos de colesterol.

Dados da literatura demonstram que a ingestão de uma dieta rica em colesterol prejudica o aprendizado espacial e a habilidade de retenção da memória de longo prazo em roedores (Granholm et al., 2008; Ullrich et al., 2010; Freeman et al., 2011). Nossos dados demonstram que camundongos $LDLr^{-/-}$ alimentados com dieta padrão ou hipercolesterolêmica não são capazes de identificar uma alteração espacial no campo aberto. A memória de localização/relocalização de objeto é proposta como um modelo de memória espacial com apenas um treinamento (Murai et al., 2007). O teste de localização do objeto é amplamente usado para avaliar a função cognitiva em camundongos transgênicos, levando em consideração a preferência de camundongos normais pelo objeto que foi movido de sua posição anterior (Faivre et al., 2011). Vários estudos demonstraram que lesões corticais levam ao prejuízo do desempenho neste teste em diferentes protocolos experimentais, uma vez que o córtex pré-frontal medial é importante para a memória espacial de trabalho (Brozoski et al., 1979; Ennaceur et al., 1997; Nelson et al., 2011). No entanto, uma desvantagem deste e de outros conhecidos testes cognitivos é fornecer apenas um índice qualitativo (aprender ou não aprender), então, provavelmente por esta razão não podemos determinar uma maior redução na função cognitiva dos camundongos $LDLr^{-/-}$ que foram expostos à dieta hipercolesterolêmica.

Como a homeostase do colesterol cerebral é independente do colesterol plasmático, o exato mecanismo responsável pelo prejuízo cognitivo induzido pela hipercolesterolemia ainda permanece desconhecido (Elder et al., 2007; Di Paolo e Kim, 2011). Uma possível explicação foi proposta por Evola e colaboradores (2010) usando camundongos ApoE $^{-/-}$ e por Thirumangalakudi e colaboradores (2008) usando camundongos $LDLr^{-/-}$ expostos á uma dieta com alto teor de gordura e/ou colesterol. Estes autores propuseram que um nocivo ciclo oxidativo e inflamatório na vasculatura poderia ter consequências deletérias para função cerebral e cognição. Na mesma linha de evidência, Hafezi-Moghadam e colaboradores (2007) demonstraram que a deleção genética da ApoE em camundongos idosos induz um efeito desestabilizador sobre a microvasculatura cerebral levando a disfunção da BHE. Portanto, alterações na BHE decorrentes da hipercolesterolemia resultam em extravasamento de componentes séricos para dentro e através da parede dos pequenos vasos cerebrais e recrutamento de células imune inflamatórias da circulação sanguínea, causando dano neuronal e glial com persistente ativação de micróglia e

astrócitos. Estes eventos podem ser direta ou indiretamente responsáveis pelo prejuízo cognitivo (Rapp et al., 2008).

O processo neuroinflamatório também pode levar ao aumento da produção de ERO, as quais são prejudiciais para os neurônios. As mitocôndrias são vítimas de extensivo dano oxidativo, enquanto a inflamação provavelmente também contribui intensificando a disfunção mitocondrial (Witte et al., 2010). Os neurônios são vulneráveis a defeitos na função mitocondrial porque requerem altos níveis de energia para sua sobrevivência e função especializada (Chen e Chan, 2006). Um grande número de evidências suporta o papel crucial da disfunção mitocondrial no desenvolvimento de prejuízos cognitivos e progressão para demência; com a diminuição no metabolismo energético e produção de ERO como principais eventos desencadeadores (Lin e Beal, 2006; Moreira et al., 2010). A dieta hipercolesterolêmica promoveu um aumento de 10 vezes no colesterol dos camundongos $LDLr^{-/-}$, o que provocou uma redução na atividade dos complexos I e II no córtex cerebral destes camundongos. De particular importância, nossos resultados apresentam evidência que esta inibição na atividade dos complexos I e II foi negativamente correlacionada com os níveis plasmáticos de colesterol. No entanto, os camundongos $LDLr^{-/-}$ alimentados com dieta padrão apesar de apresentarem prejuízo cognitivo; uma elevação de duas vezes nos níveis de colesterol plasmático não foi capaz de causar disfunção mitocondrial, bem como estresse oxidativo. Portanto, o aumento do colesterol sérico com o auxílio de uma dieta rica em colesterol acelera as consequências de um nocivo ciclo oxidativo e inflamatório na vasculatura.

Recentes estudos conduzidos em mitocôndrias isoladas de vários tecidos (cérebro, fígado e rim) de camundongos $LDLr^{-/-}$ demonstraram que a disfunção mitocondrial e estresse oxidativo causados pela hipercolesterolemia são devido ao baixo conteúdo de nucleotídeos piridinas no estado reduzido (NADPH), os quais são depletados pela lipogênese aumentada (Oliveira et al., 2005; Paim et al., 2008). As mitocôndrias são ao mesmo tempo alvos e fontes biologicamente importantes de ERO principalmente no SNC (Kowaltowski e Vercesi, 1999; Kowaltowski et al., 2001) e por outro lado, o aumento dos níveis de ERO causa disfunção mitocondrial (Tan et al., 1998). Entretanto, a produção de ERO aumenta quando os complexos respiratórios estão inibidos (Leonard e Schapira, 2000; Fontanesi et al., 2009).

Estresse oxidativo celular e suas consequências são também características bem estabelecidas da patofisiologia de diversas patologias neurodegenerativas, e o SNC é especialmente suscetível ao

dano induzido por ERO (Harish et al., 2011; Sultana e Butterfield, 2011). Nossos resultados indicaram um desequilíbrio na atividade das enzimas GPx e GR, caracterizado por uma diminuição na atividade da GPx e um aumento da atividade da GR no córtex cerebral dos camundongos expostos à dieta hipercolesterolêmica, o que pode estar relacionado com o estresse oxidativo induzido pela hipercolesterolemia, principalmente por causar lipoperoxidação e prejudicar o sistema antioxidante dependente da GSH. Muitos estudos tem mostrado uma relação entre a exposição a dietas com alto teor de gordura e/ou colesterol e estresse oxidativo em cérebros de camundongos e ratos (Crisby et al., 2004; Montilla et al., 2006) e os nossos resultados podem estabelecer o papel da disfunção mitocondrial no consequente desequilíbrio oxidativo induzido pela hipercolesterolemia.

GSH e as GPxs constituem um dos mais importantes sistemas de defesa antioxidante (Raes et al., 1987; Brigelius-Flohe, 1999). A GSH é um importante antioxidante e sequestrador de ERO intracelular, proporcionando a célula neuronal uma importante proteção contra os danos oxidativos (Dringen e Hirrlinger, 2003). Estudos anteriores demonstraram que o sistema GSH pode ser ativado como uma resposta ao estresse oxidativo em cérebros de indivíduos com doenças neurodegenerativas (Lovell et al., 1995; Aksenov et al., 2001). Adicionalmente, a hipercolesterolemia desencadeia estresse oxidativo por aumentar a geração de ERO levando à depleção dos níveis de GSH, além de prejudicar o transporte mitocondrial de GSH, resultando em depleção de GSH mitocondrial e estresse oxidativo mitocondrial (Colell et al., 1997; Mari et al., 2008; Fernandez et al., 2009). Corroborando com estas informações, Esposito e colaboradores (2000) demonstraram que mitocôndrias isoladas de fígados de camundongos deficientes de GPx-1 têm altas taxas de produção de H_2O_2 e reduzida taxa de respiração mitocondrial. A GPx é um enzima de defesa antioxidante que pode diretamente detoxificar H_2O_2 ou hidroperóxidos à custa de GSH. Várias evidências propõem que a GPx funciona como uma proteção primária contra estresse oxidativo agudo, principalmente em situações neuropatológicas (Brigelius-Flohe, 1999; De Haan et al., 2003; Ran et al., 2006). De acordo com nossos dados podemos indicar uma nova evidência sobre a associação entre níveis plasmáticos elevados de colesterol e prejuízos cognitivos em camundongos $LDLr^{-/-}$ hipercolesterolêmicos, o que foi mediado em parte pela disfunção mitocondrial e estresse oxidativo também encontrados no córtex cerebral destes camundongos expostos à uma dieta com alto teor de colesterol.

Vários estudos têm reportado efeitos benéficos dos compostos orgânicos de selênio contra condições patológicas associadas ao estresse oxidativo (inflamação, diabetes, neurotoxicidade e hepatotoxicidade) (Nogueira e Rocha, 2011). Dentre estes compostos, o (PhSe)₂, primeiramente utilizado como intermediário em reações de química orgânica, demonstrou ações farmacológicas interessantes em diversos modelos experimentais relacionados à produção exacerbada de espécies reativas (Wilson et al., 1989; Nogueira et al., 2004; Nogueira e Rocha, 2010), especialmente em modelos de hipercolesterolemia, apresentando um potencial efeito protetor em doenças cardiovasculares (de Bem et al., 2008; de Bem et al., 2009; Hort et al., 2011). O conjunto de dados apresentados no manuscrito 1 indicam um significativo efeito neuroprotetor do tratamento com (PhSe)₂ sobre o estresse oxidativo induzido pela hipercolesterolemia no córtex cerebral dos camundongos LDLr^{-/-} expostos à dieta hipercolesterolêmica. Este efeito antioxidante foi relacionado ao aumento dos níveis de GSH e diminuição da lipoperoxidação.

O (PhSe)₂, atua como mimético da GPx, reagindo com tióis para gerar selenol, o intermediário responsável pelo efeito antioxidante (Mugesh e Singh, 2000). De fato, Posser e colaboradores (2008) demonstraram efeito neuroprotetor do (PhSe)₂ em fatias hipocampais expostas ao dano oxidativo induzido pelo H₂O₂, e este efeito foi relacionado a sua atividade mimética da GPx. Nesse sentido, o (PhSe)₂ pode exercer sua proteção por decompor lipoperóxidos. O tratamento oral de 30 dias com (PhSe)₂ na dose de 1 mg/kg, não foi capaz de modificar a atividade da GPx no córtex cerebral dos camundongos LDLr^{-/-} expostos à dieta hipercolesterolêmica, no entanto aumentou os níveis de GSH, seu substrato, o que possivelmente contribui para o aumento da detoxificação dos produtos de lipoperoxidação, diminuindo os níveis de MDA. Além disso, a diminuição na atividade da GR, enzima responsável por regenerar a forma reduzida da GSH, também foi observada no córtex cerebral dos camundongos LDLr^{-/-} expostos à dieta hipercolesterolêmica tratados com (PhSe)₂. Esta redução pode ser resultado do aumento na síntese de GSH nestes tecidos, podendo estar relacionada ao aumento da atividade da enzima gama-glutamilcisteína sintetase, passo limitante na síntese de GSH. Desta forma, acreditamos que os efeitos protetores do (PhSe)₂ frente ao estresse oxidativo induzido pela hipercolesterolemia no córtex cerebral dos camundongos LDLr^{-/-} expostos à dieta hipercolesterolêmica vão além de sua atividade mimética da GPx.

Como já demonstrado recentemente para o ebselen, outro importante composto orgânico de selênio (Tamasi et al., 2004; Kim et al., 2009); acreditamos que o (PhSe)₂ possa ser capaz de promover a translocação do fator Nrf-2, sinalizando diversas respostas celulares vinculadas à ativação dos elementos de resposta antioxidante (ARE), entre elas o aumento nos níveis de GSH e na atividade de enzimas antioxidantes. A hipótese para este mecanismo é que o (PhSe)₂, da mesma forma que o ebselen, reage com grupamentos sulfidrila da proteína inibitória Keap-1, liberando o Nrf-2 e ativando vias de sinalização vinculadas ao sistema redox (Zhao e Holmgren, 2002; Tamasi et al., 2004). Confirmando nossos dados, estudos anteriores verificaram um aumento na concentração de GSH em diferentes tecidos, em animais tratados com (PhSe)₂ e expostos a danos oxidativo (Borges et al., 2006, de Bem et al., 2007; Straliootto et al., 2010). Nossos dados, constituem uma evidência do papel neuroprotetor *in vivo* do (PhSe)₂ sobre o estresse oxidativo induzido pela hipercolesterolemia no córtex cerebral dos camundongos LDLr^{-/-} expostos à dieta hipercolesterolêmica. E este efeito antioxidante do (PhSe)₂ em parte está associado com o envolvimento do sistema GSH. Portanto, em adição aos seus efeitos protetores demonstrados anteriormente em modelos de doenças cardiovasculares desencadeadas pela hipercolesterolemia, baseados em nossos dados podemos apontar o (PhSe)₂ como uma molécula neuroprotetora contra efeitos deletérios centrais induzidos pela hipercolesterolemia.

Finalmente, os resultados obtidos neste trabalho de mestrado apontam a utilização dos camundongos LDLr^{-/-} hipercolesterolêmicos como um modelo para o entendimento dos mecanismos envolvidos na patogênese de demências como a DA, bem como o composto orgânico de selênio, (PhSe)₂ como um efetivo agente neuroprotetor.

6 CONCLUSÕES

Os resultados obtidos neste trabalho indicam que:

- Os camundongos $\text{LDLr}^{-/-}$ hipercolesterolêmicos independentemente da dieta adotada apresentam prejuízo cognitivo em teste de memória espacial;
- A exposição de camundongos $\text{LDLr}^{-/-}$ à uma dieta hipercolesterolêmica durante 30 dias leva à diminuição da atividade dos complexos I e II da cadeia respiratória mitocondrial no córtex cerebral destes camundongos;
- A função mitocondrial nos camundongos $\text{LDLr}^{-/-}$ foi negativamente correlacionada com os respectivos níveis de colesterol destes camundongos;
- A disfunção cognitiva e mitocondrial nos camundongos $\text{LDLr}^{-/-}$ expostos à uma dieta hipercolesterolêmica foram associadas com um quadro de estresse oxidativo no córtex cerebral destes camundongos caracterizado por um desequilíbrio no sistema antioxidante vinculado a glutationa bem como peroxidação lipídica;
- O tratamento com composto orgânico de selênio, $(\text{PhSe})_2$ (1 mg/kg) durante 30 dias previou o estresse oxidativo no córtex cerebral de camundongos $\text{LDLr}^{-/-}$ expostos à uma dieta hipercolesterolêmica.

7 PERSPECTIVAS

Alguns aspectos relacionados a este trabalho ainda precisam ser melhor elucidados, desta forma, nosso grupo propõe algumas perspectivas:

- Primeiramente, avaliar o efeito do (PhSe)₂ contra o prejuízo cognitivo e a disfunção mitocondrial induzida pela dieta hipercolesterolêmica em camundongos LDLr^{-/-};
- Analisar a influência da hipercolesterolemia na toxicidade induzida pelo peptídeo β -amilóide em camundongos LDLr^{-/-}, através da injeção intracerebroventricular do peptídeo β -amilóide, a qual é uma ferramenta amplamente utilizada como modelo de Doença de Alzheimer em roedores;
- Desenvolver cultivo primário de neurônios hipocampais de camundongos LDLr^{-/-}, para analisar a resposta dos mesmos a toxicidade do peptídeo beta-amilóide.

8 REFERÊNCIAS BIBLIOGRÁFICAS

- Aksenov MY, Aksenova MV, Butterfield DA, Geddes JW, Markesberry WR. Protein oxidation in the brain in Alzheimer's disease. *Neuroscience* 103:373-383. 2001.
- Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Walter P. Molecular Biology of the Cell. In: Garland Science. Nova York: 1616 pp. 2002.
- Aliev G, Smith MA, de la Torre JC, Perry G. Mitochondria as a primary target for vascular hypoperfusion and oxidative stress in Alzheimer's disease. *Mitochondrion* 4: 649-663. 2004.
- Aliev G, Palacios HH, Walrafen B, Lipsitt AE, Obrenovich ME, Morales L. Brain mitochondria as a primary target in the development of treatment strategies for Alzheimer disease. *Int J Biochem Cell Biol* 41:1989-2004. 2009.
- Aliyev A, Chen SG, Seyidova D, Smith MA, Perry G, de la Torre J, Aliev G. Mitochondria DNA deletions in atherosclerotic hypoperfused brain microvessels as a primary target for the development of Alzheimer's disease. *J Neurol Sci* 229-230:285-292. 2005.
- Aoyama K, Watabe M, Nakaki T. Regulation of neuronal glutathione synthesis. *J Pharmacol Sci* 108:227-238. 2008.
- Babcock GT, Wikstrom M. Oxygen activation and the conservation of energy in cell respiration. *Nature* 356:301-309. 1992.
- Barber DA, Harris SR. Oxygen free radicals and antioxidants: a review. *Am Pharm NS34*:26-35. 1994.
- Barrientos A, Barros MH, Valnot I, Rotig A, Rustin P, Tzagoloff A. Cytochrome oxidase in health and disease. *Gene* 286:53-63. 2002.
- Beach TG, Wilson JR, Sue LI, Newell A, Poston M, Cisneros R, Pandya Y, Esh C, Connor DJ, Sabbagh M, Walker DG, Roher AE. Circle of Willis atherosclerosis: association with Alzheimer's disease, neuritic plaques and neurofibrillary tangles. *Acta Neuropathol* 113:13-21. 2007.
- Beeri MS, Rapp M, Silverman JM, Schmeidler J, Grossman HT, Fallon JT, Purohit DP, Perl DP, Siddiqui A, Lesser G, Rosendorff C, Haroutunian V. Coronary artery disease is associated with Alzheimer disease neuropathology in APOE4 carriers. *Neurology* 66:1399-1404. 2006.
- Blankenberg S, Rupprecht HJ, Bickel C, Torzewski M, Hafner G, Tiret L, Smieja M, Cambien F, Meyer J, Lackner KJ. Glutathione

- peroxidase 1 activity and cardiovascular events in patients with coronary artery disease. *N Engl J Med* 349:1605-1613. 2003.
- Borges LP, Borges VC, Moro AV, Nogueira CW, Rocha JB, Zeni G. Protective effect of diphenyl diselenide on acute liver damage induced by 2-nitropropane in rats. *Toxicology* 210:1-8. 2005.
- Borges LP, Nogueira CW, Panatieri RB, Rocha JB, Zeni G. Acute liver damage induced by 2-nitropropane in rats: effect of diphenyl diselenide on antioxidant defenses. *Chem Biol Interact* 160:99-107. 2006.
- Brigelius-Flohe R. Tissue-specific functions of individual glutathione peroxidases. *Free Radic Biol Med* 27:951-965. 1999.
- Brodsky SV, Gealekman O, Chen J, Zhang F, Togashi N, Crabtree M, Gross SS, Nasjletti A, Goligorsky MS. Prevention and reversal of premature endothelial cell senescence and vasculopathy in obesity-induced diabetes by ebselen. *Circ Res* 94:377-384. 2004.
- Brown MS, Goldstein JL. A receptor-mediated pathway for cholesterol homeostasis. *Science* 232:34-47. 1986.
- Brozoski TJ, Brown RM, Rosvold HE, Goldman PS. Cognitive deficit caused by regional depletion of dopamine in prefrontal cortex of rhesus monkey. *Science* 205:929-932. 1979.
- Burger M, Fachinetto R, Calegari L, Paixao MW, Braga AL, Rocha JB. Effects of age on reserpine-induced orofacial dyskinesia and possible protection of diphenyl diselenide. *Brain Res Bull* 64:339-345. 2004.
- Casserly I, Topol E. Convergence of atherosclerosis and Alzheimer's disease: inflammation, cholesterol, and misfolded proteins. *Lancet* 363:1139-1146. 2004.
- Centuriao FB, Corte CL, Paixao MW, Braga AL, Zeni G, Emanuelli T, Rocha JB. Effect of ebselen and organochalcogenides on excitotoxicity induced by glutamate in isolated chick retina. *Brain Res* 1039:146-152. 2005.
- Chance B, Sies H, Boveris A. Hydroperoxide metabolism in mammalian organs. *Physiol Rev* 59:527-605. 1979.
- Chen H, Chan DC. Critical dependence of neurons on mitochondrial dynamics. *Current opinion in cell biology* 18:453-459. 2006.
- Chew P, Yuen DY, Koh P, Stefanovic N, Febbraio MA, Kola I, Cooper ME, de Haan JB. Site-specific antiatherogenic effect of the antioxidant ebselen in the diabetic apolipoprotein E-deficient mouse. *Arterioscler Thromb Vasc Biol* 29:823-830. 2009.

- Circu ML, Aw TY. Glutathione and apoptosis. *Free Radic Res* 42:689-706. 2008.
- Civeira F. Guidelines for the diagnosis and management of heterozygous familial hypercholesterolemia. *Atherosclerosis* 173:55-68. 2004.
- Colell A, Garcia-Ruiz C, Morales A, Ballesta A, Ookhtens M, Rodes J, Kaplowitz N, Fernandez-Checa JC. Transport of reduced glutathione in hepatic mitochondria and mitoplasts from ethanol-treated rats: effect of membrane physical properties and S-adenosyl-L-methionine. *Hepatology* 26:699-708. 1997.
- Crisby M, Rahman SM, Sylven C, Winblad B, Schultzberg M. Effects of high cholesterol diet on gliosis in apolipoprotein E knockout mice. Implications for Alzheimer's disease and stroke. *Neurosci Lett* 369:87-92. 2004.
- Csont T, Bereczki E, Bencsik P, Fodor G, Gorbe A, Zvara A, Csonka C, Puskas LG, Santha M, Ferdinand P. Hypercholesterolemia increases myocardial oxidative and nitrosative stress thereby leading to cardiac dysfunction in apoB-100 transgenic mice. *Cardiovasc Res* 76:100-109. 2007.
- da Silva MH, da Rosa EJ, de Carvalho NR, Dobrachinski F, da Rocha JB, Mauriz JL, Gonzalez-Gallego J, Soares FA. Acute brain damage induced by acetaminophen in mice: Effect of diphenyl diselenide on oxidative stress and mitochondrial dysfunction. *Neurotox Res*. 2011.
- Daugherty A. Mouse models of atherosclerosis. *Am J Med Sci* 323:3-10. 2002.
- Davalos A. [New treatments in cerebrovascular diseases]. *Neurologia* 14 Suppl 6:77-83. 1999.
- Davignon J, Gregg RE, Sing CF. Apolipoprotein E polymorphism and atherosclerosis. *Arteriosclerosis* 8:1-21. 1988.
- de Bem AF, de Lima Portella R, Farina M, Perottoni J, Paixao MW, Nogueira CW, Teixeira Rocha JB. Low toxicity of diphenyl diselenide in rabbits: a long-term study. *Basic Clin Pharmacol Toxicol* 101:47-55. 2007.
- de Bem AF, Farina M, Portella Rde L, Nogueira CW, Dinis TC, Laranjinha JA, Almeida LM, Rocha JB. Diphenyl diselenide, a simple glutathione peroxidase mimetic, inhibits human LDL oxidation in vitro. *Atherosclerosis* 201:92-100. 2008.
- de Bem AF, Portella Rde L, Colpo E, Duarte MM, Frediane A, Taube PS, Nogueira CW, Farina M, da Silva EL, Teixeira Rocha JB. Diphenyl diselenide decreases serum levels of total cholesterol

- and tissue oxidative stress in cholesterol-fed rabbits. *Basic Clin Pharmacol Toxicol* 105:17-23. 2009.
- De Haan JB, Crack PJ, Flentjar N, Iannello RC, Hertzog PJ, Kola I. An imbalance in antioxidant defense affects cellular function: the pathophysiological consequences of a reduction in antioxidant defense in the glutathione peroxidase-1 (Gpx1) knockout mouse. *Redox Rep* 8:69-79. 2003.
- de la Torre JC. Alzheimer disease as a vascular disorder: nosological evidence. *Stroke* 33:1152-1162. 2002.
- de la Torre JC. Is Alzheimer's disease a neurodegenerative or a vascular disorder? Data, dogma, and dialectics. *Lancet Neurol* 3:184-190. 2004.
- de la Torre JC. Pathophysiology of neuronal energy crisis in Alzheimer's disease. *Neurodegener Dis* 5:126-132. 2008.
- de Moura MB, dos Santos LS, Van Houten B. Mitochondrial dysfunction in neurodegenerative diseases and cancer. *Environ Mol Mutagen* 51:391-405. 2010.
- Dede DS, Yavuz B, Yavuz BB, Cankurtaran M, Halil M, Ulger Z, Cankurtaran ES, Aytemir K, Kabakci G, Ariogul S. Assessment of endothelial function in Alzheimer's disease: is Alzheimer's disease a vascular disease? *J Am Geriatr Soc* 55:1613-1617. 2007.
- Di Donato S. Disorders related to mitochondrial membranes: pathology of the respiratory chain and neurodegeneration. *J Inherit Metab Dis* 23:247-263. 2000.
- Di Paolo G, Kim TW. Linking lipids to Alzheimer's disease: cholesterol and beyond. *Nat Rev Neurosci* 12:284-296. 2011.
- Dikalov S. Cross talk between mitochondria and NADPH oxidases. *Free Radic Biol Med* 51:1289-1301. 2011.
- DiMauro S, Schon EA. Mitochondrial respiratory-chain diseases. *N Engl J Med* 348:2656-2668. 2003.
- Dirmagl U, Simon RP, Hallenbeck JM. Ischemic tolerance and endogenous neuroprotection. *Trends Neurosci* 26:248-254. 2003.
- Dringen R, Pawlowski PG, Hirrlinger J. Peroxide detoxification by brain cells. *J Neurosci Res* 79:157-165. 2005.
- Dringen R, Hirrlinger J. Glutathione pathways in the brain. *Biol Chem* 384:505-516. 2003.
- Dringen R, Pfeiffer B, Hamprecht B. Synthesis of the antioxidant glutathione in neurons: supply by astrocytes of CysGly as

- precursor for neuronal glutathione. *J Neurosci* 19:562-569. 1999.
- Droge W. Free radicals in the physiological control of cell function. *Physiol Rev* 82:47-95. 2002.
- Drummond GR, Selemidis S, Griendlings KK, Sobey CG. Combating oxidative stress in vascular disease: NADPH oxidases as therapeutic targets. *Nat Rev Drug Discov* 10:453-471. 2011.
- Duchen MR. Roles of mitochondria in health and disease. *Diabetes* 53 Suppl 1:S96-102. 2004.
- Duron E, Hanon O. Vascular risk factors, cognitive decline, and dementia. *Vasc Health Risk Manag* 4:363-381. 2008.
- Eichner JE, Dunn ST, Perveen G, Thompson DM, Stewart KE, Stroehla BC. Apolipoprotein E polymorphism and cardiovascular disease: a HuGE review. *Am J Epidemiol* 155:487-495. 2002.
- Elder GA, Cho JY, English DF, Franciosi S, Schmeidler J, Sosa MA, Gasperi RD, Fisher EA, Mathews PM, Haroutunian V, Buxbaum JD. Elevated plasma cholesterol does not affect brain Abeta in mice lacking the low-density lipoprotein receptor. *J Neurochem* 102:1220-1231. 2007.
- Emerit J, Edeas M, Bricaire F. Neurodegenerative diseases and oxidative stress. *Biomed Pharmacother* 58:39-46. 2004.
- Ennaceur A, Neave N, Aggleton JP. Spontaneous object recognition and object location memory in rats: the effects of lesions in the cingulate cortices, the medial prefrontal cortex, the cingulum bundle and the fornix. *Exp Brain Res* 113:509-519. 1997.
- Espinola-Klein C, Rupprecht HJ, Bickel C, Schnabel R, Genth-Zotz S, Torzewski M, Lackner K, Munzel T, Blankenberg S. Glutathione peroxidase-1 activity, atherosclerotic burden, and cardiovascular prognosis. *Am J Cardiol* 99:808-812. 2007.
- Esposito LA, Kokoszka JE, Waymire KG, Cottrell B, MacGregor GR, Wallace DC. Mitochondrial oxidative stress in mice lacking the glutathione peroxidase-1 gene. *Free Radic Biol Med* 28:754-766. 2000.
- Evola M, Hall A, Wall T, Young A, Grammas P. Oxidative stress impairs learning and memory in apoE knockout mice. *Pharmacol Biochem Behav* 96:181-186. 2010.
- Faivre E, Gault VA, Thorens B, Holscher C. Glucose-dependent insulinotropic polypeptide receptor knockout mice are impaired in learning, synaptic plasticity, and neurogenesis. *J Neurophysiol* 105:1574-1580. 2011.

- Fernandez A, Llacuna L, Fernandez-Checa JC, Colell A. Mitochondrial cholesterol loading exacerbates amyloid beta peptide-induced inflammation and neurotoxicity. *J Neurosci* 29:6394-6405. 2009.
- Ferri CP, Prince M, Brayne C, Brodaty H, Fratiglioni L, Ganguli M, Hall K, Hasegawa K, Hendrie H, Huang Y, Jorm A, Mathers C, Menezes PR, Rimmer E, Scazufca M. Global prevalence of dementia: a Delphi consensus study. *Lancet* 366:2112-2117. 2005.
- Flohe L, Gunzler WA, Schock HH. Glutathione peroxidase: a selenoenzyme. *FEBS Lett* 32:132-134. 1973.
- Fontanesi F, Diaz F, Barrientos A. Evaluation of the mitochondrial respiratory chain and oxidative phosphorylation system using yeast models of OXPHOS deficiencies. Current protocols in human genetics / editorial board, Jonathan L Haines [et al] Chapter 19:Unit19 15. 2009.
- Forgione MA, Weiss N, Heydrick S, Cap A, Klings ES, Bierl C, Eberhardt RT, Farber HW, Loscalzo J. Cellular glutathione peroxidase deficiency and endothelial dysfunction. *Am J Physiol Heart Circ Physiol* 282(4): H1255-61. 2002
- Forstermann U. Nitric oxide and oxidative stress in vascular disease. *Pflugers Arch* 459:923-939. 2010.
- Freeman LR, Haley-Zitlin V, Stevens C, Granholm AC. Diet-induced effects on neuronal and glial elements in the middle-aged rat hippocampus. *Nutr Neurosci* 14:32-44. 2011.
- Ghisleni G, Porciuncula LO, Cimarosti H, Batista TRJ, Salbego CG, Souza DO. Diphenyl diselenide protects rat hippocampal slices submitted to oxygen-glucose deprivation and diminishes inducible nitric oxide synthase immunocontent. *Brain Res* 986:196-199. 2003.
- Gotto AM, Jr., Grundy SM. Lowering LDL cholesterol: questions from recent meta-analyses and subset analyses of clinical trial dataIssues from the interdisciplinary council on reducing the risk for coronary heart disease, ninth council meeting. *Circulation* 99:E1-7. 1999.
- Granholm AC, Bimonte-Nelson HA, Moore AB, Nelson ME, Freeman LR, Sambamurti K. Effects of a saturated fat and high cholesterol diet on memory and hippocampal morphology in the middle-aged rat. *J Alzheimers Dis* 14:133-145. 2008.

- Hafezi-Moghadam A, Thomas KL, Wagner DD. ApoE deficiency leads to a progressive age-dependent blood-brain barrier leakage. *Am J Physiol Cell Physiol* 292:C1256-1262. 2007.
- Halliwell B. Reactive oxygen species and the central nervous system. *J Neurochem* 59:1609-1623. 1992.
- Halliwell B. Role of free radicals in the neurodegenerative diseases: therapeutic implications for antioxidant treatment. *Drugs Aging* 18:685-716. 2001.
- Halliwell B. Oxidative stress and neurodegeneration: where are we now? *J Neurochem* 97:1634-1658. 2006.
- Harish G, Venkateshappa C, Mahadevan A, Pruthi N, Srinivas Bharath MM, Shankar SK. Glutathione metabolism is modulated by postmortem interval, gender difference and agonal state in postmortem human brains. *Neurochem Int* 59:1029-1042. 2011.
- Harrison D, Griendling KK, Landmesser U, Hornig B, Drexler H. Role of oxidative stress in atherosclerosis. *Am J Cardiol* 91:7A-11A. 2003.
- Hensley K, Hall N, Subramaniam R, Cole P, Harris M, Aksenov M, Aksenova M, Gabbita SP, Wu JF, Carney JM, et al. Brain regional correspondence between Alzheimer's disease histopathology and biomarkers of protein oxidation. *J Neurochem* 65:2146-2156. 1995.
- Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS, Johnson AB, Kress Y, Vinters HV, Tabaton M, Shimohama S, Cash AD, Siedlak SL, Harris PL, Jones PK, Petersen RB, Perry G, Smith MA. Mitochondrial abnormalities in Alzheimer's disease. *J Neurosci* 21:3017-3023. 2001.
- Hofman A, Ott A, Breteler MM, Bots ML, Slooter AJ, van Harskamp F, van Duijn CN, Van Broeckhoven C, Grobbee DE. Atherosclerosis, apolipoprotein E, and prevalence of dementia and Alzheimer's disease in the Rotterdam Study. *Lancet* 349:151-154. 1997.
- Hort MA, Straliotto MR, Netto PM, da Rocha JB, de Bem AF, Ribeiro-Valle RM. Diphenyl diselenide effectively reduces atherosclerotic lesions in LDL_r $-/-$ mice by attenuation of oxidative stress and inflammation. *J Cardiovasc Pharmacol* 58:91-101. 2011.
- Hulsmans M, Holvoet P. The vicious circle between oxidative stress and inflammation in atherosclerosis. *J Cell Mol Med* 14:70-78. 2010

- Humpel C. Chronic mild cerebrovascular dysfunction as a cause for Alzheimer's disease? *Exp Gerontol* 46:225-232. 2011
- Ishibashi S, Brown MS, Goldstein JL, Gerard RD, Hammer RE, Herz J. Hypercholesterolemia in low density lipoprotein receptor knockout mice and its reversal by adenovirus-mediated gene delivery. *J Clin Invest* 92:883-893. 1993.
- Jacques-Silva MC, Nogueira CW, Broch LC, Flores EM, Rocha JB. Diphenyl diselenide and ascorbic acid changes deposition of selenium and ascorbic acid in liver and brain of mice. *Pharmacol Toxicol* 88:119-125. 2001.
- Jawien J, Nastalek P, Korbut R. Mouse models of experimental atherosclerosis. *J Physiol Pharmacol* 55:503-517. 2004.
- Kaur H, Halliwell B. Evidence for nitric oxide-mediated oxidative damage in chronic inflammation. Nitrotyrosine in serum and synovial fluid from rheumatoid patients. *FEBS Lett* 350:9-12. 1994.
- Kharrazi H, Vaisi-Raygani A, Rahimi Z, Tavilani H, Aminian M, Pourmotabbed T. Association between enzymatic and non-enzymatic antioxidant defense mechanism with apolipoprotein E genotypes in Alzheimer disease. *Clin Biochem* 41:932-936. 2008.
- Khatri JJ, Johnson C, Magid R, Lessner SM, Laude KM, Dikalov SI, Harrison DG, Sung HJ, Rong Y, Galis ZS. Vascular oxidant stress enhances progression and angiogenesis of experimental atheroma. *Circulation* 109:520-525. 2004.
- Kim SJ, Park C, Han AL, Youn MJ, Lee JH, Kim Y, Kim ES, Kim HJ, Kim JK, Lee HK, Chung SY, So H, Park R. Ebselen attenuates cisplatin-induced ROS generation through Nrf2 activation in auditory cells. *Hear Res* 251:70-82. 2009.
- Kivipelto M, Ngandu T, Fratiglioni L, Viitanen M, Kareholt I, Winblad B, Helkala EL, Tuomilehto J, Soininen H, Nissinen A. Obesity and vascular risk factors at midlife and the risk of dementia and Alzheimer disease. *Arch Neurol* 62:1556-1560. 2005.
- Kivipelto M, Helkala EL, Laakso MP, Hanninen T, Hallikainen M, Alhainen K, Iivonen S, Mannermaa A, Tuomilehto J, Nissinen A, Soininen H. Apolipoprotein E epsilon4 allele, elevated midlife total cholesterol level, and high midlife systolic blood pressure are independent risk factors for late-life Alzheimer disease. *Ann Intern Med* 137:149-155. 2002.
- Kivipelto M, Helkala EL, Laakso MP, Hanninen T, Hallikainen M, Alhainen K, Soininen H, Tuomilehto J, Nissinen A. Midlife

- vascular risk factors and Alzheimer's disease in later life: longitudinal, population based study. *BMJ* 322:1447-1451. 2001.
- Klotz LO, Sies H. Defenses against peroxy nitrite: selenocompounds and flavonoids. *Toxicol Lett* 140-141:125-132. 2003.
- Kolovou G, Yiannakouris N, Hatzivassiliou M, Malakos J, Daskalova D, Hatzigeorgiou G, Cariolou MA, Cokkinos DV. Association of apolipoprotein E polymorphism with myocardial infarction in Greek patients with coronary artery disease. *Curr Med Res Opin* 18:118-124. 2002.
- Kowala MC, Recce R, Beyer S, Gu C, Valentine M. Characterization of atherosclerosis in LDL receptor knockout mice: macrophage accumulation correlates with rapid and sustained expression of aortic MCP-1/JE. *Atherosclerosis* 149:323-330. 2000.
- Kowaltowski AJ, Castilho RF, Vercesi AE. Mitochondrial permeability transition and oxidative stress. *FEBS Lett* 495:12-15. 2001.
- Kowaltowski AJ, Vercesi AE. Mitochondrial damage induced by conditions of oxidative stress. *Free Radic Biol Med* 26:463-471. 1999.
- Kryukov GV, Castellano S, Novoselov SV, Lobanov AV, Zehtab O, Guigo R, Gladyshev VN. Characterization of mammalian selenoproteomes. *Science* 300:1439-1443. 2003.
- Lapenna D, de Gioia S, Ciofani G, Mezzetti A, Ucchino S, Calafiore AM, Napolitano AM, Di Ilio C, Cuccurullo F. Glutathione-related antioxidant defenses in human atherosclerotic plaques. *Circulation* 97:1930-1934. 1998.
- Lash LH. Mitochondrial glutathione transport: physiological, pathological and toxicological implications. *Chem Biol Interact* 163:54-67. 2006.
- LeDoux SP, Driggers WJ, Hollensworth BS, Wilson GL. Repair of alkylation and oxidative damage in mitochondrial DNA. *Mutat Res* 434:149-159. 1999.
- Lehninger A, Nelson D, Cox MM. Oxidative phosphorylation and photophosphorylation. In: *Principles of Biochemistry* (Lehninger, A. et al., eds), 690 – 750 New York: W.H. Freeman. 2004.
- Leonard JV, Schapira AH. Mitochondrial respiratory chain disorders I: mitochondrial DNA defects. *Lancet* 355:299-304. 2000.
- Libby P. Inflammation in atherosclerosis. *Nature* 420:868-874. 2002.
- Lin MT, Beal MF. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 443:787-795. 2006.

- Liscum L, Underwood KW. Intracellular cholesterol transport and compartmentation. *J Biol Chem* 270:15443-15446. 1995.
- Lovell MA, Ehmann WD, Butler SM, Markesberry WR. Elevated thiobarbituric acid-reactive substances and antioxidant enzyme activity in the brain in Alzheimer's disease. *Neurology* 45:1594-1601. 1995.
- Luthra K, Bhargav B, Chabria S, Das N, Misra A, Agarwal DP, Pandey RM, Srivastava LM. Apolipoprotein E polymorphism in Northern Indian patients with coronary heart disease: phenotype distribution and relation to serum lipids and lipoproteins. *Mol Cell Biochem* 232:97-102. 2002.
- Maciel EN, Flores EM, Rocha JB, Folmer V. Comparative deposition of diphenyl diselenide in liver, kidney, and brain of mice. *Bull Environ Contam Toxicol* 70:470-476. 2003.
- Maher P. The effects of stress and aging on glutathione metabolism. *Ageing Res Rev* 4:288-314. 2005.
- Mansego ML, Redon J, Martinez-Hervas S, Real JT, Martinez F, Blesa S, Gonzalez-Albert V, Saez GT, Carmena R, Chaves FJ. Different impacts of cardiovascular risk factors on oxidative stress. *Int J Mol Sci* 12:6146-6163. 2011.
- Mao P, Reddy PH. Aging and amyloid beta-induced oxidative DNA damage and mitochondrial dysfunction in Alzheimer's disease: implications for early intervention and therapeutics. *Biochim Biophys Acta* 1812:1359-1370. 2011.
- Mari M, Colell A, Morales A, Caballero F, Moles A, Fernandez A, Terrones O, Basanez G, Antonsson B, Garcia-Ruiz C, Fernandez-Checa JC. Mechanism of mitochondrial glutathione-dependent hepatocellular susceptibility to TNF despite NF-kappaB activation. *Gastroenterology* 134:1507-1520. 2008.
- McCommis KS, McGee AM, Laughlin MH, Bowles DK, Baines CP. Hypercholesterolemia increases mitochondrial oxidative stress and enhances the MPT response in the porcine myocardium: beneficial effects of chronic exercise. *Am J Physiol Regul Integr Comp Physiol* 301:R1250-1258. 2011.
- Meotti FC, Borges VC, Zeni G, Rocha JB, Nogueira CW. Potential renal and hepatic toxicity of diphenyl diselenide, diphenyl ditelluride and Ebselen for rats and mice. *Toxicol Lett* 143:9-16. 2003.
- Meotti FC, Stangherlin EC, Zeni G, Nogueira CW, Rocha JB. Protective role of aryl and alkyl diselenides on lipid peroxidation. *Environ Res* 94:276-282. 2004.

- Mielke MM, Zandi PP, Sjogren M, Gustafson D, Ostling S, Steen B, Skoog I. High total cholesterol levels in late life associated with a reduced risk of dementia. *Neurology* 64:1689-1695. 2005.
- Montilla P, Espejo I, Munoz MC, Bujalance I, Munoz-Castaneda JR, Tunez I. Protective effect of red wine on oxidative stress and antioxidant enzyme activities in the brain and kidney induced by feeding high cholesterol in rats. *Clin Nutr* 25:146-153. 2006.
- Moreira PI, Carvalho C, Zhu X, Smith MA, Perry G. Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. *Biochim Biophys Acta* 1802:2-10. 2010.
- Moreira PI, Zhu X, Wang X, Lee HG, Nunomura A, Petersen RB, Perry G, Smith MA. Mitochondria: a therapeutic target in neurodegeneration. *Biochim Biophys Acta* 1802:212-220.2010.
- Morgan MJ, Kim YS, Liu Z. Lipid rafts and oxidative stress-induced cell death. *Antioxid Redox Signal* 9:1471-1483. 2007.
- Mugesh G, Singh H. Synthetic organoselenium compounds as antioxidants: glutathione peroxidase activity. *Chem Soc Rev* 29:347-357. 2000.
- Mulder M, Jansen PJ, Janssen BJ, van de Berg WD, van der Boom H, Havekes LM, de Kloet RE, Ramaekers FC, Blokland A. Low-density lipoprotein receptor-knockout mice display impaired spatial memory associated with a decreased synaptic density in the hippocampus. *Neurobiol Dis* 16:212-219. 2004.
- Muller A, Cadenas E, Graf P, Sies H. A novel biologically active seleno-organic compound--I. Glutathione peroxidase-like activity in vitro and antioxidant capacity of PZ 51 (Ebselen). *Biochem Pharmacol* 33:3235-3239. 1984.
- Murai T, Okuda S, Tanaka T, Ohta H. Characteristics of object location memory in mice: Behavioral and pharmacological studies. *Physiol Behav* 90:116-124. 2007.
- Nelson AJ, Cooper MT, Thur KE, Marsden CA, Cassaday HJ. The effect of catecholaminergic depletion within the prelimbic and infralimbic medial prefrontal cortex on recognition memory for recency, location, and objects. *Behav Neurosci* 125:396-403. 2011.
- Niki E. Lipid peroxidation: physiological levels and dual biological effects. *Free Radic Biol Med* 47:469-484. 2009.
- Nogueira CW, Rocha JB. Toxicology and pharmacology of selenium: emphasis on synthetic organoselenium compounds. *Arch Toxicol* 85:1313-1359. 2011.

- Nogueira CW, Rocha JB. Diphenyl Diselenide a Janus-Faced Molecule. *J Braz Chem Soc* 2010.
- Nogueira CW, Zeni G, Rocha JB. Organoselenium and organotellurium compounds: toxicology and pharmacology. *Chem Rev* 104:6255-6285. 2004.
- Nogueira CW, Borges VC, Zeni G, Rocha JB. Organochalcogens effects on delta-aminolevulinate dehydratase activity from human erythrocytic cells in vitro. *Toxicology* 191:169-178. 2003a.
- Nogueira CW, Quinhones EB, Jung EA, Zeni G, Rocha JB. Anti-inflammatory and antinociceptive activity of diphenyl diselenide. *Inflamm Res* 52:56-63. 2003b.
- O'Donovan DJ, Fernandes CJ. Mitochondrial glutathione and oxidative stress: implications for pulmonary oxygen toxicity in premature infants. *Mol Genet Metab* 71:352-358. 2000.
- Ohashi R, Mu H, Yao Q, Chen C. Cellular and molecular mechanisms of atherosclerosis with mouse models. *Trends Cardiovasc Med* 14:187-190. 2004.
- Oliveira HC, Cocco RG, Alberici LC, Maciel EN, Salerno AG, Dorighelli GG, Velho JA, de Faria EC, Vercesi AE. Oxidative stress in atherosclerosis-prone mouse is due to low antioxidant capacity of mitochondria. *FASEB J* 19:278-280. 2005.
- Paigen B, Holmes PA, Novak EK, Swank RT. Analysis of atherosclerosis susceptibility in mice with genetic defects in platelet function. *Arteriosclerosis* 10:648-652. 1990.
- Paim BA, Velho JA, Castilho RF, Oliveira HC, Vercesi AE. Oxidative stress in hypercholesterolemic LDL (low-density lipoprotein) receptor knockout mice is associated with low content of mitochondrial NADP-linked substrates and is partially reversed by citrate replacement. *Free Radic Biol Med* 44:444-451. 2008.
- Panza F, D'Introno A, Colacicco AM, Capurso C, Pichichero G, Capurso SA, Capurso A, Solfrizzi V. Lipid metabolism in cognitive decline and dementia. *Brain Res Rev* 51:275-292. 2006.
- Pappolla MA, Smith MA, Bryant-Thomas T, Bazan N, Petanceska S, Perry G, Thal LJ, Sano M, Refolo LM. Cholesterol, oxidative stress, and Alzheimer's disease: expanding the horizons of pathogenesis. *Free Radic Biol Med* 33:173-181. 2002.
- Parnham MJ, Kindt S. A novel biologically active seleno-organic compound-III. Effects of PZ 51 (Ebselen) on glutathione peroxidase and secretory activities of mouse macrophages. *Biochem Pharmacol* 33:3247-3250. 1984.

- Pfrieger FW. Cholesterol homeostasis and function in neurons of the central nervous system. *Cell Mol Life Sci* 60:1158-1171. 2003.
- Pieczenik SR, Neustadt J. Mitochondrial dysfunction and molecular pathways of disease. *Exp Mol Pathol* 83:84-92. 2007.
- Porciuncula LO, Rocha JB, Boeck CR, Vendite D, Souza DO. Ebselen prevents excitotoxicity provoked by glutamate in rat cerebellar granule neurons. *Neurosci Lett* 299:217-220. 2001.
- Posser T, Franco JL, dos Santos DA, Rigon AP, Farina M, Dafre AL, Teixeira Rocha JB, Leal RB. Diphenyl diselenide confers neuroprotection against hydrogen peroxide toxicity in hippocampal slices. *Brain Res* 1199:138-147. 2008.
- Raes M, Michiels C, Remacle J. Comparative study of the enzymatic defense systems against oxygen-derived free radicals: the key role of glutathione peroxidase. *Free Radic Biol Med* 3:3-7. 1987.
- Ramirez C, Sierra S, Tercero I, Vazquez JA, Pineda A, Manrique T, Burgos JS. ApoB100/LDLR-/- hypercholesterolaemic mice as a model for mild cognitive impairment and neuronal damage. *PLoS One* 6:e22712. 2011.
- Ran Q, Gu M, Van Remmen H, Strong R, Roberts JL, Richardson A. Glutathione peroxidase 4 protects cortical neurons from oxidative injury and amyloid toxicity. *J Neurosci Res* 84:202-208. 2006.
- Rapp JH, Pan XM, Neumann M, Hong M, Hollenbeck K, Liu J. Microemboli composed of cholesterol crystals disrupt the blood-brain barrier and reduce cognition. *Stroke* 39:2354-2361. 2008.
- Ray R, Shah AM. NADPH oxidase and endothelial cell function. *Clin Sci (Lond)* 109:217-226. 2005.
- Refolo LM, Malester B, LaFrancois J, Bryant-Thomas T, Wang R, Tint GS, Sambamurti K, Duff K, Pappolla MA. Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. *Neurobiol Dis* 7:321-331. 2000.
- Rhee SG, Yang KS, Kang SW, Woo HA, Chang TS. Controlled elimination of intracellular H₂O₂: regulation of peroxiredoxin, catalase, and glutathione peroxidase via post-translational modification. *Antioxid Redox Signal* 7:619-626. 2005.

- Richter C, Park JW, Ames BN. Normal oxidative damage to mitochondrial and nuclear DNA is extensive. *Proc Natl Acad Sci U S A* 85:6465-6467. 1988.
- Rosa RM, Flores DG, Appelt HR, Braga AL, Henriques JA, Roesler R. Facilitation of long-term object recognition memory by pretraining administration of diphenyl diselenide in mice. *Neurosci Lett* 341:217-220. 2003.
- Ross R, Harker L. Hyperlipidemia and atherosclerosis. *Science* 193:1094-1100. 1976.
- Rousset S, Alves-Guerra MC, Mozo J, Miroux B, Cassard-Doulcier AM, Bouillaud F, Ricquier D. The biology of mitochondrial uncoupling proteins. *Diabetes* 53 Suppl 1:S130-135. 2004.
- Rubbo H, Radi R, Trujillo M, Telleri R, Kalyanaraman B, Barnes S, Kirk M, Freeman BA. Nitric oxide regulation of superoxide and peroxynitrite-dependent lipid peroxidation. Formation of novel nitrogen-containing oxidized lipid derivatives. *J Biol Chem* 269:26066-26075. 1994.
- Saito I, Asano T, Sano K, Takakura K, Abe H, Yoshimoto T, Kikuchi H, Ohta T, Ishibashi S. Neuroprotective effect of an antioxidant, ebselen, in patients with delayed neurological deficits after aneurysmal subarachnoid hemorrhage. *Neurosurgery* 42:269-277; discussion 277-268. 1998.
- Saraste M. Structural features of cytochrome oxidase. *Q Rev Biophys* 23:331-366. 1990.
- Schulz JB, Matthews RT, Klockgether T, Dichgans J, Beal MF. The role of mitochondrial dysfunction and neuronal nitric oxide in animal models of neurodegenerative diseases. *Mol Cell Biochem* 174:193-197. 1997.
- Setia N, Verma IC, Khan B, Arora A. Premature coronary artery disease and familial hypercholesterolemia: need for early diagnosis and cascade screening in the Indian population. *Cardiol Res Pract* 2012:658526. 2012.
- Shobab LA, Hsiung GY, Feldman HH. Cholesterol in Alzheimer's disease. *Lancet Neurol* 4:841-852. 2005.
- Sies H. Oxidative stress: oxidants and antioxidants. *Exp Physiol* 82:291-295. 1997.
- Sies H, Arteel GE. Interaction of peroxynitrite with selenoproteins and glutathione peroxidase mimics. *Free Radic Biol Med* 28:1451-1455. 2000.
- Simons K, Ikonen E. How cells handle cholesterol. *Science* 290:1721-1726. 2000.

- Sparks DL, Sabbagh MN, Connor DJ, Lopez J, Launer LJ, Browne P, Wasser D, Johnson-Traver S, Lochhead J, Ziolkowski C. Atorvastatin for the treatment of mild to moderate Alzheimer disease: preliminary results. *Arch Neurol* 62:753-757. 2005.
- Sparks DL, Scheff SW, Hunsaker JC, 3rd, Liu H, Landers T, Gross DR. Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. *Exp Neurol* 126:88-94. 1994.
- Spees JL, Olson SD, Whitney MJ, Prockop DJ. Mitochondrial transfer between cells can rescue aerobic respiration. *Proc Natl Acad Sci U S A* 103:1283-1288. 2006.
- Stadtman ER, Levine RL. Protein oxidation. *Ann N Y Acad Sci* 899:191-208. 2000.
- Steinberg D. Atherogenesis in perspective: hypercholesterolemia and inflammation as partners in crime. *Nat Med* 8:1211-1217. 2002.
- Stokes J, 3rd, Kannel WB, Wolf PA, Cupples LA, D'Agostino RB. The relative importance of selected risk factors for various manifestations of cardiovascular disease among men and women from 35 to 64 years old: 30 years of follow-up in the Framingham Study. *Circulation* 75:V65-73. 1987.
- Straliotto MR, Mancini G, de Oliveira J, Nazari EM, Muller YM, Dafre A, Ortiz S, Silva EL, Farina M, Latini A, Rocha JB, de Bem AF. Acute exposure of rabbits to diphenyl diselenide: a toxicological evaluation. *J Appl Toxicol* 30:761-768. 2010.
- Sultana R, Butterfield DA. Identification of the oxidative stress proteome in the brain. *Free Radic Biol Med* 50:487-494. 2011.
- Tamasi V, Jeffries JM, Arteel GE, Falkner KC. Ebselen augments its peroxidase activity by inducing nrf-2-dependent transcription. *Arch Biochem Biophys* 431:161-168. 2004.
- Tan S, Sagara Y, Liu Y, Maher P, Schubert D. The regulation of reactive oxygen species production during programmed cell death. *J Cell Biol* 141:1423-1432. 1998.
- Thirumangalakudi L, Prakasam A, Zhang R, Bimonte-Nelson H, Sambamurti K, Kindy MS, Bhat NR. High cholesterol-induced neuroinflammation and amyloid precursor protein processing correlate with loss of working memory in mice. *J Neurochem* 106:475-485. 2008.
- Torzewski M, Ochsenhirt V, Kleschyov AL, Oelze M, Daiber A, Li H, Rossmann H, Tsimikas S, Reifenberg K, Cheng F, Lehr HA, Blankenberg S, Forstermann U, Munzel T, Lackner KJ. Deficiency of glutathione peroxidase-1 accelerates the

- progression of atherosclerosis in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* 27:850-857. 2007.
- Turrens JF. Mitochondrial formation of reactive oxygen species. *J Physiol* 552:335-344. 2003.
- Ullrich C, Pirchl M, Humpel C. Hypercholesterolemia in rats impairs the cholinergic system and leads to memory deficits. *Mol Cell Neurosci* 45:408-417. 2010.
- Ursini F, Heim S, Kiess M, Maiorino M, Roveri A, Wissing J, Flohe L. Dual function of the selenoprotein PHGPx during sperm maturation. *Science* 285:1393-1396. 1999.
- Vercesi AE, Castilho RF, Kowaltowski AJ, Oliveira HC. Mitochondrial energy metabolism and redox state in dyslipidemias. *IUBMB Life* 59:263-268. 2007.
- Vindis C, Elbaz M, Escargueil-Blanc I, Auge N, Heniquez A, Thiers JC, Negre-Salvayre A, Salvayre R. Two distinct calcium-dependent mitochondrial pathways are involved in oxidized LDL-induced apoptosis. *Arterioscler Thromb Vasc Biol* 25:639-645. 2005.
- Wallace DC. Mitochondrial diseases in man and mouse. *Science* 283:1482-1488. 1999.
- Wilson SR, Zucker PA, Huang RRC, Spector A. Development of synthetic compounds with glutathione peroxidase activity. *J Am Chem Soc* 111: 5936–5939. 1989.
- Whitehouse PJ, Sciulli CG, Mason RM. Dementia drug development: use of information systems to harmonize global drug development. *Psychopharmacol Bull* 33:129-133. 1997.
- Whitmer RA, Sidney S, Selby J, Johnston SC, Yaffe K. Midlife cardiovascular risk factors and risk of dementia in late life. *Neurology* 64:277-281. 2005.
- Witte ME, Geurts JJ, de Vries HE, van der Valk P, van Horssen J. Mitochondrial dysfunction: a potential link between neuroinflammation and neurodegeneration? *Mitochondrion* 10:411-418. 2010.
- Wolozin B. Cholesterol, statins and dementia. *Curr Opin Lipidol* 15:667-672. 2004.
- Yamagata K, Ichinose S, Miyashita A, Tagami M. Protective effects of ebselen, a seleno-organic antioxidant on neurodegeneration induced by hypoxia and reperfusion in stroke-prone spontaneously hypertensive rat. *Neuroscience* 153:428-435. 2008.
- Yin Z, Lee E, Ni M, Jiang H, Milatovic D, Rongzhu L, Farina M, Rocha JB, Aschner M. Methylmercury-induced alterations in astrocyte

- functions are attenuated by ebselen. *Neurotoxicology* 32:291-299. 2011.
- Zadelaar S, Kleemann R, Verschuren L, de Vries-Van der Weij J, van der Hoorn J, Princen HM, Kooistra T. Mouse models for atherosclerosis and pharmaceutical modifiers. *Arterioscler Thromb Vasc Biol* 27:1706-1721. 2007.
- Zambon D, Quintana M, Mata P, Alonso R, Benavent J, Cruz-Sanchez F, Gich J, Pocovi M, Civeira F, Capurro S, Bachman D, Sambamurti K, Nicholas J, Pappolla MA. Higher incidence of mild cognitive impairment in familial hypercholesterolemia. *Am J Med* 123:267-274. 2010.
- Zhao R, Holmgren A. A novel antioxidant mechanism of ebselen involving ebselen diselenide, a substrate of mammalian thioredoxin and thioredoxin reductase. *J Biol Chem* 277:39456-39462. 2002.
- Zmijewski JW, Moellering DR, Le Goffe C, Landar A, Ramachandran A, Darley-Usmar VM. Oxidized LDL induces mitochondrially associated reactive oxygen/nitrogen species formation in endothelial cells. *Am J Physiol Heart Circ Physiol* 289:H852-861. 2005.